

Synthesis of Branched Monodisperse Oligoethylene Glycols and ^{19}F MRI-Traceable Biomaterials through Reductive Dimerization of Azides

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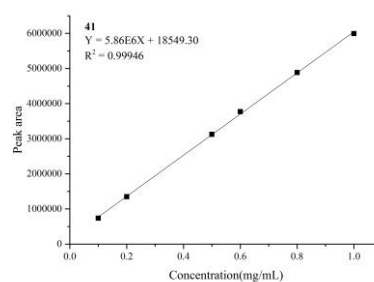
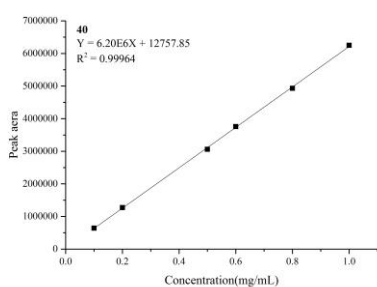
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1. General information

^1H , ^{19}F and ^{13}C NMR spectra were recorded on a Bruker 400 MHz. Chemical shifts are in ppm and coupling constants (J) are in Hertz (Hz). ^1H NMR spectra were referenced to tetramethylsilane (d, 0.00 ppm) using CDCl_3 as solvent, ^{13}C NMR spectra were referenced to solvent carbons (77.16 ppm for CDCl_3). ^{19}F NMR spectra were referenced to 2% perfluorobenzene (s, -164.90 ppm) in CDCl_3 . The splitting patterns for ^1H NMR spectra are denoted as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), b (broad) and combinations thereof. MS spectra were recorded on an Asilent LCMS-1100 spectrometer. MALDI-TOF mass spectra were recorded on a MALDI-TOF/TOF 5800 (AB SCIEX) spectrometer using the reflector mode for positive ions with α -cyano-4-hydroxycinnamic acid as matrix.

2. *n*-Octanol/Water partition coefficients (logP) measurement

The logP values of compounds **40-41** were measured following shake-flask method. Briefly, the product was dissolved in distilled *n*-octanol saturated with water. Then 1 mL of this solution was mixed with an equal volume of water saturated with distilled *n*-octanol and mixed on a vortex device. After shaking the mixture overnight, water phase was separated by centrifugation. Equal-volume samples of the shaken water phase and the starting solution were subsequently taken and analyzed by HPLC. The peak area was measured at $\lambda = 254$ nm, and compared with calibration curve to obtain the concentration of the peptide. LogP values were determined from: $\text{Lg}[(C_s - C_w)/C_w]$, where C_s and C_w are the concentrations of the starting water solution and the water phase of the compound, respectively.



3. Solvent-dependent ^{19}F NMR of 40 and 41

Solvent-dependent ^{19}F NMR spectra were referenced to 10% sodium trifluoromethanesulfonate (s, -79.61 ppm) in D_2O at 25 °C.

4. Dynamic light scattering of 41

Solution of **41** in H₂O at 0.07 mM was used for DLS analysis. The particle size was measured at an angle of 90° in a 10 mm diameter cell at the room temperature with a Dynamic Light Scattering (DLS) Analyzer (Malvern ZetasizerNano 3690). Eleven scans were run for each measurement and the measurement was repeated 3 times. The particle size and polydispersity index (PDI) were calculated by Malvern software.

5. In vitro ¹⁹F MRI experiments of 40 and 41

All magnetic resonance imaging (MRI) experiments were performed on a 400 MHz Bruker BioSpec MRI system. The temperature of the magnet room was maintained at 25 °C during the entire MRI experiment. The ¹⁹F in vitro images were acquired using a gradient-echo (GRE) pulse sequence, method = RARE, matrix size = 32 × 32, SI = 20 mm, FOV = 3.0 cm, TR = 2500 ms, TE = 2.8 ms, scan time = 160 s.

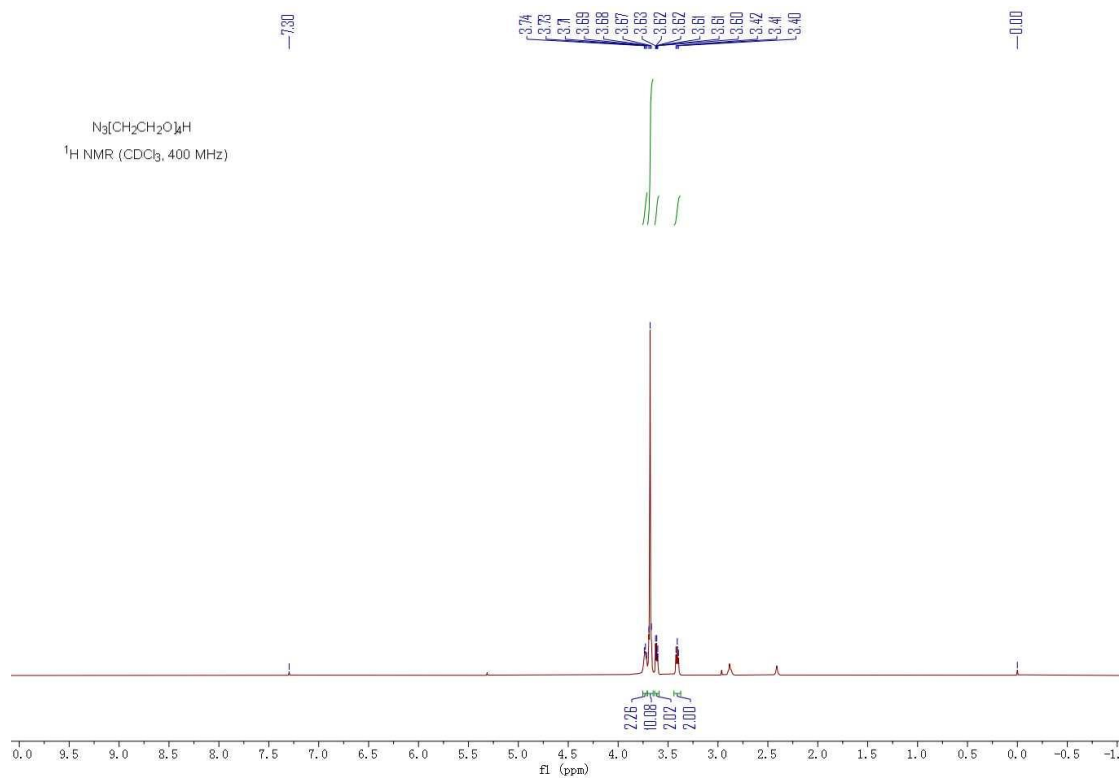
6. Cytotoxicity assay of 40 and 41

HepG2 cells were cultured in DMEM medium containing 10% FBS and 1% streptomycin double antibody. L929 cells were cultured in alpha-MEM medium containing 10% FBS and 1% streptomycin double antibody. All cells were cultured at 37 °C in humidified atmosphere containing 5% CO₂ and the growth medium was replaced with fresh media every 24 h.

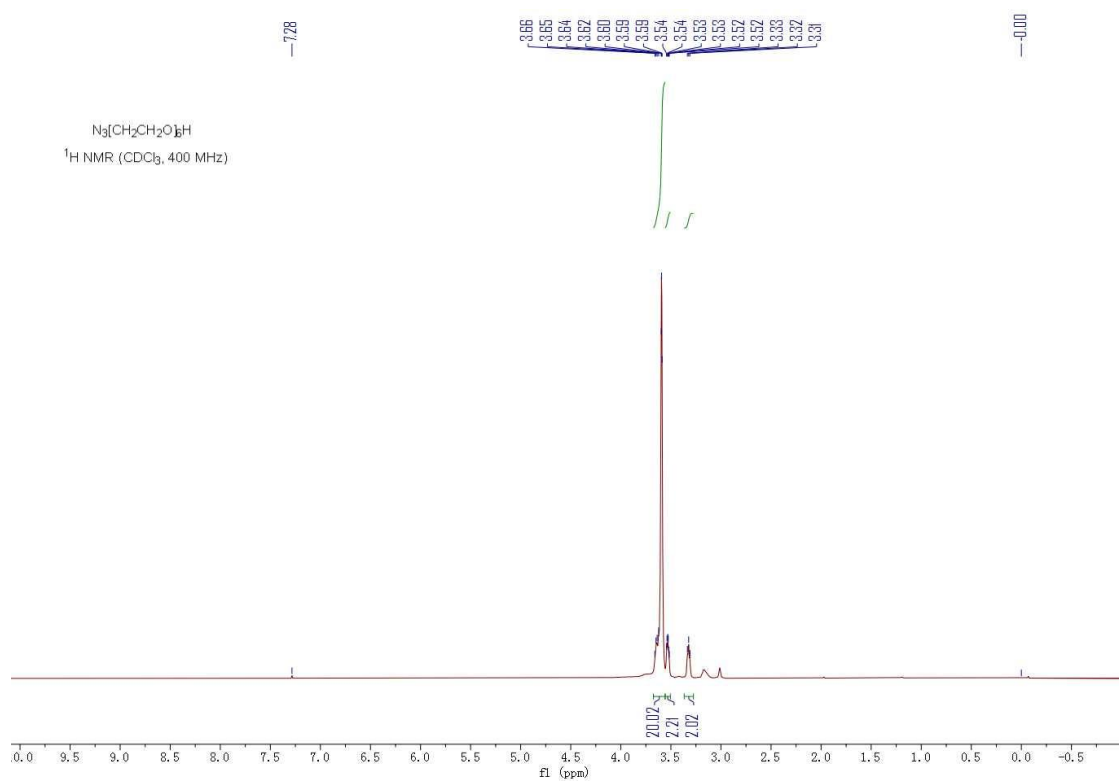
The cell viability assay of compounds **40** and **41** were investigated in L929 cell lines and HepG2 cell lines *in vitro* by MTT assay. L929 cells HepG2 cells were seeded into a 96-well plate for several hours. Subsequently, a gradient concentration of the compounds ranging from 62 µg/mL to 1000 µg/mL were added in a series of wells. Every concentration was set with five wells at least. The wells with 100 µL culture medium alone were used as negative control and wells containing cells alone were used as positive control. After incubation for 24 h, the medium was replaced with 100 µL MTT (1.0 mg/mL) solution and incubated for 4 h. Then the medium was replaced with 200 µL DMSO and the absorbance value was measured at 490 nm using a microplate reader (Bio Tek Instruments, USA).

7. Copies of $^1\text{H}/^{13}\text{C}/^{19}\text{F}$ NMR, MS and mass spectra (HRMS) of compounds

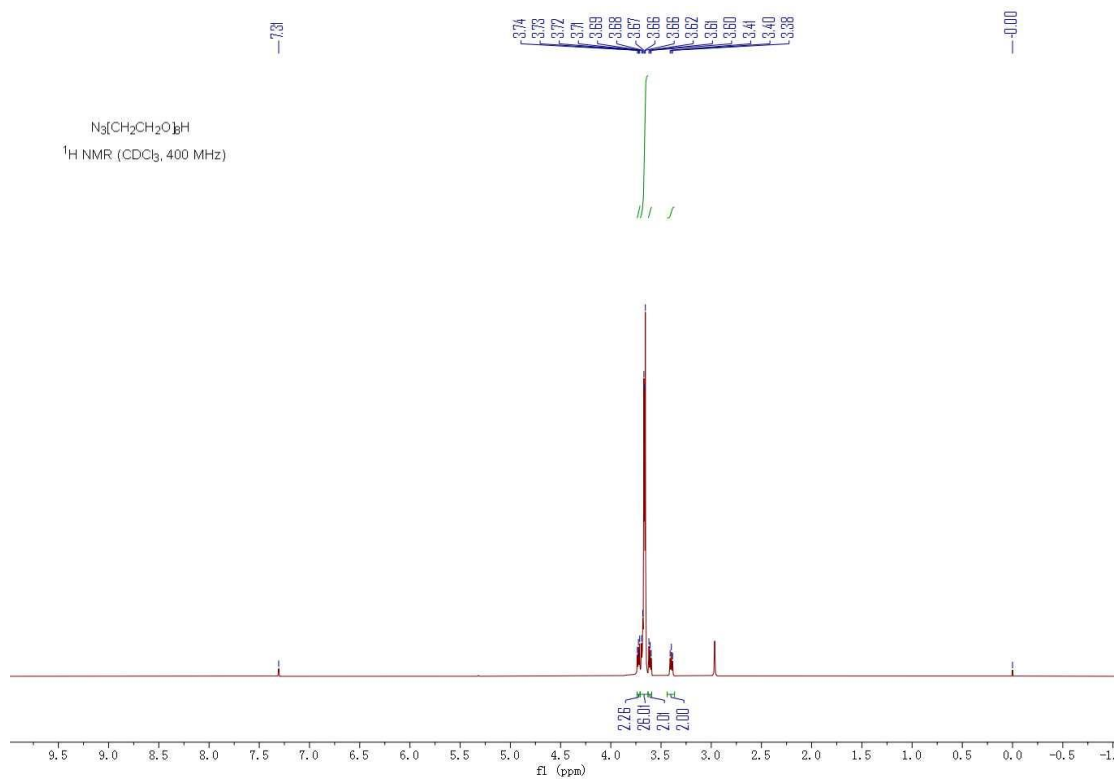
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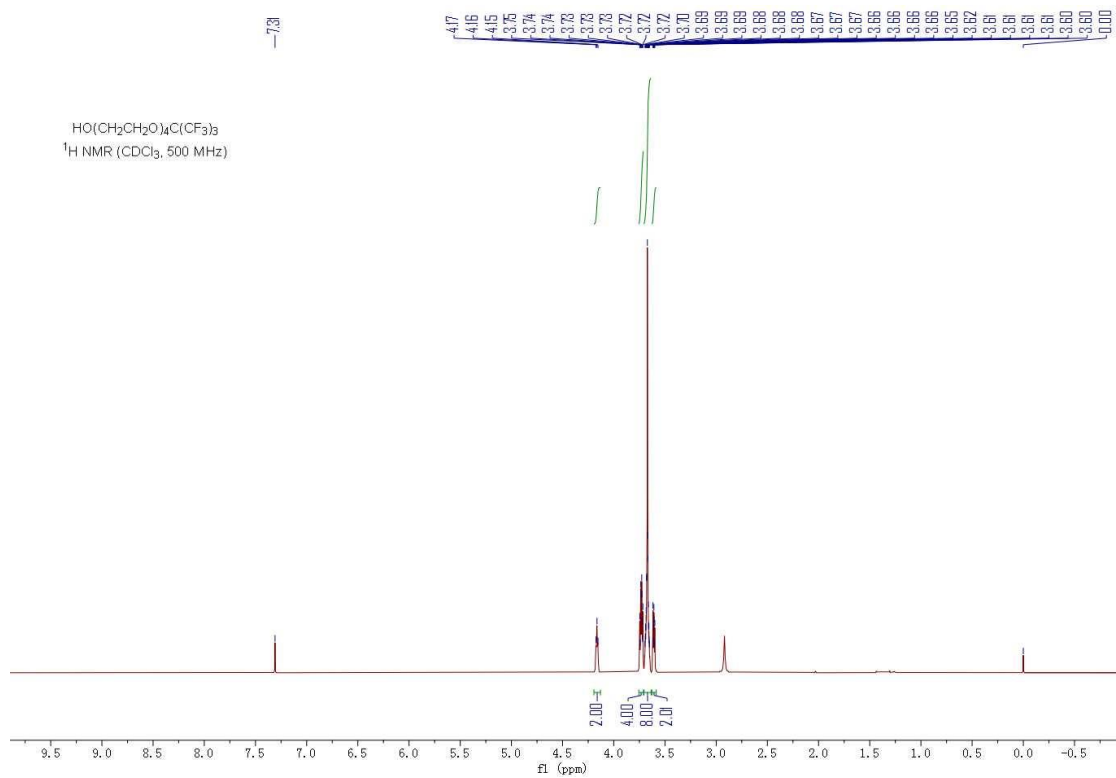
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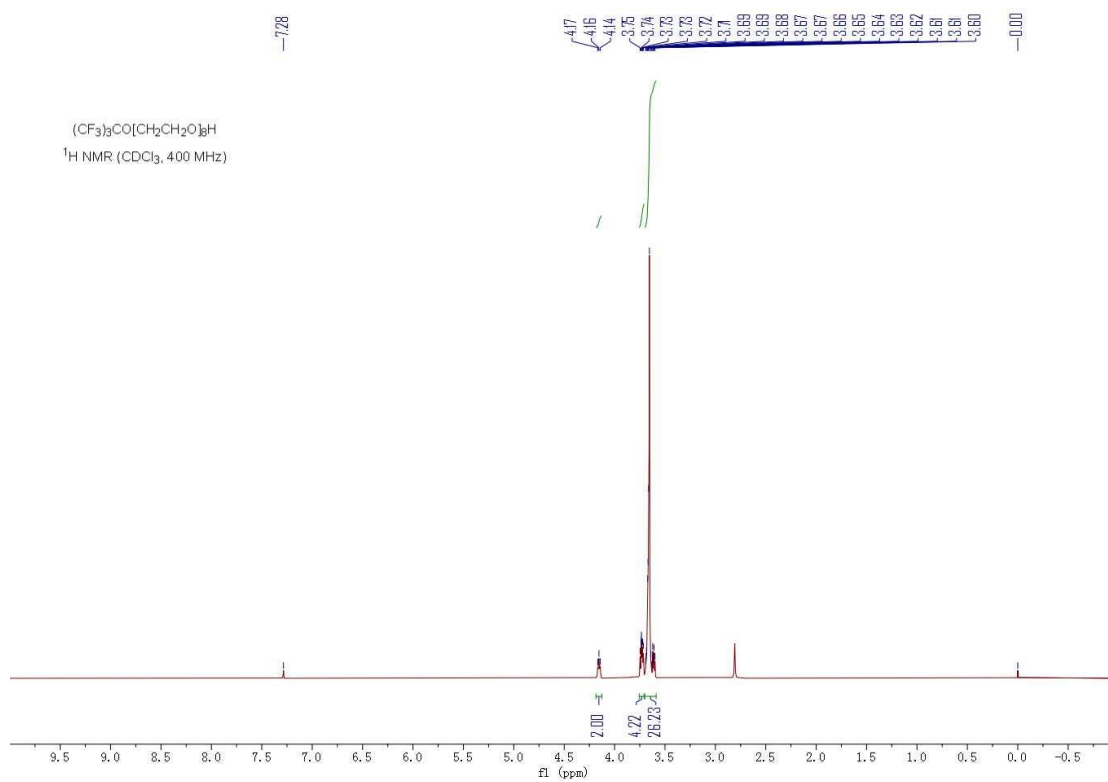
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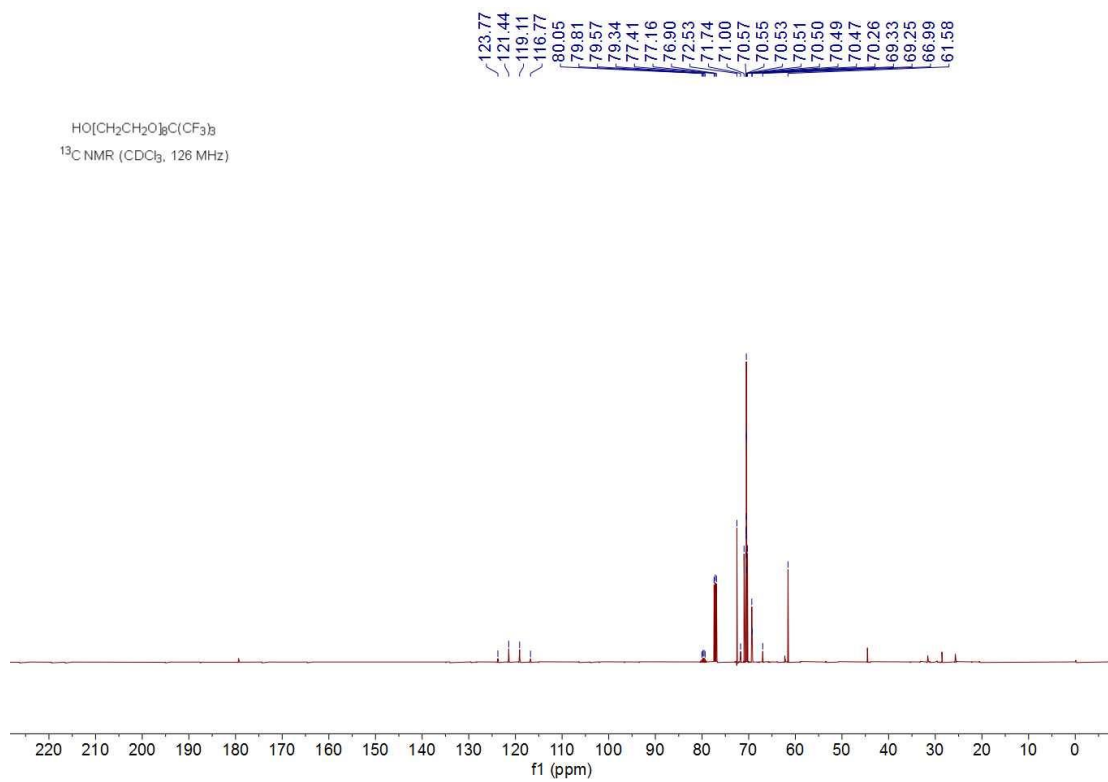
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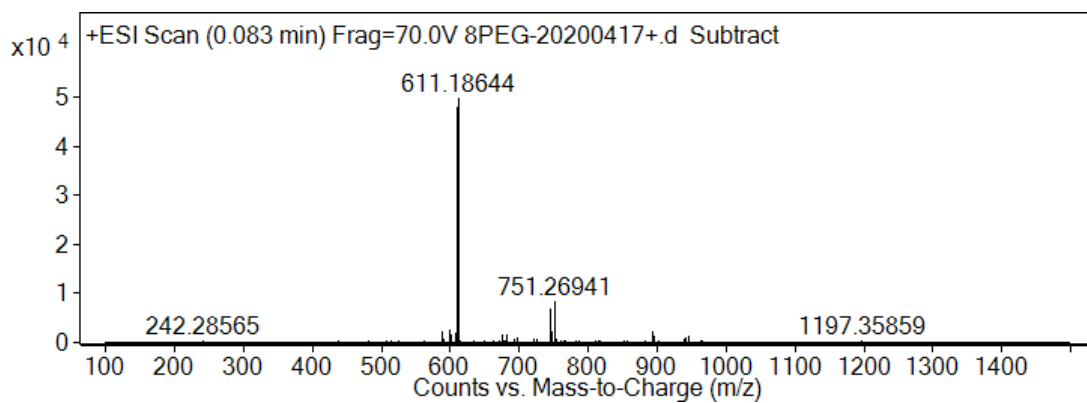
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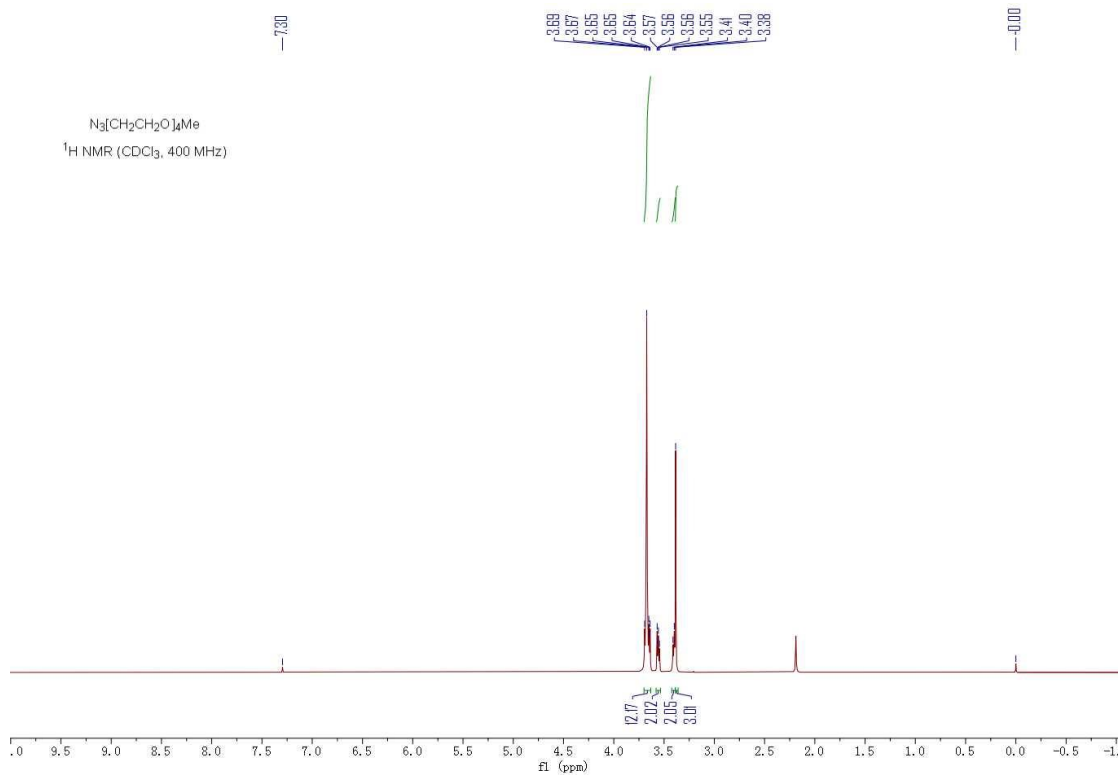
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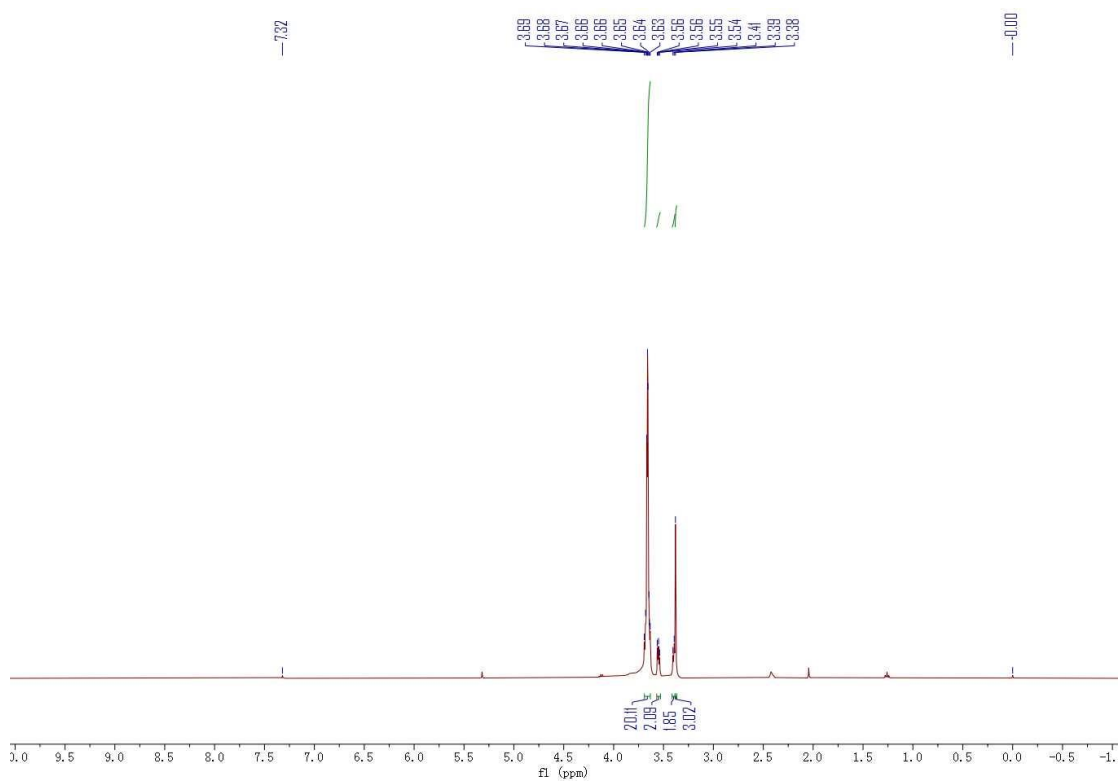
HRMS of compound 6



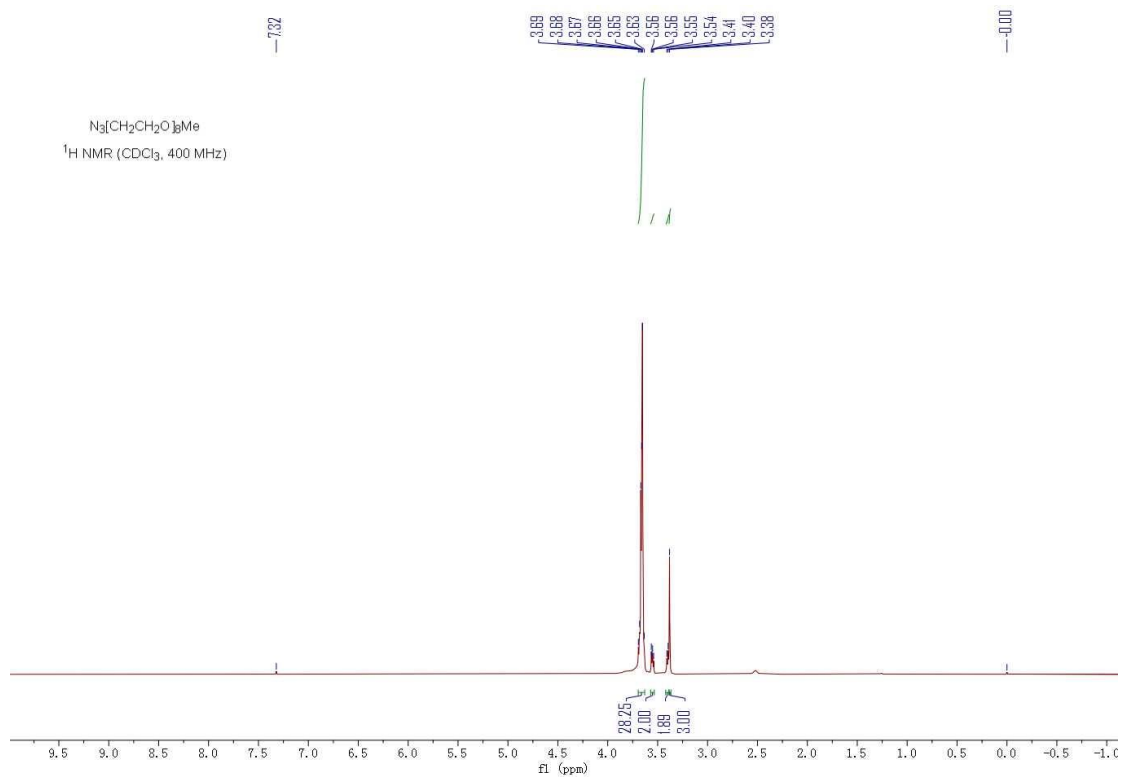
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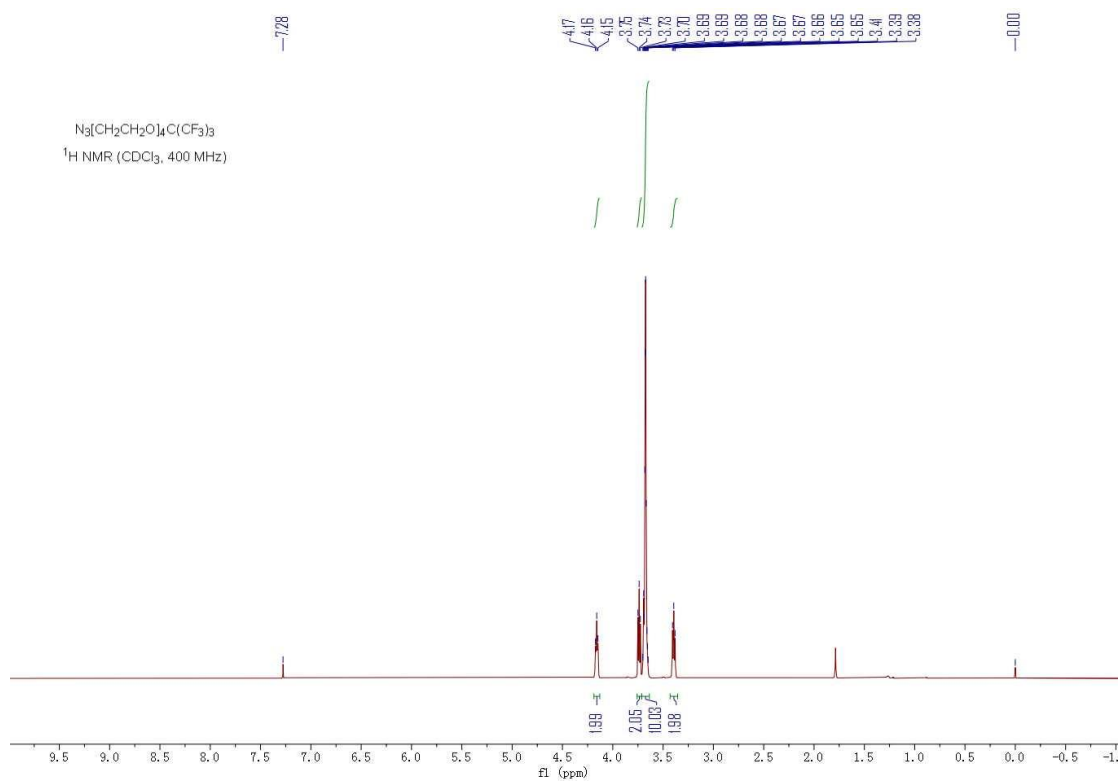
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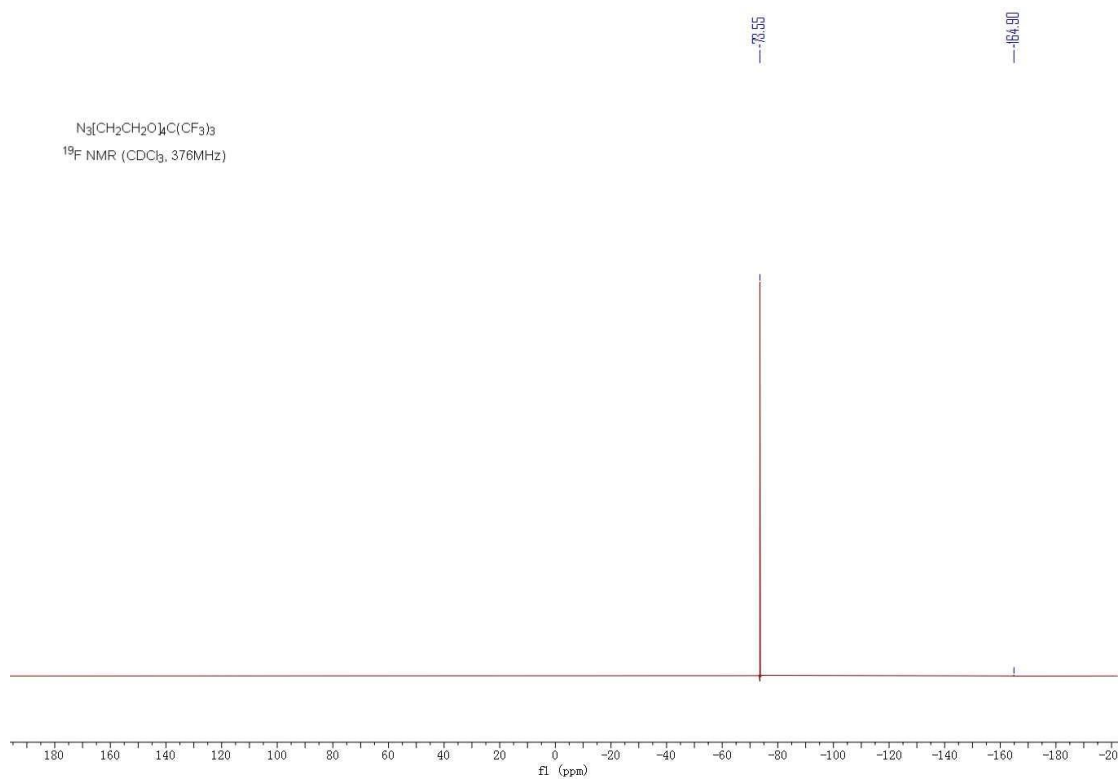
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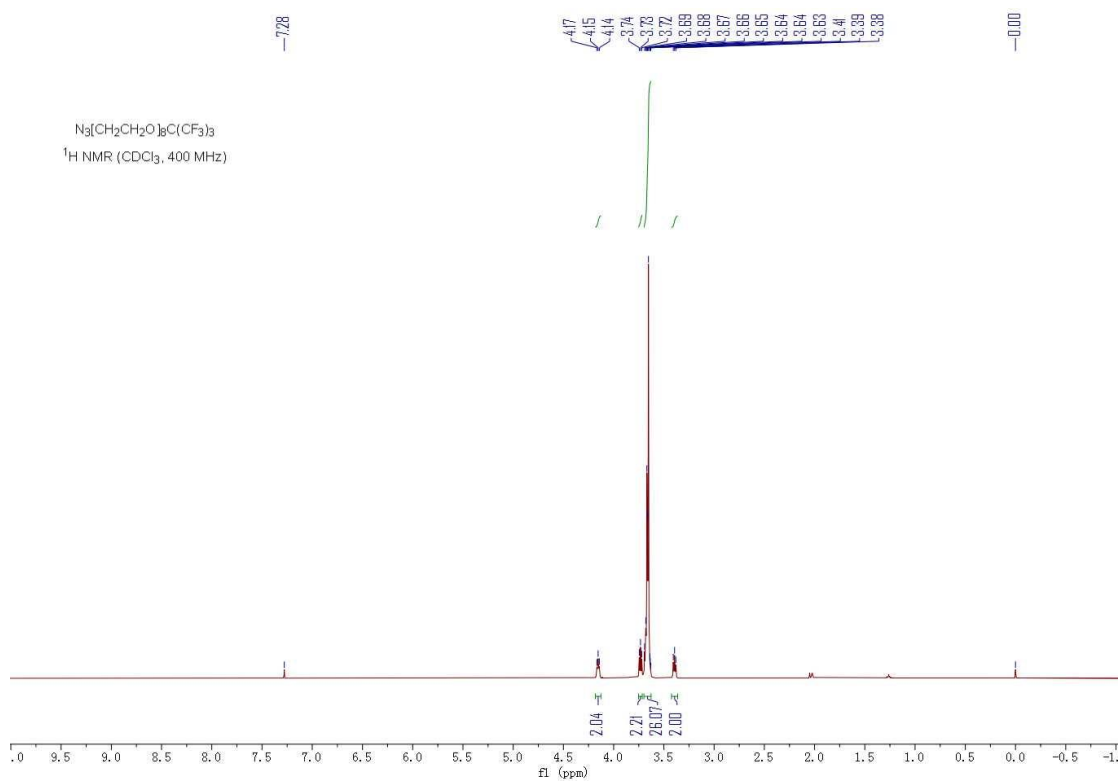
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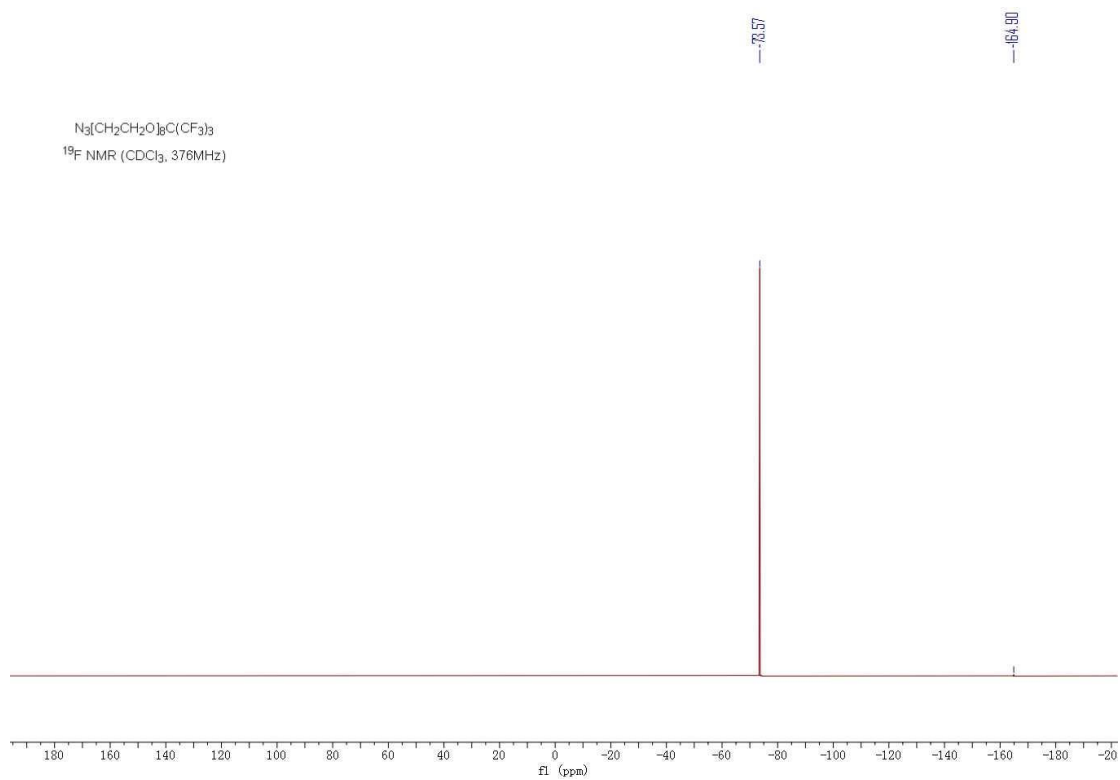
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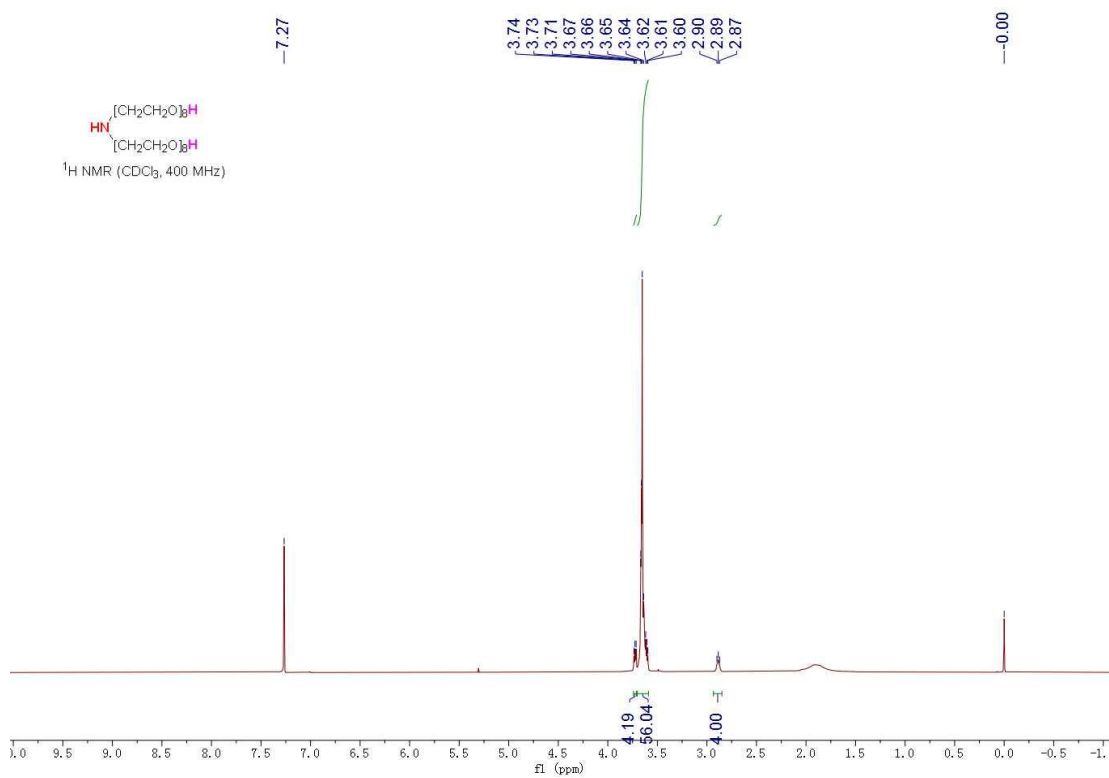
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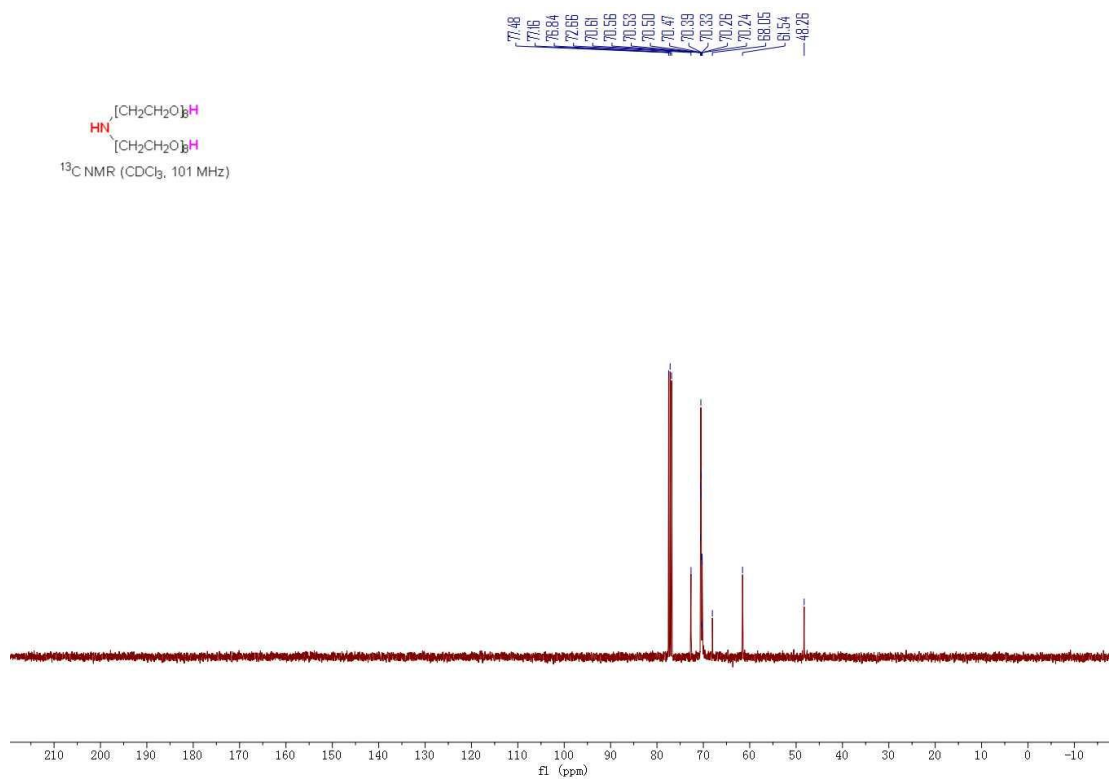
^{19}F NMR of compound **11**



¹H NMR of compound 12

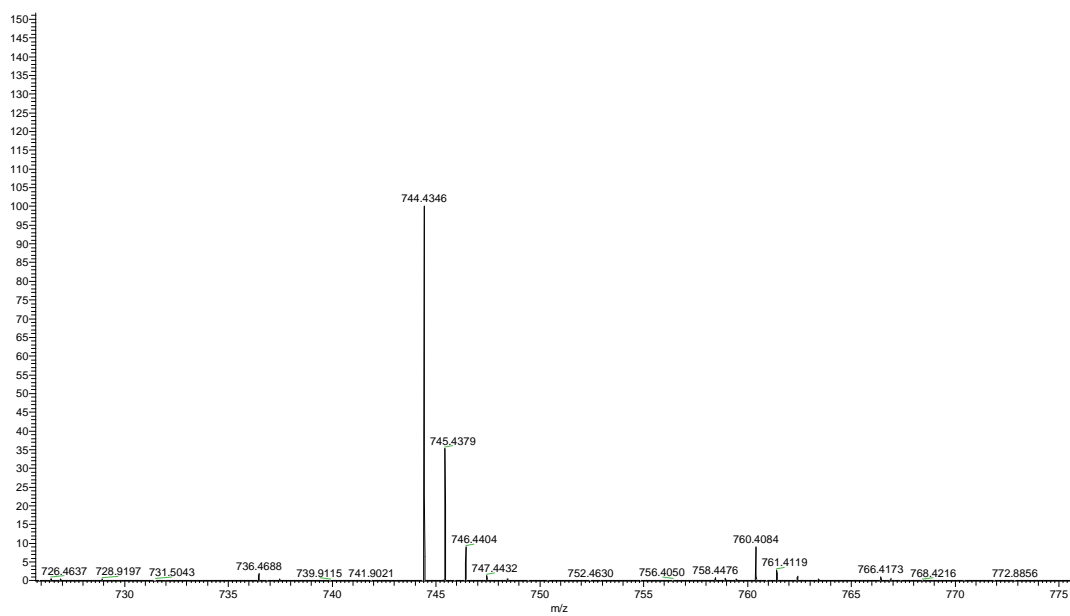


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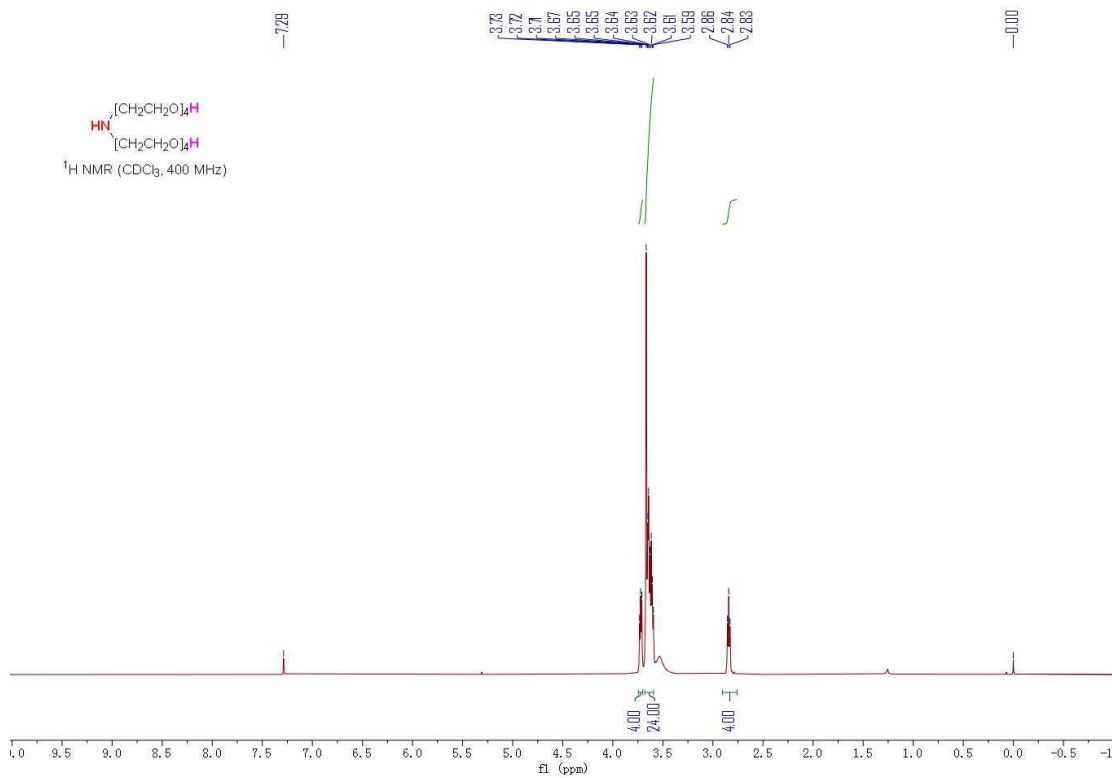


HRMS of compound 12

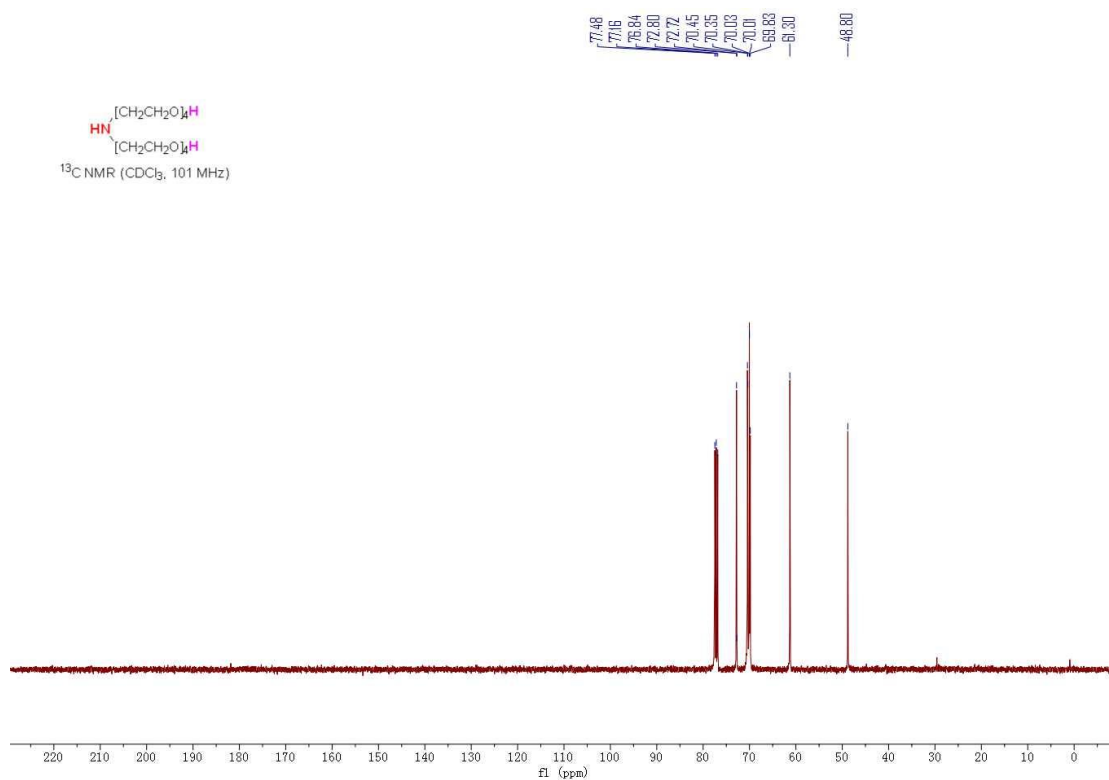
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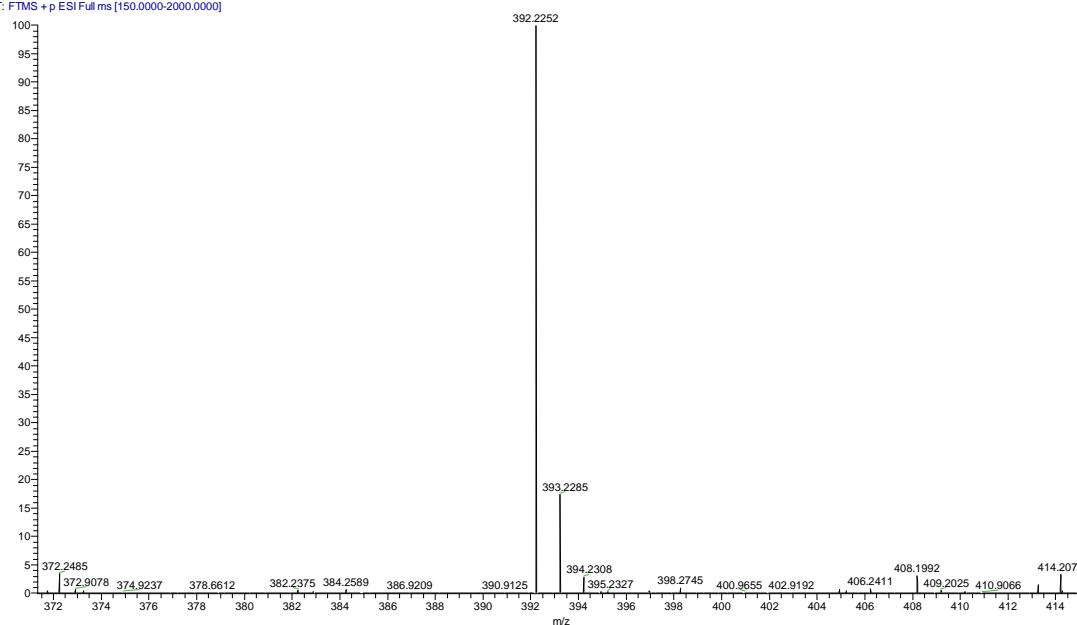


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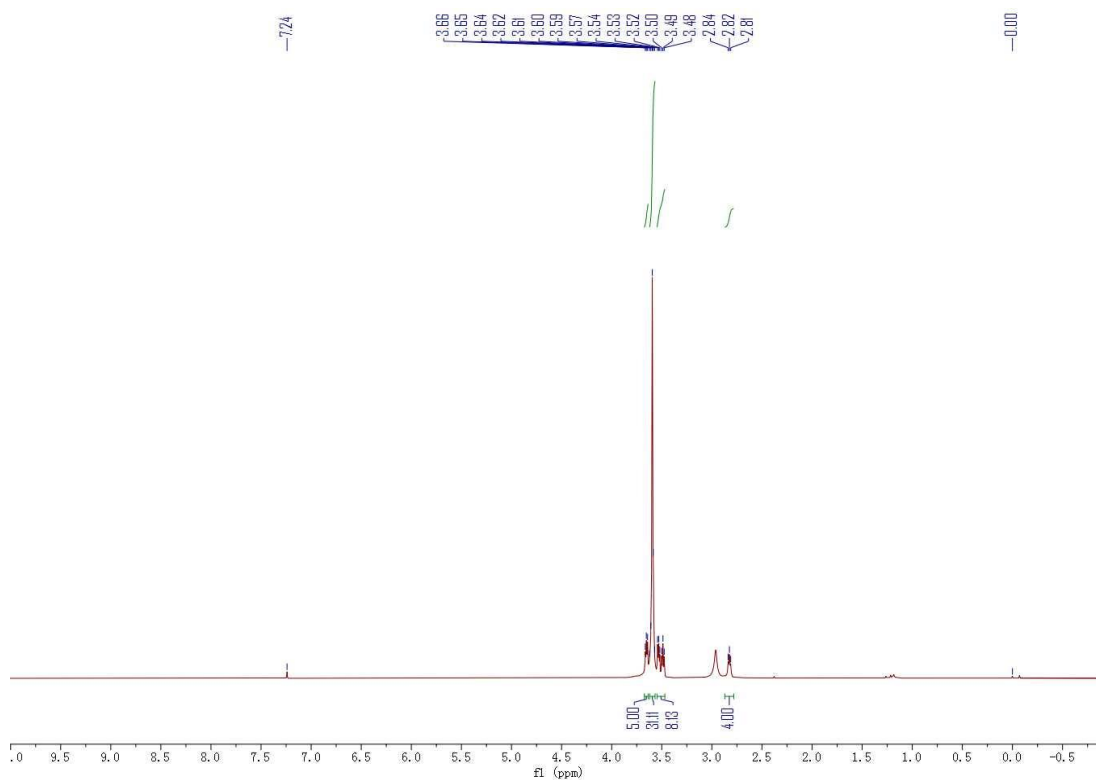


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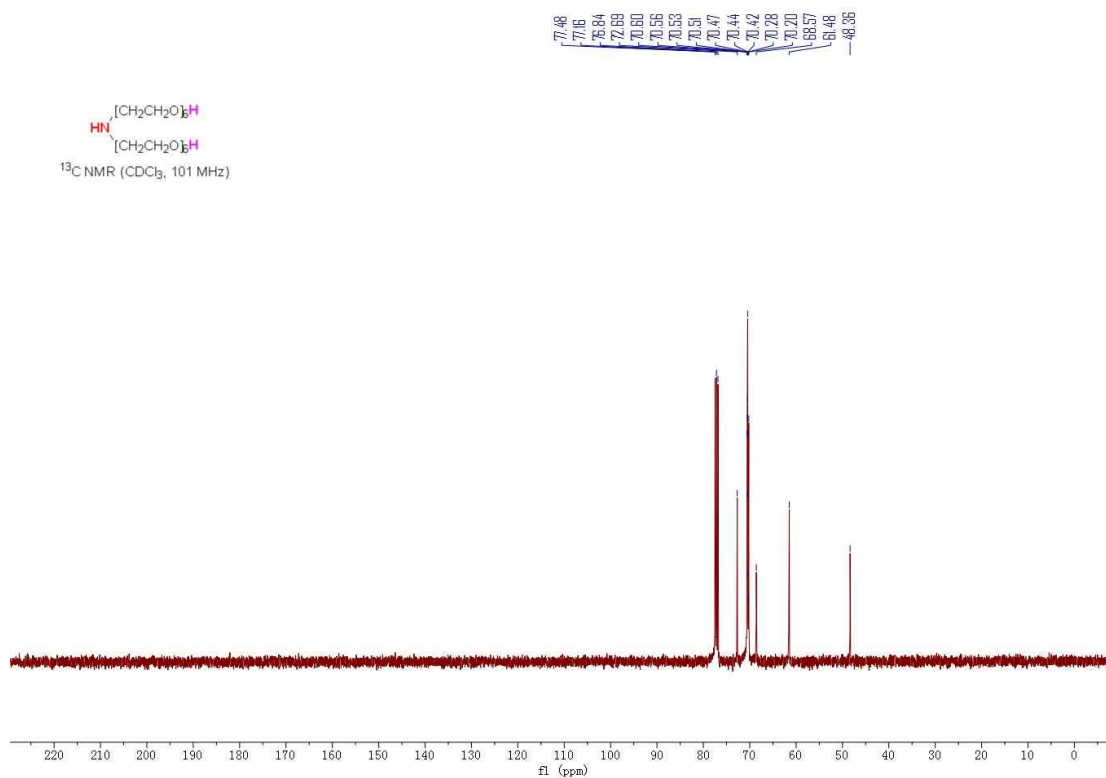
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¹H NMR of compound 14

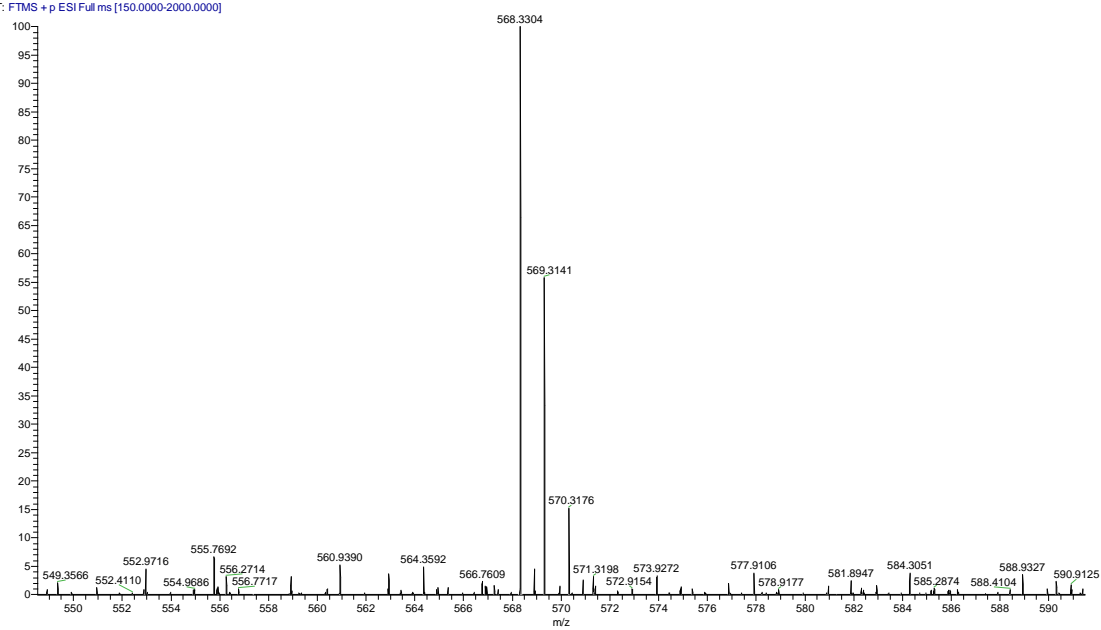


¹³C NMR of compound 14

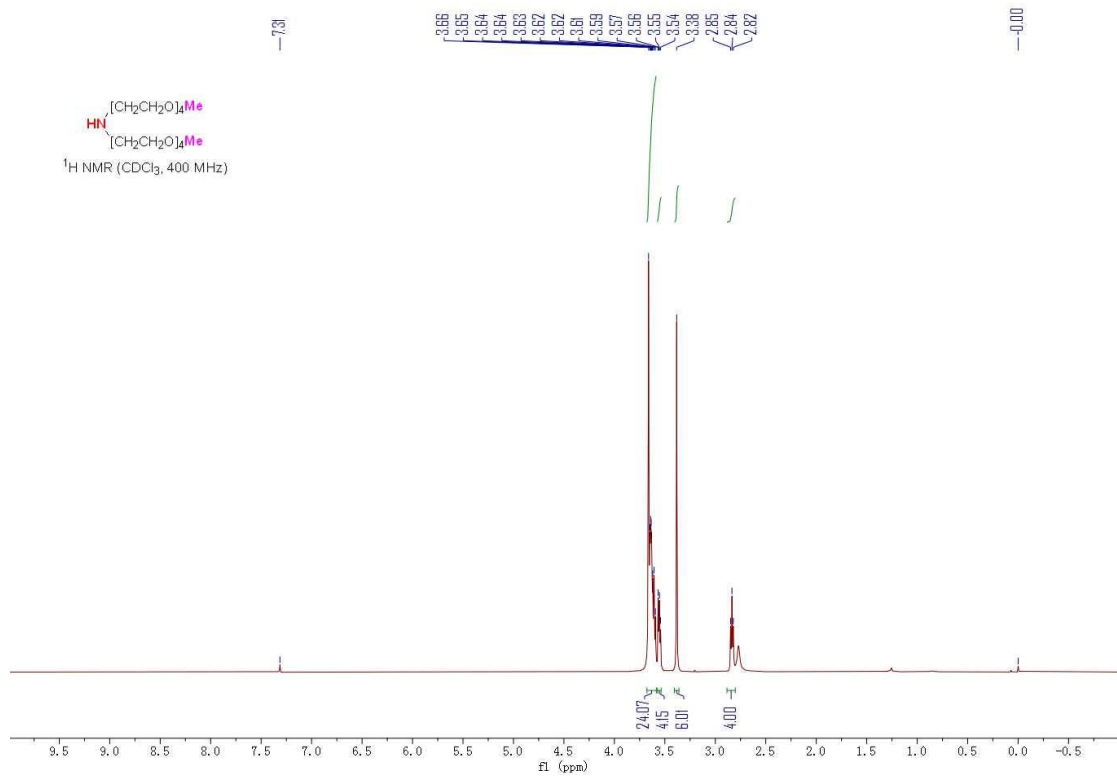


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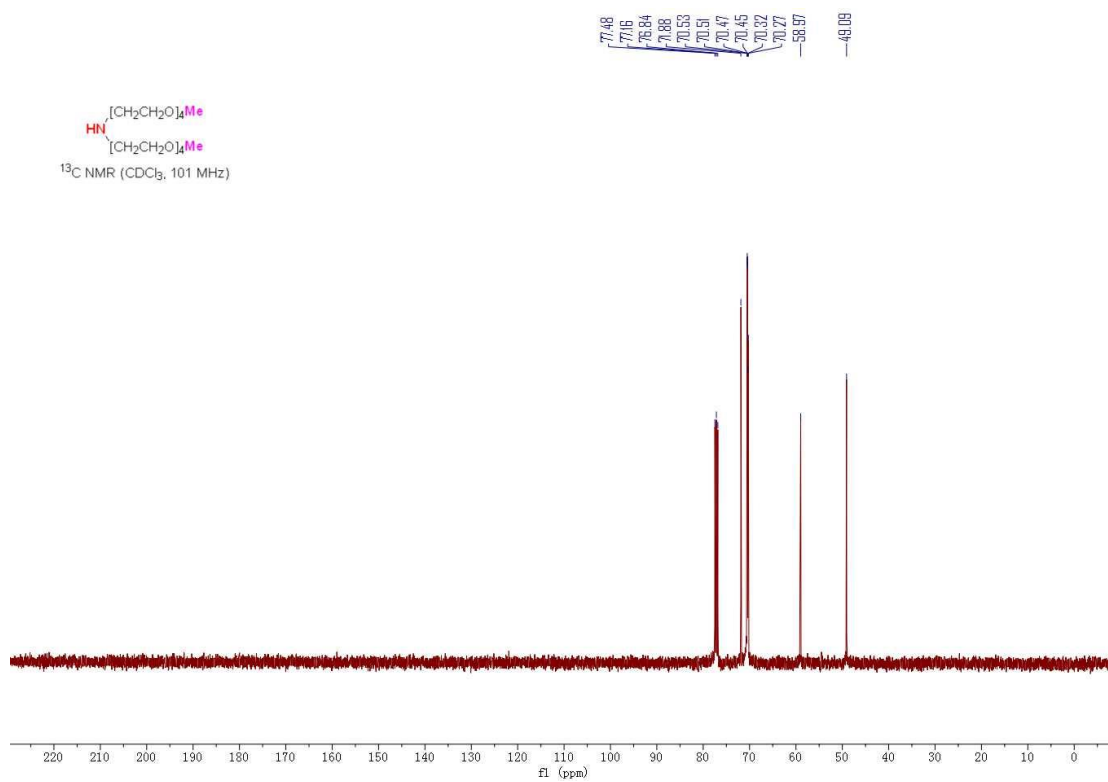
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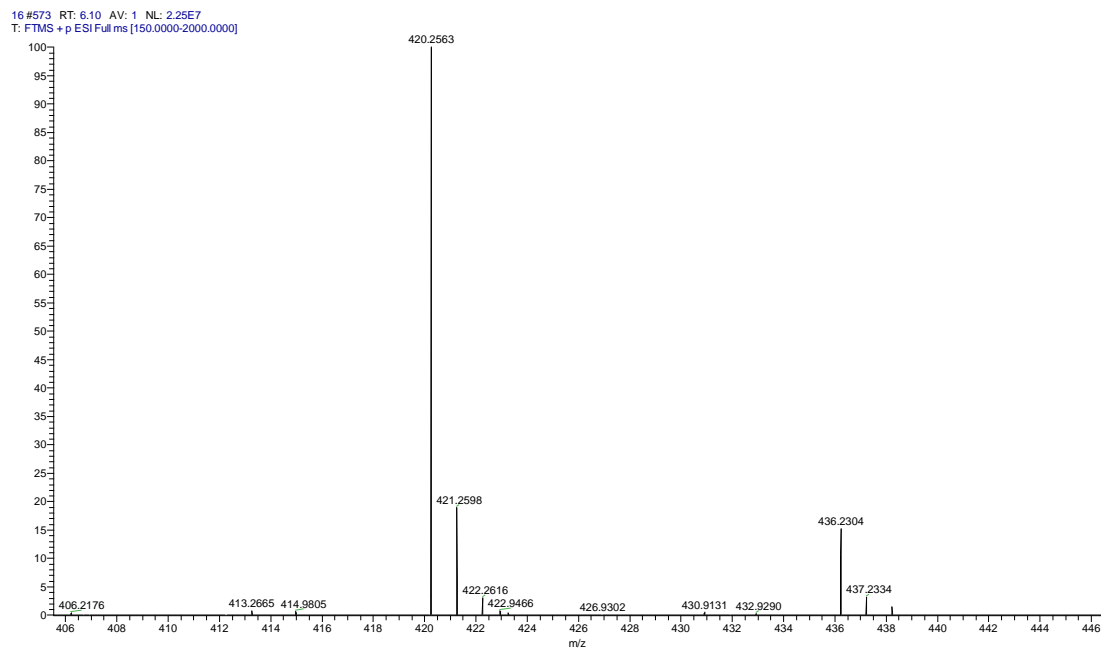
¹H NMR of compound 15



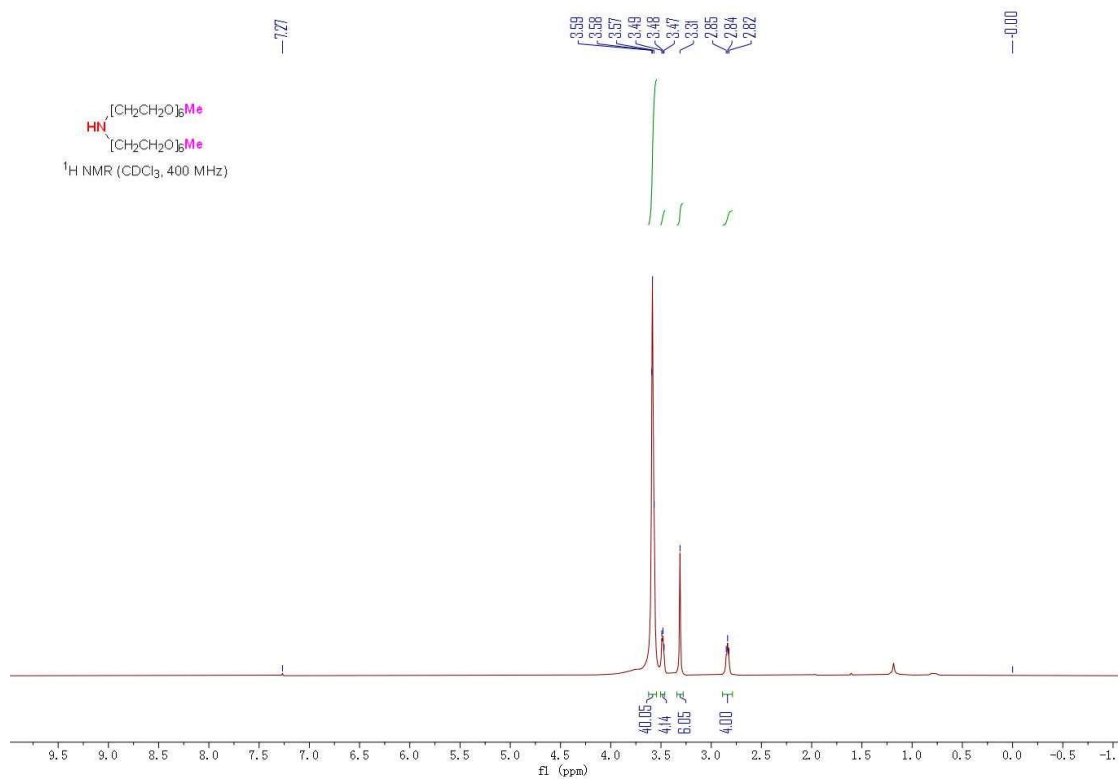
¹³C NMR of compound **15**



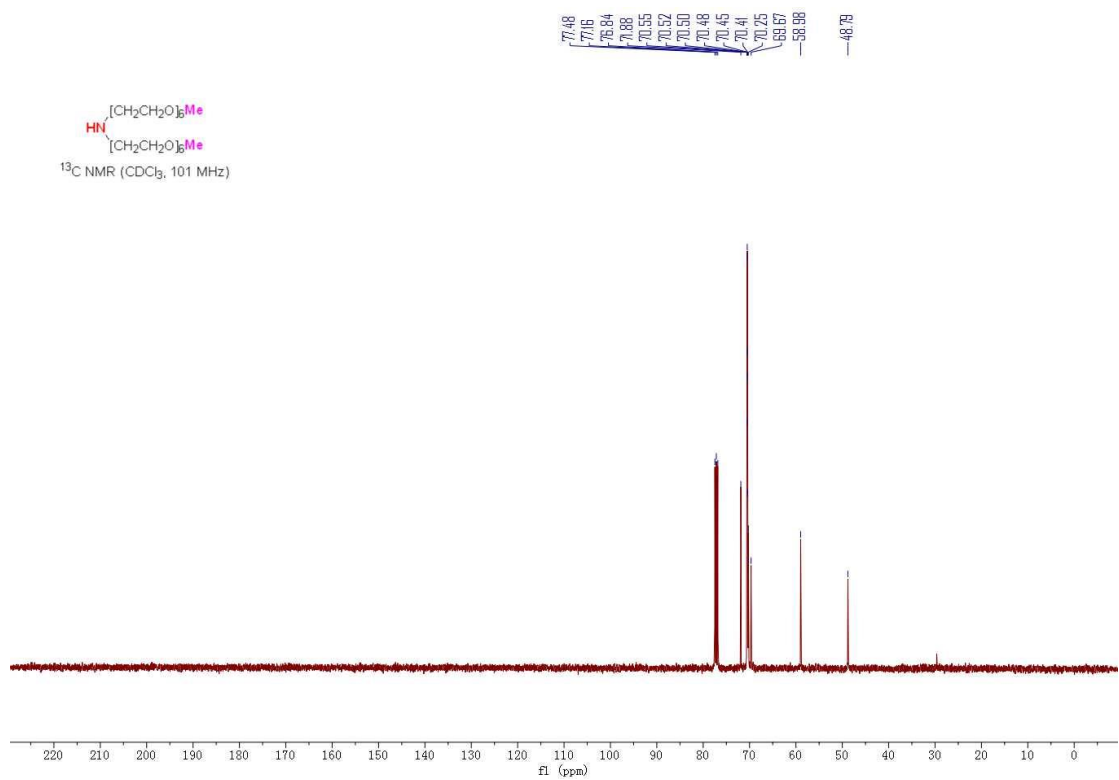
HRMS of compound **15**



¹H NMR of compound 16

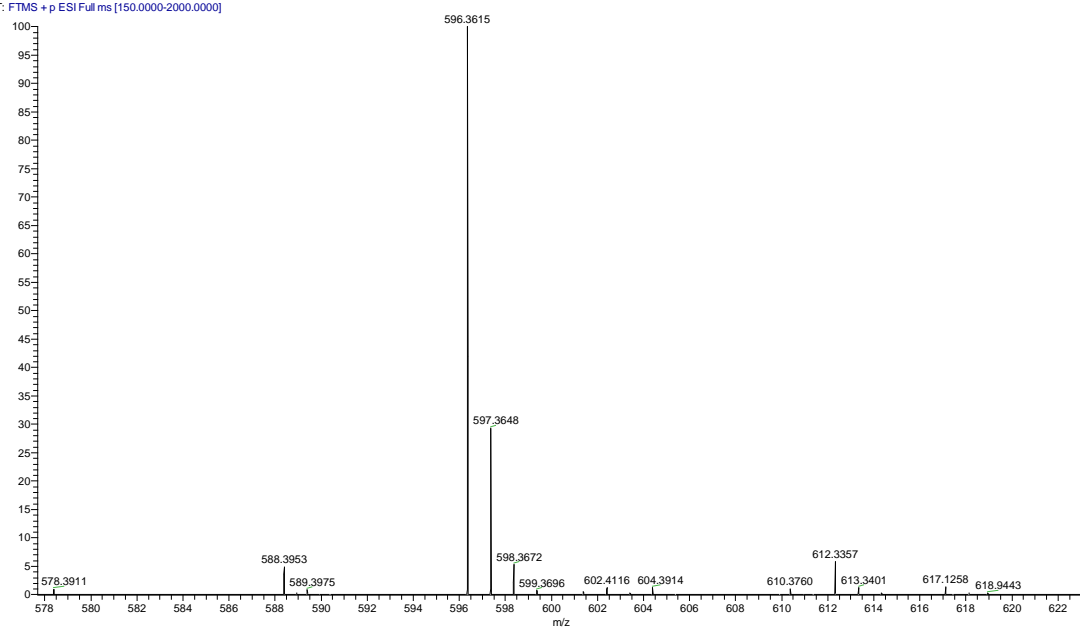


¹³C NMR of compound 16

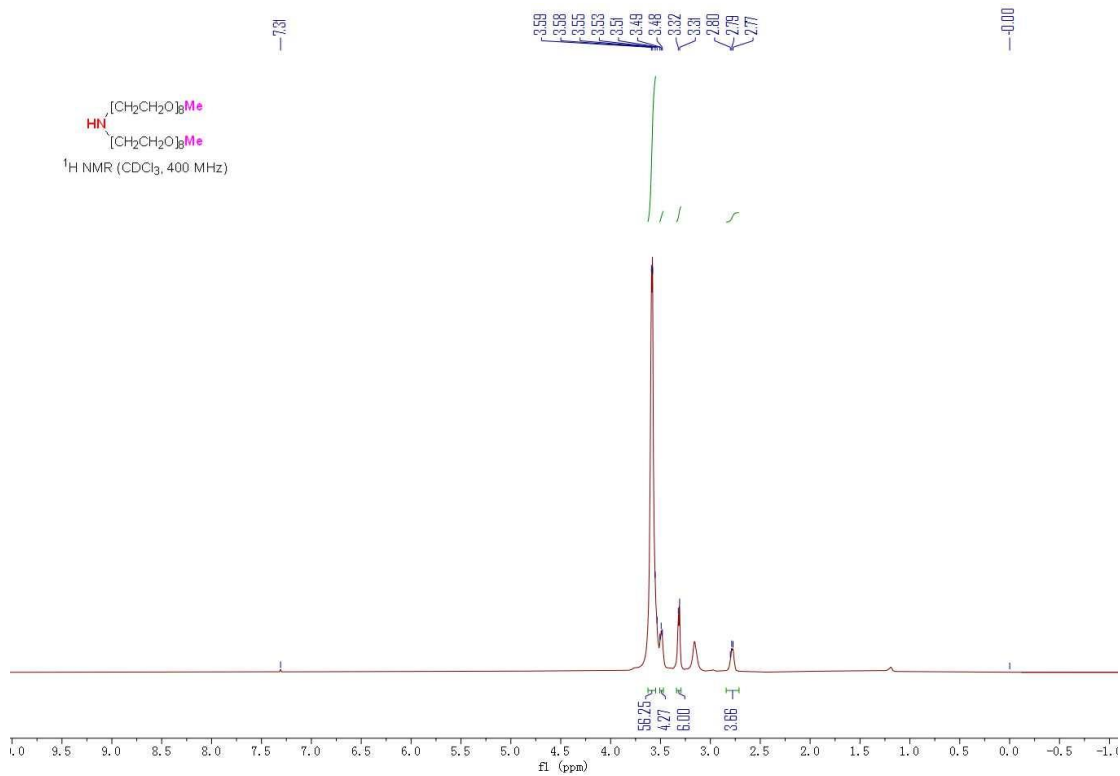


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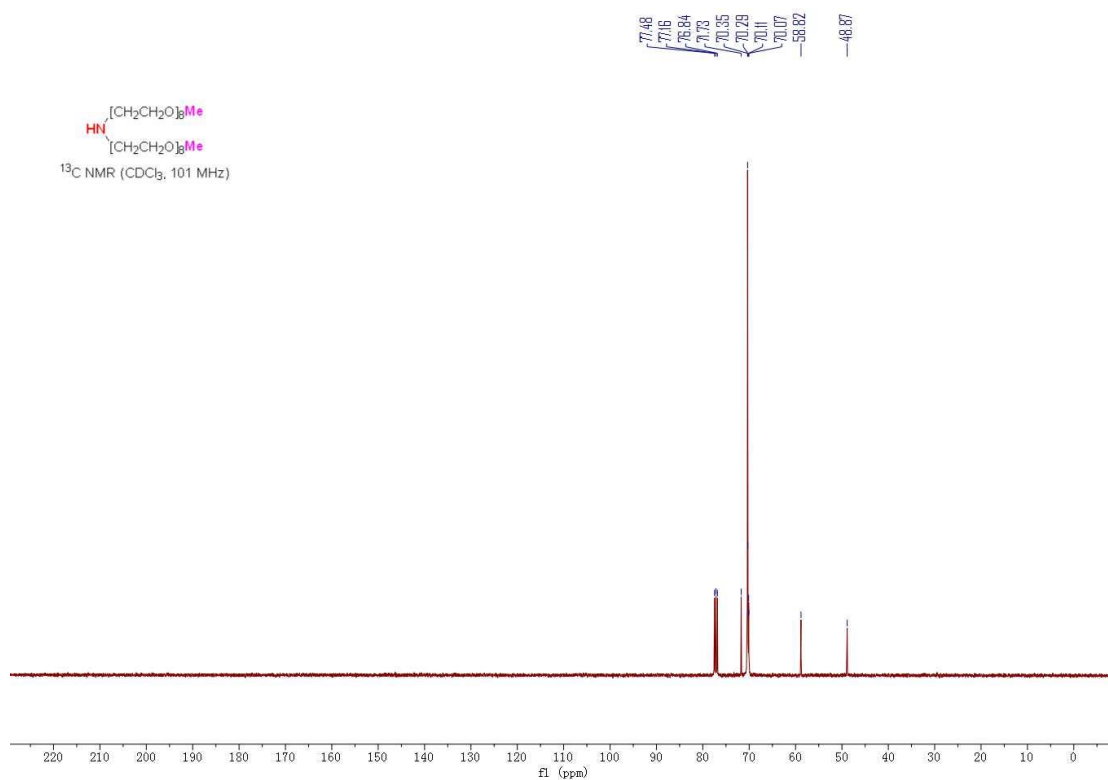
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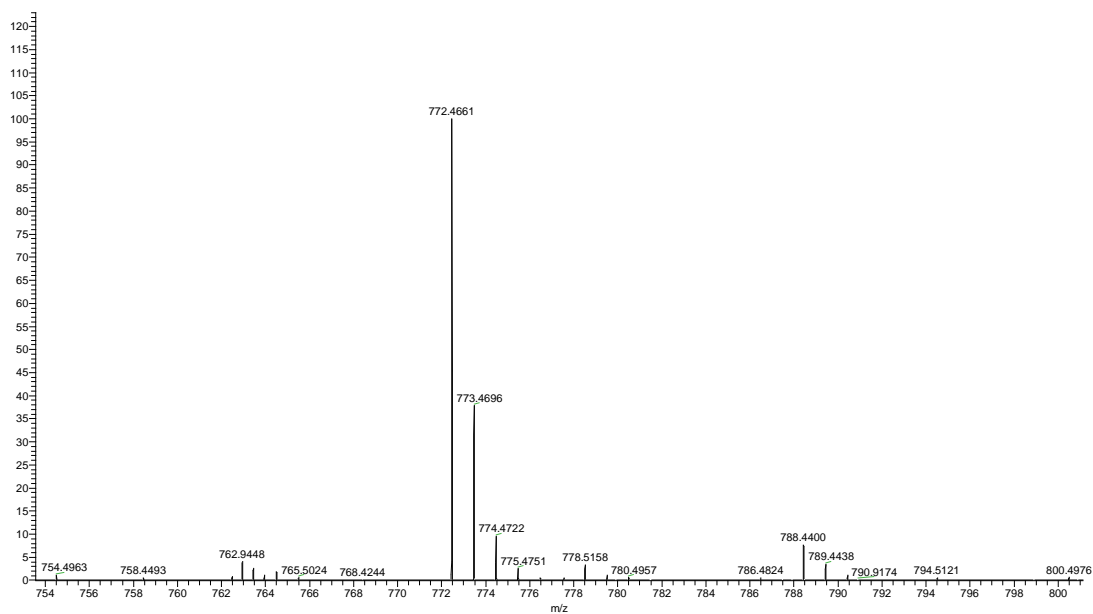


¹³C NMR of compound 17

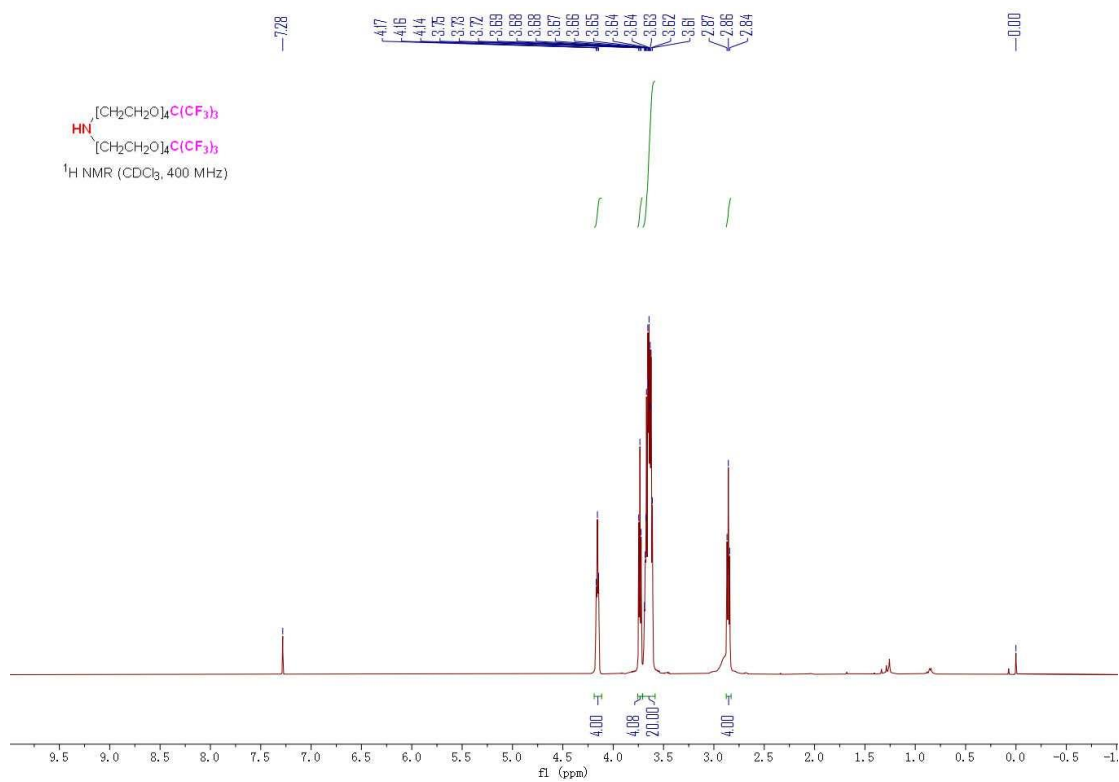


HRMS of compound 17

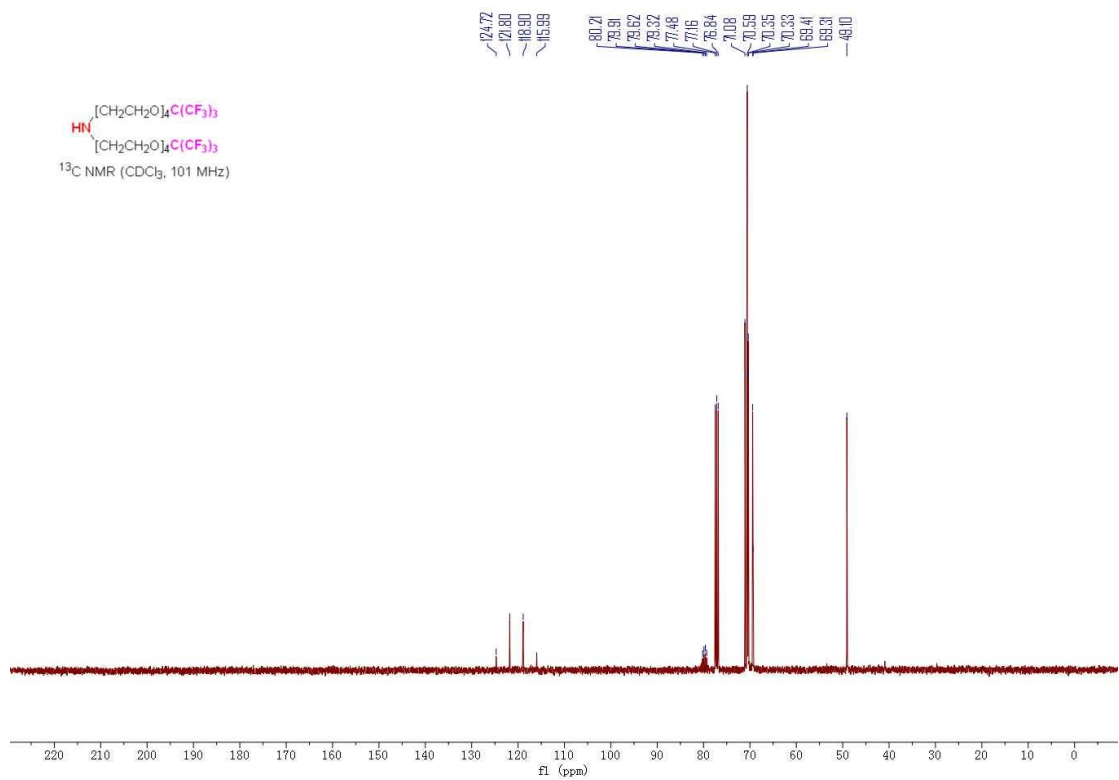
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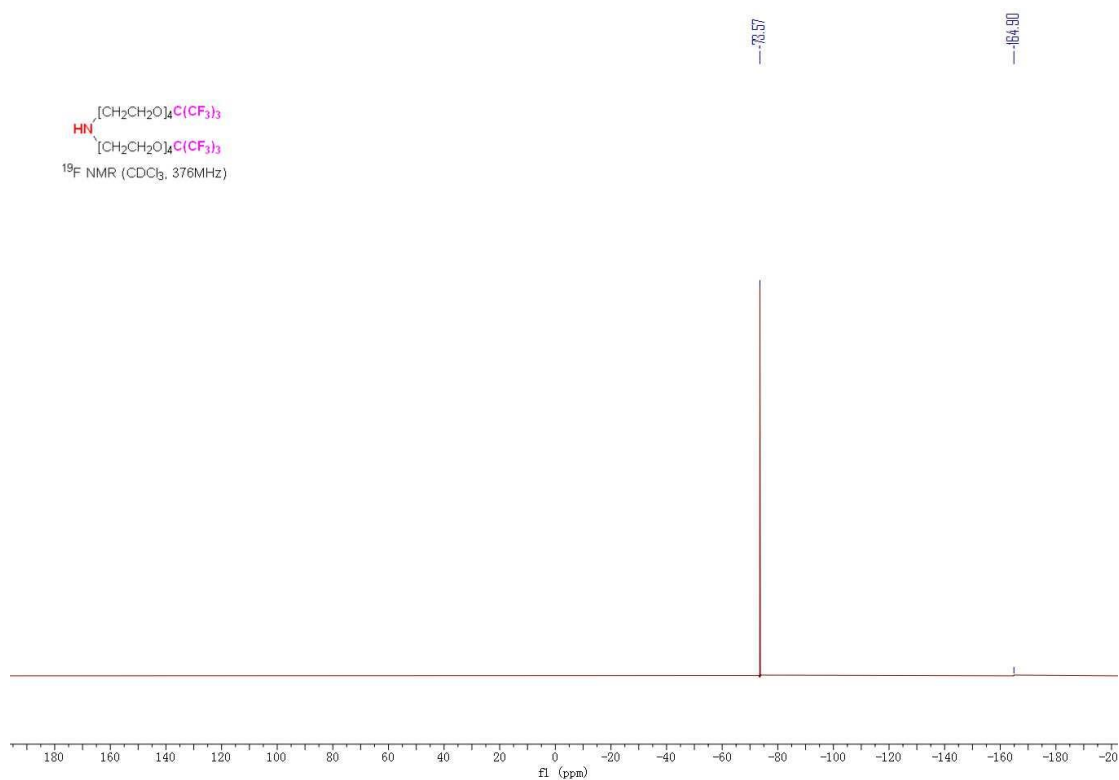
¹H NMR of compound 18



¹³C NMR of compound 18

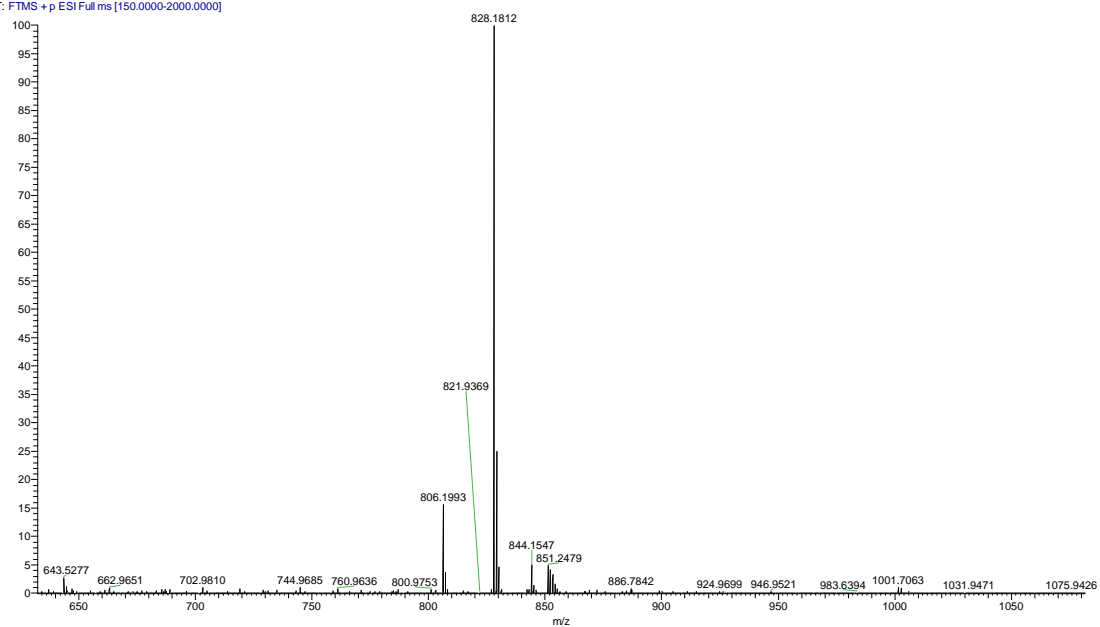


¹⁹F NMR of compound **18**

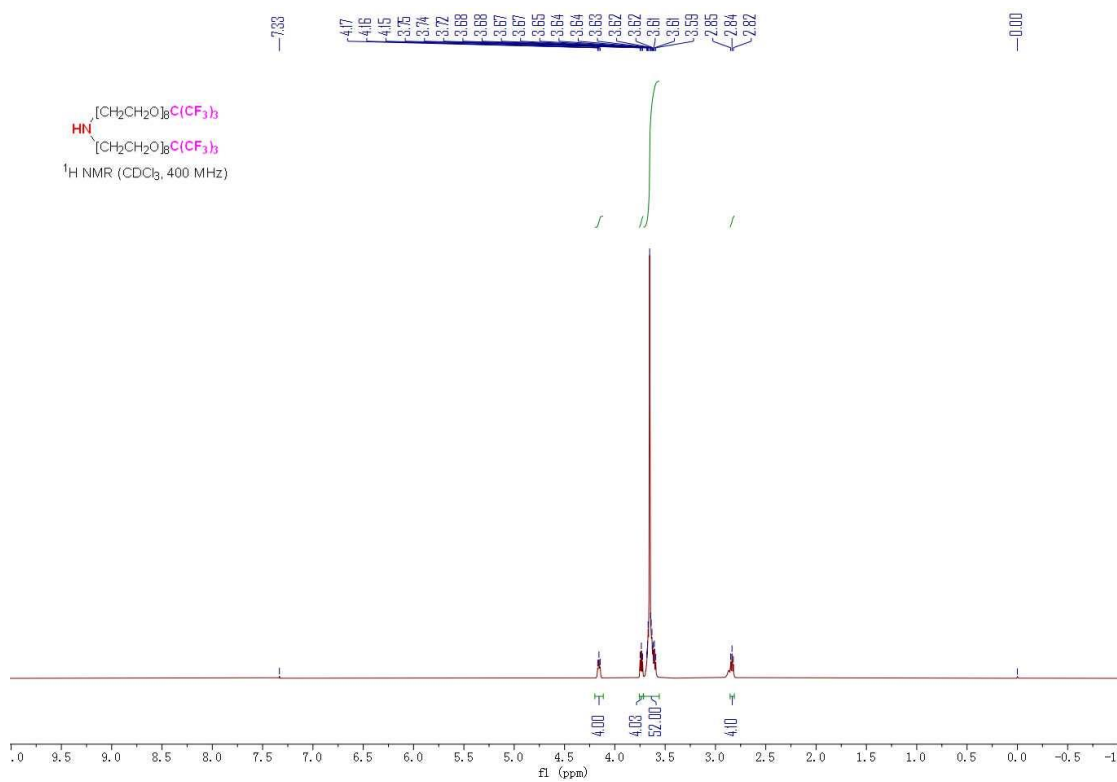


HRMS of compound **18**

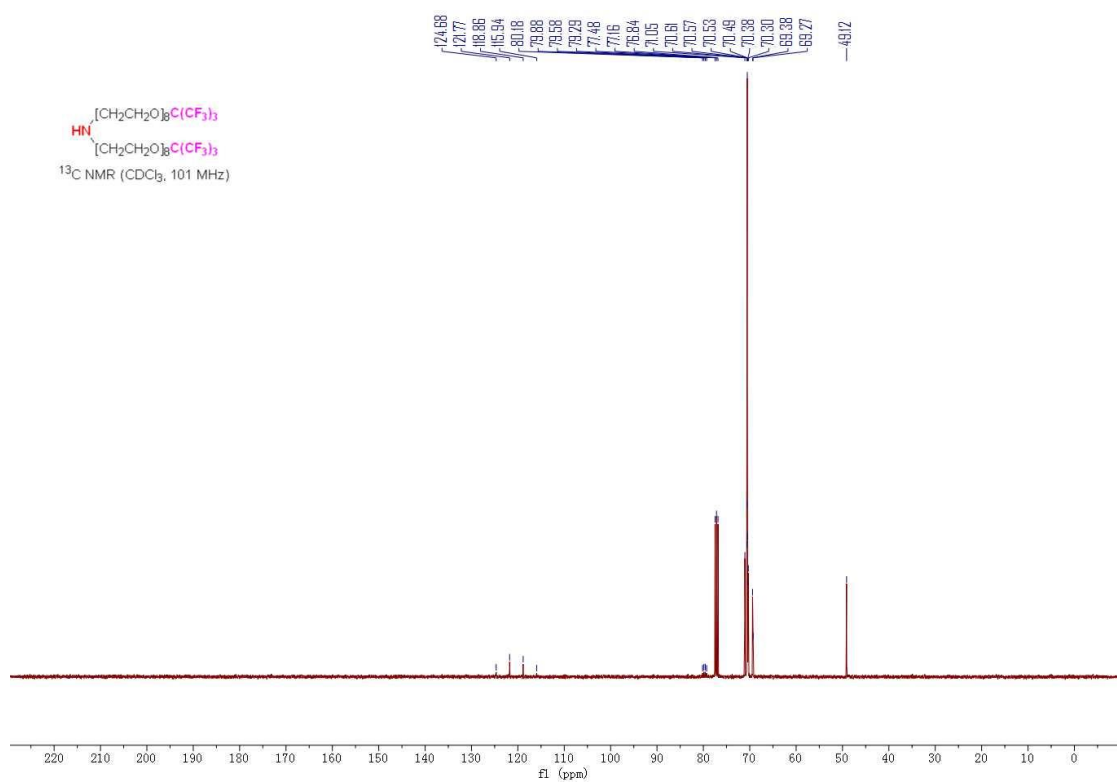
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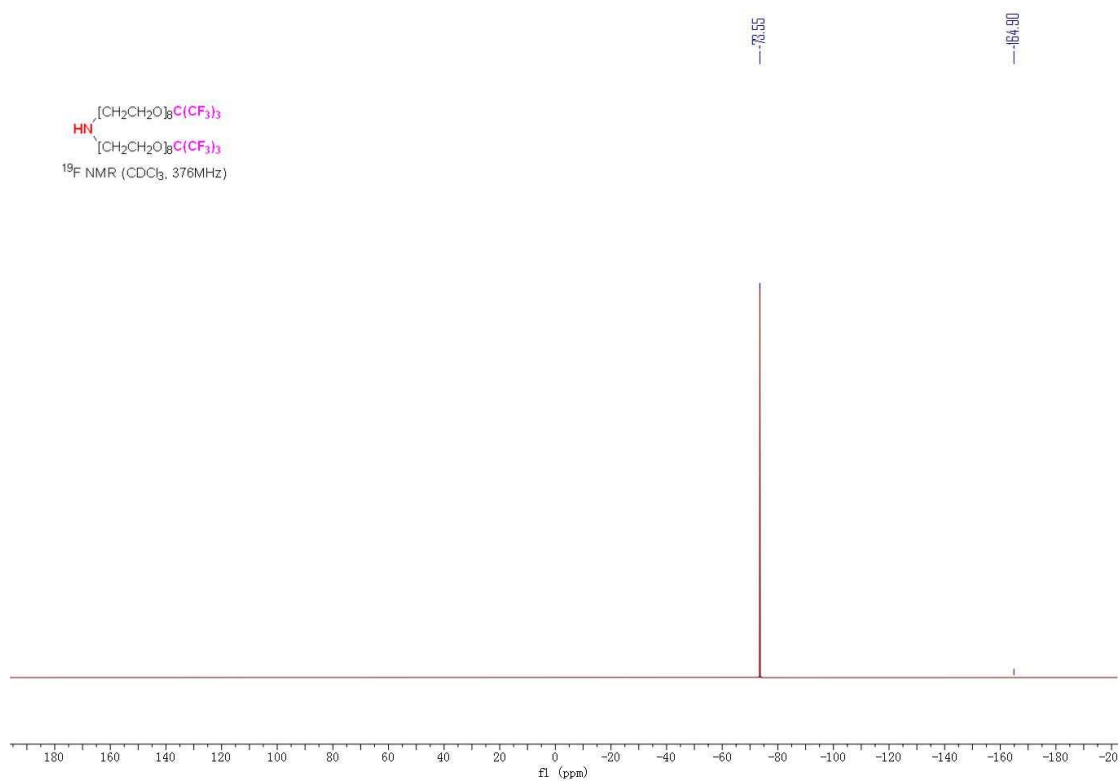
¹H NMR of compound 19



¹³C NMR of compound 19

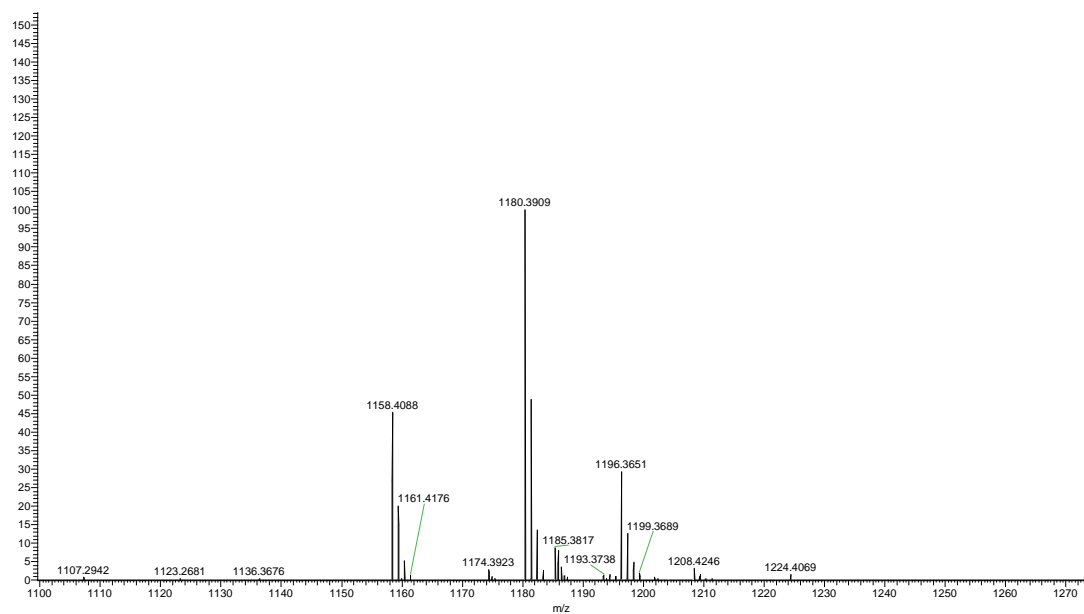


¹⁹F NMR of compound **19**

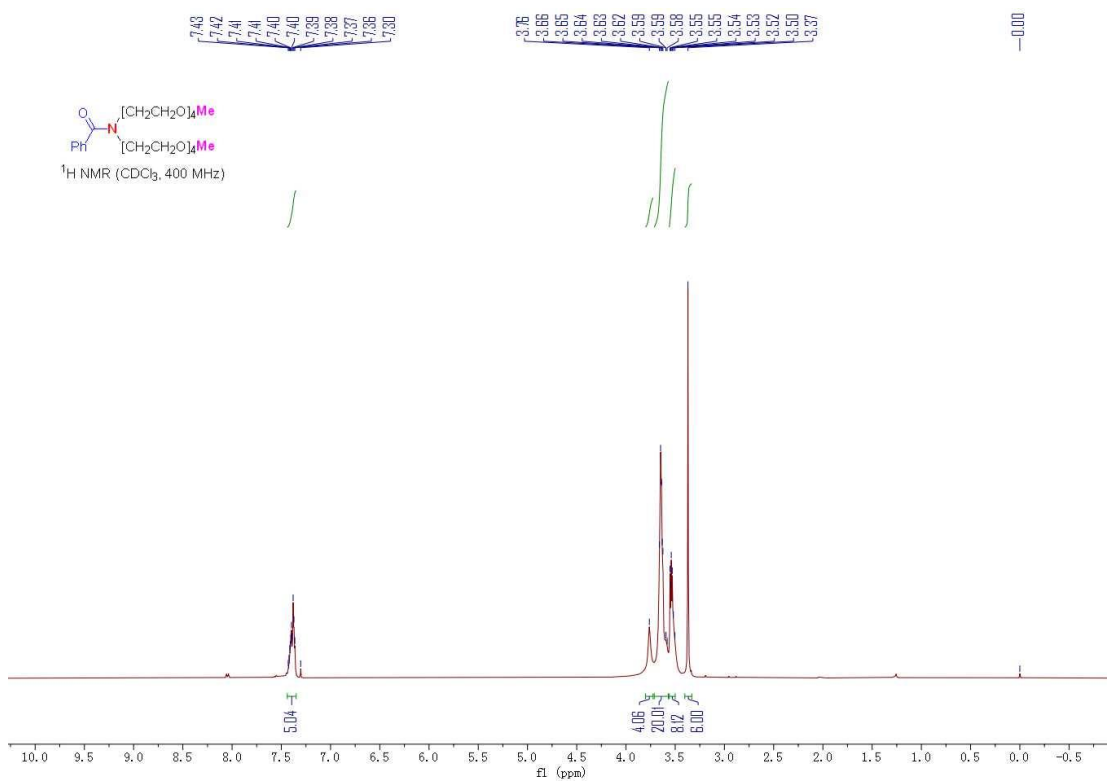


HRMS of compound **19**

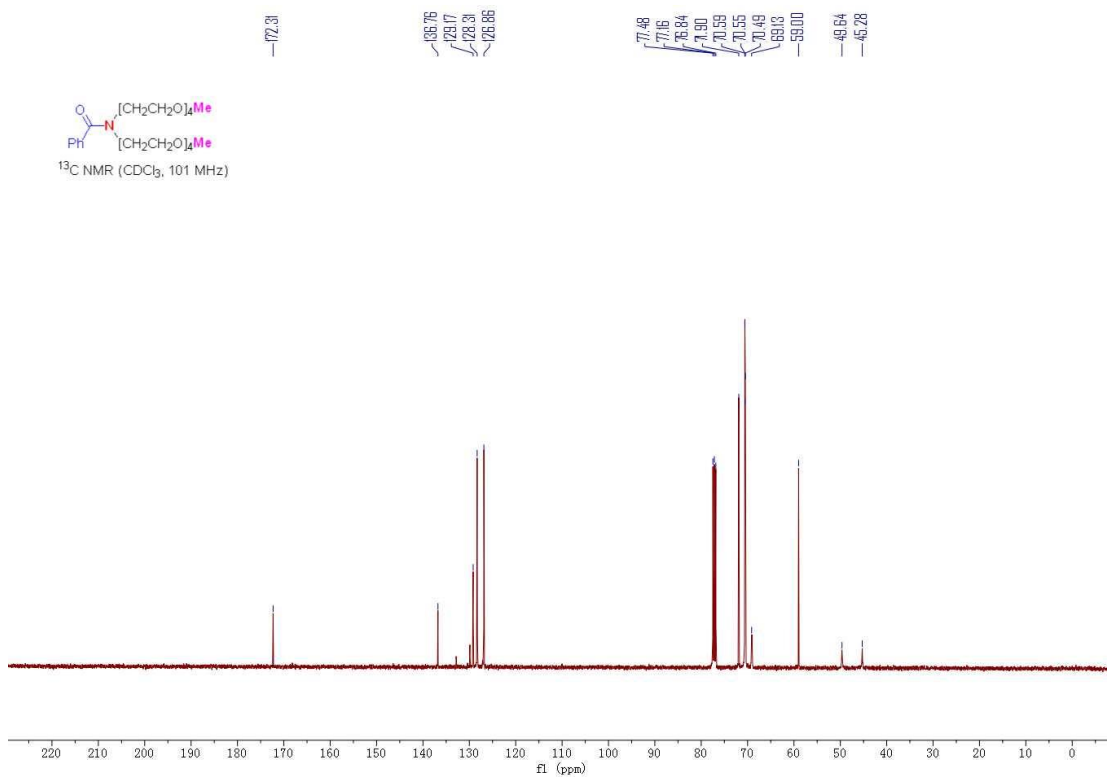
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¹H NMR of compound 20

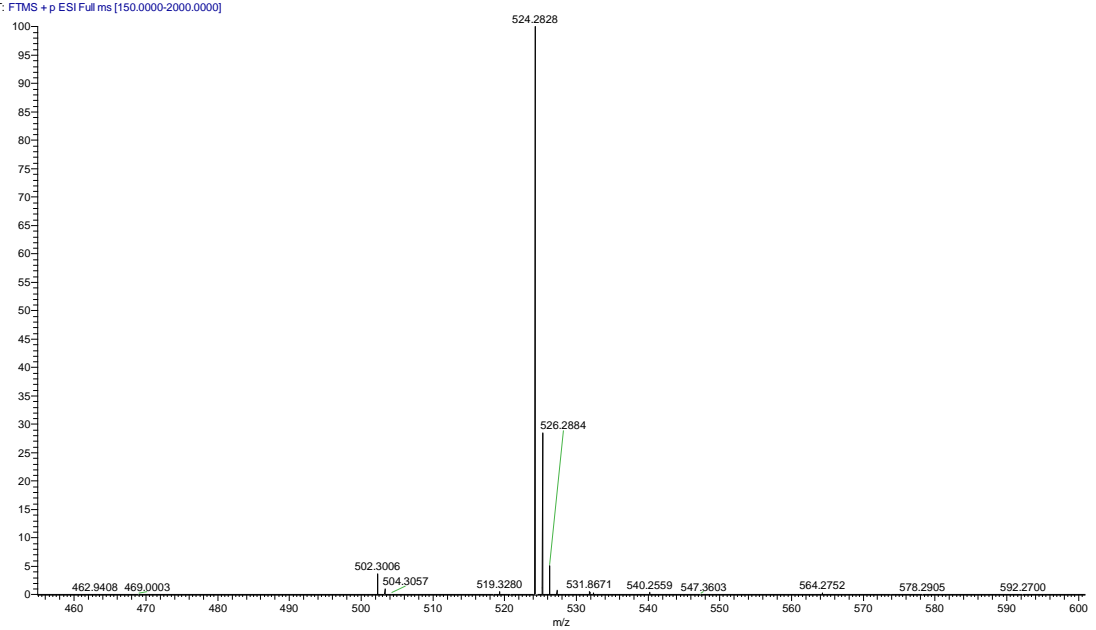


¹³C NMR of compound 20

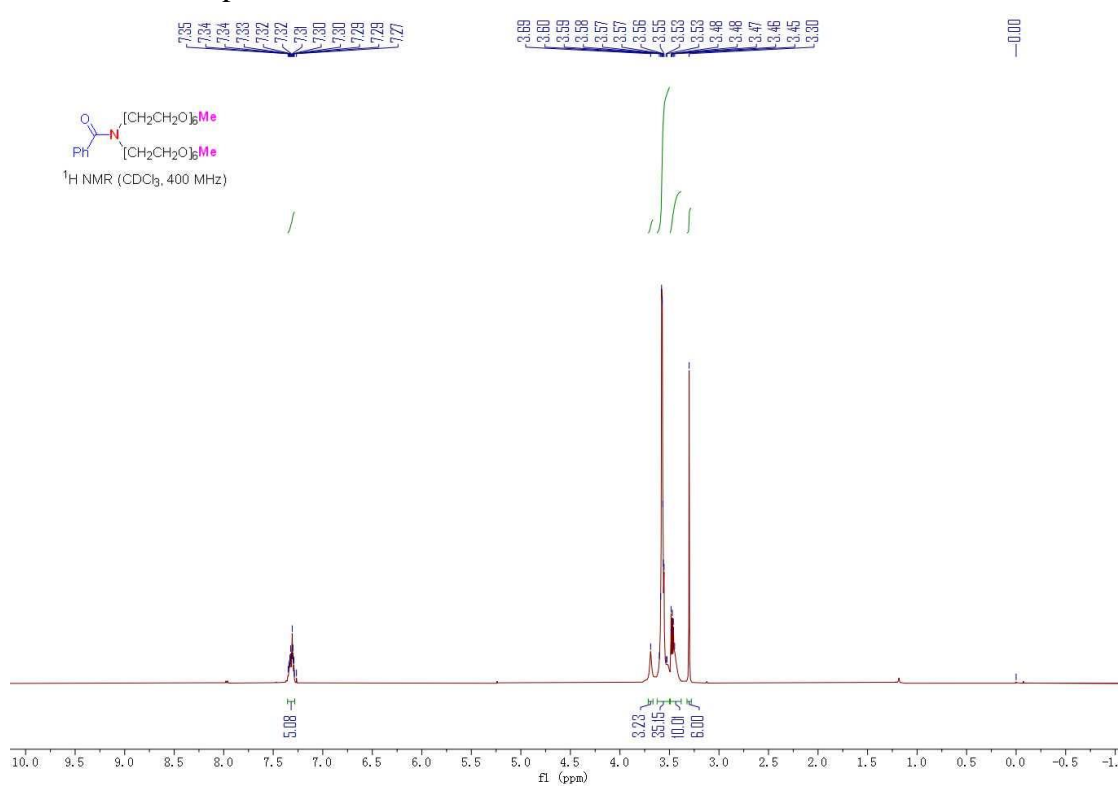
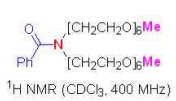


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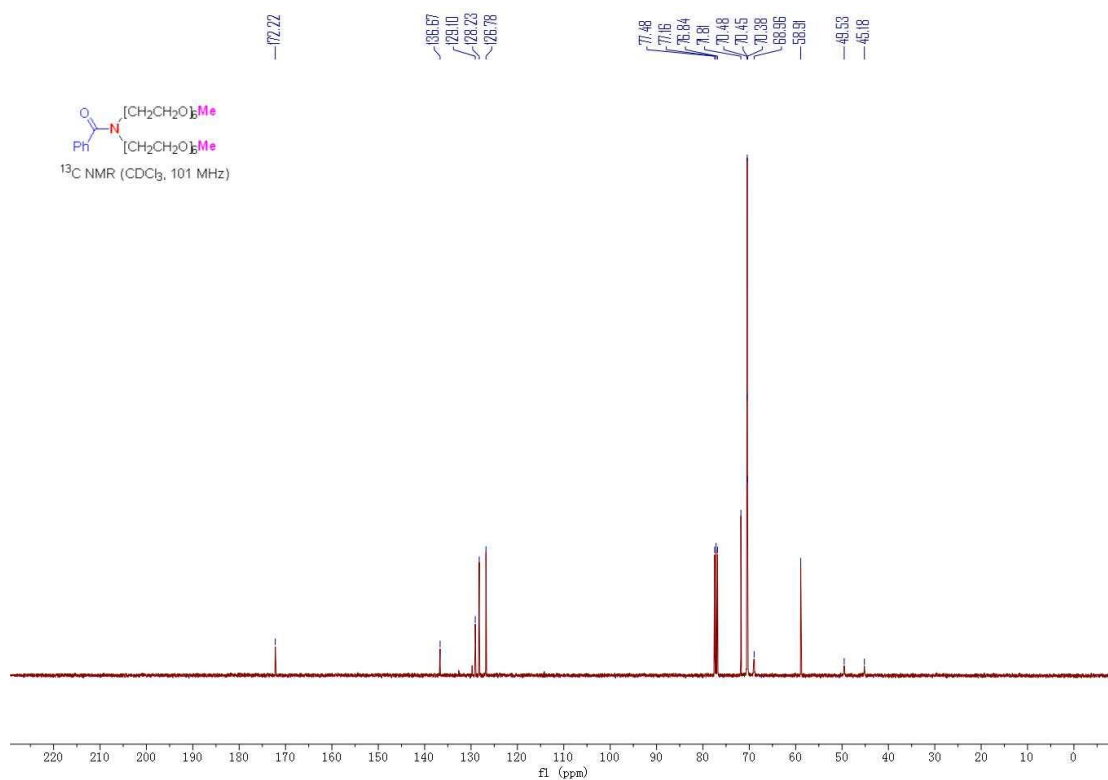
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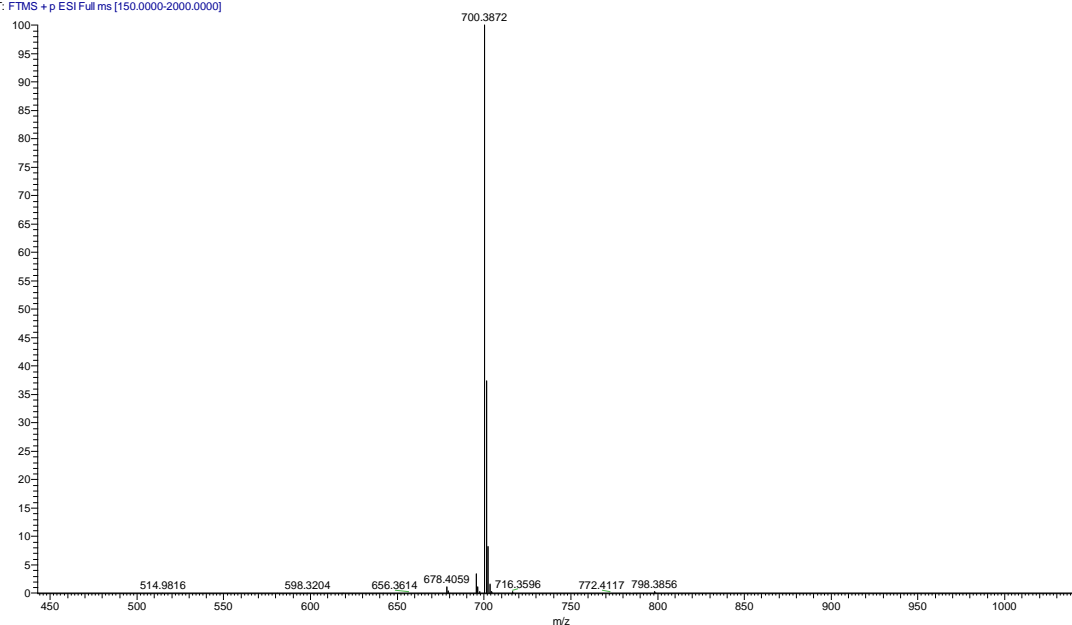


¹³C NMR of compound 21

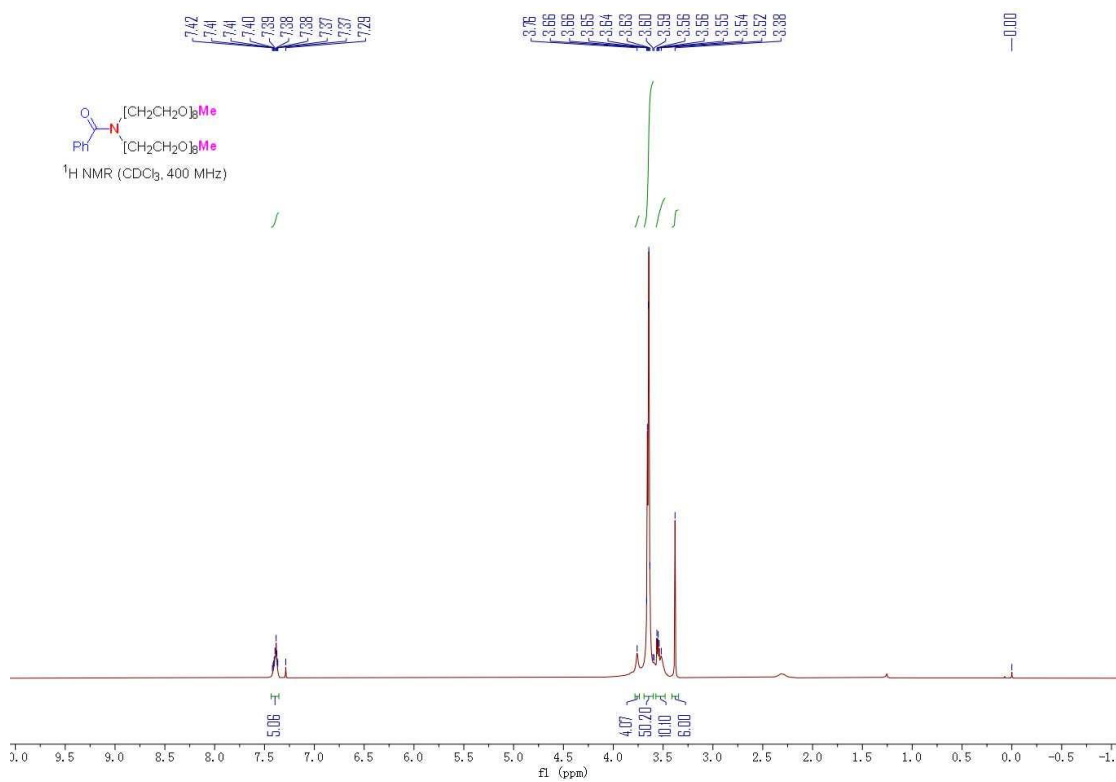


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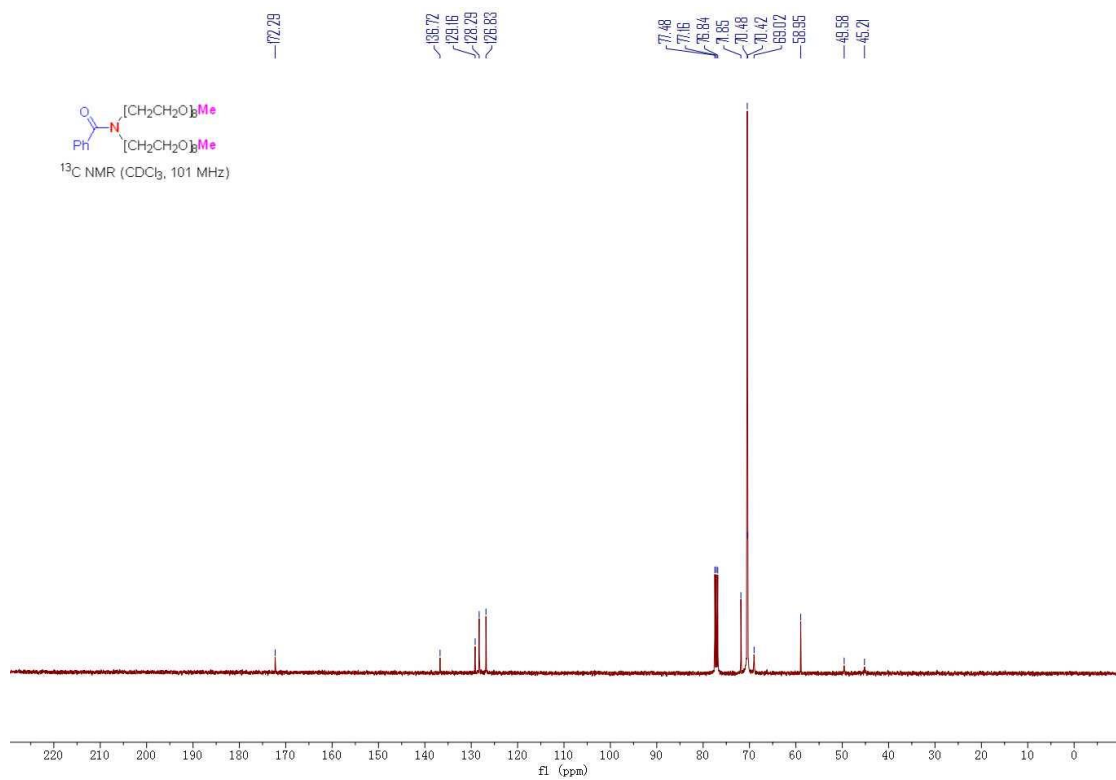
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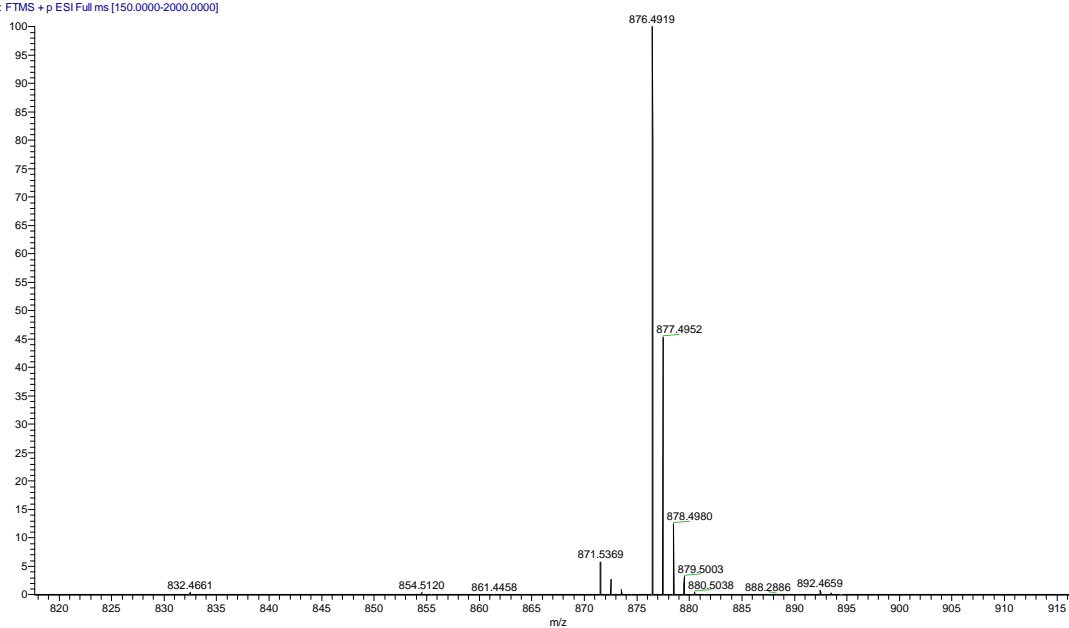


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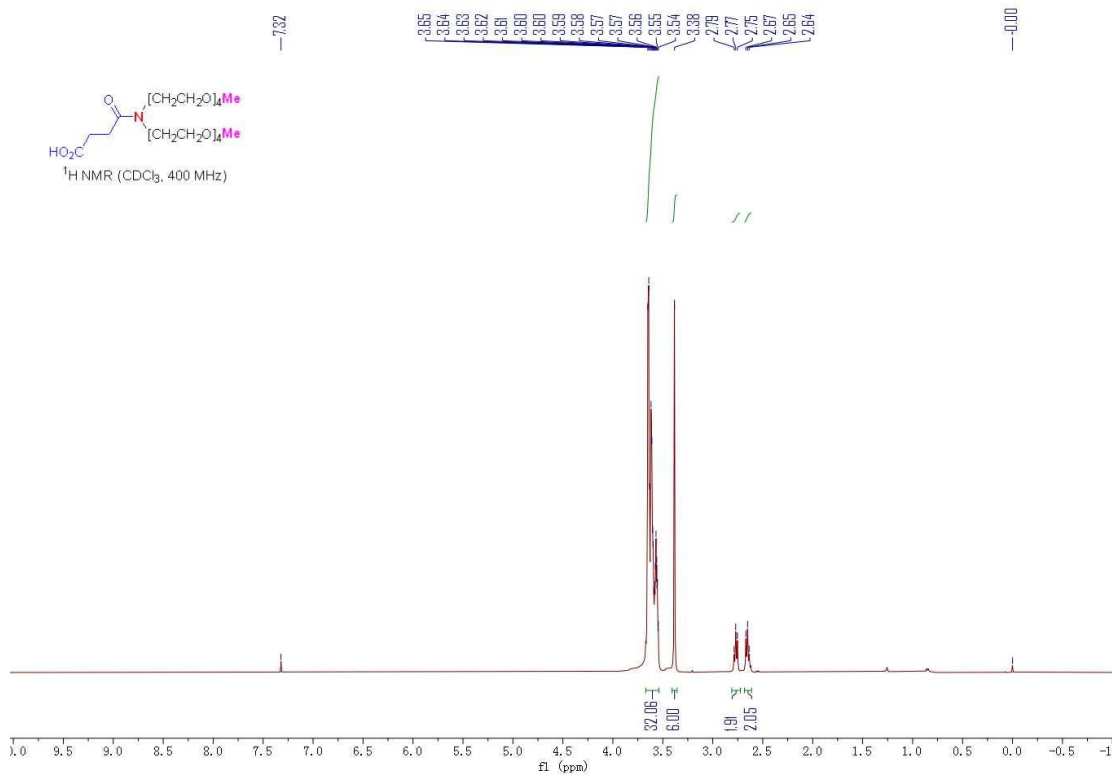


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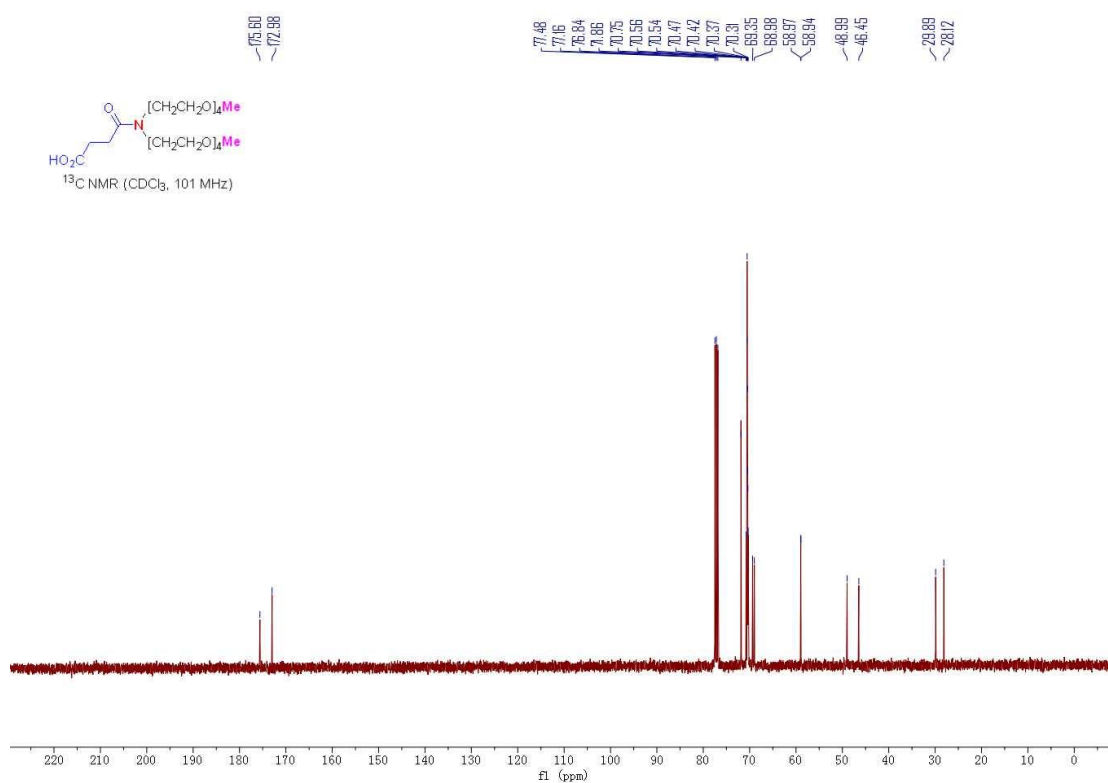
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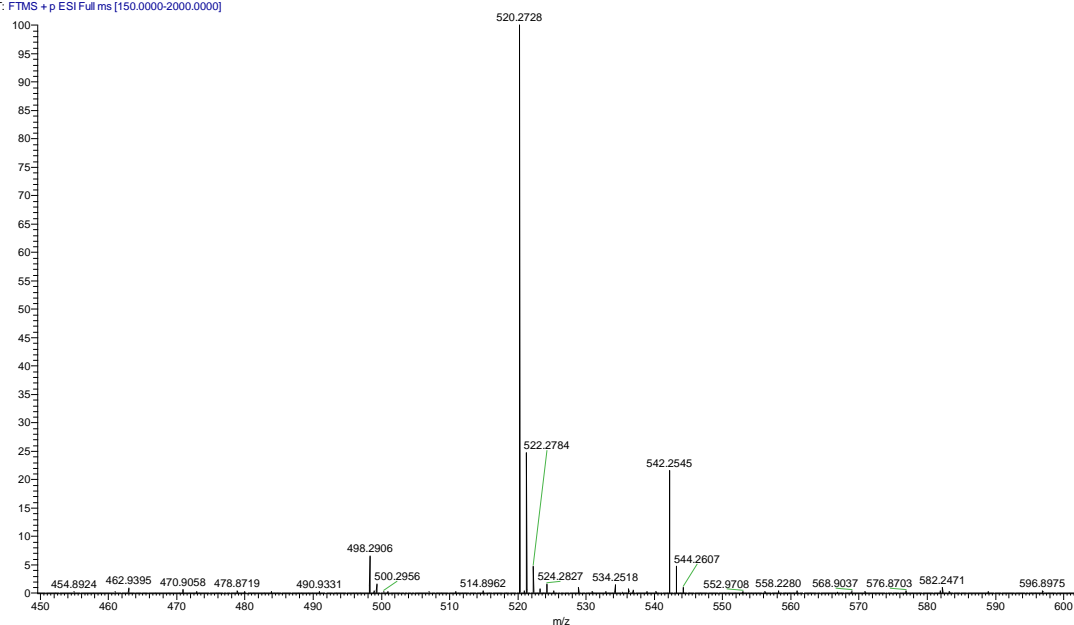


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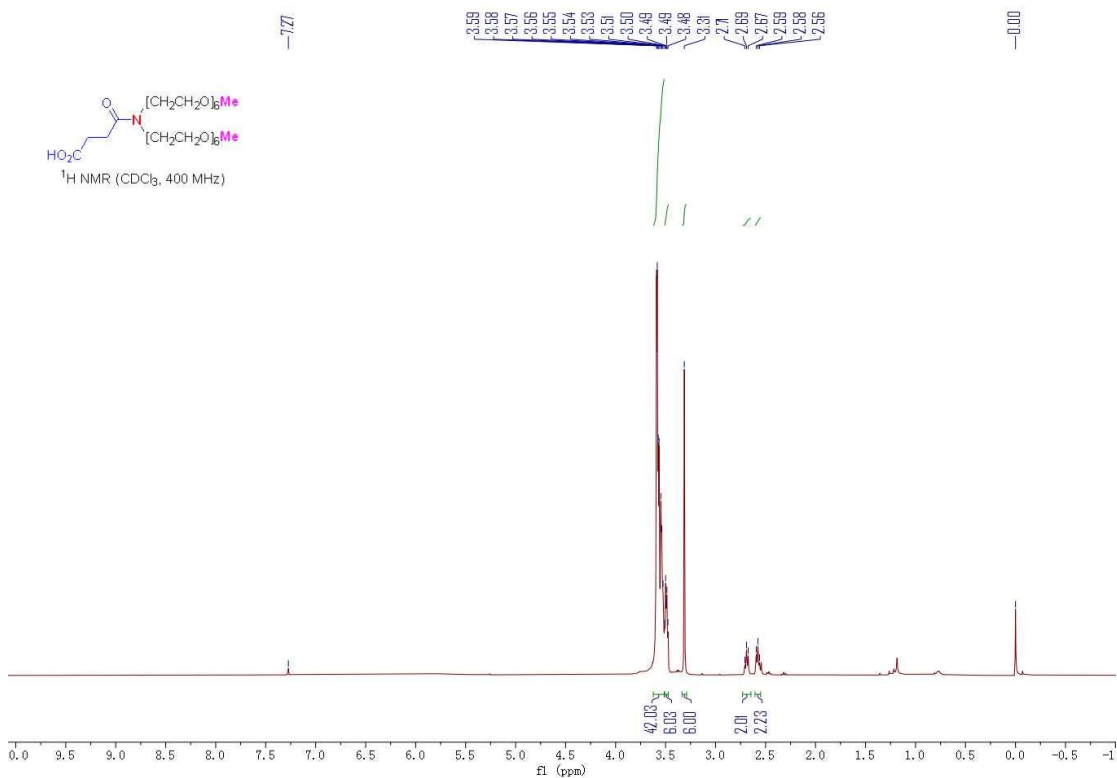


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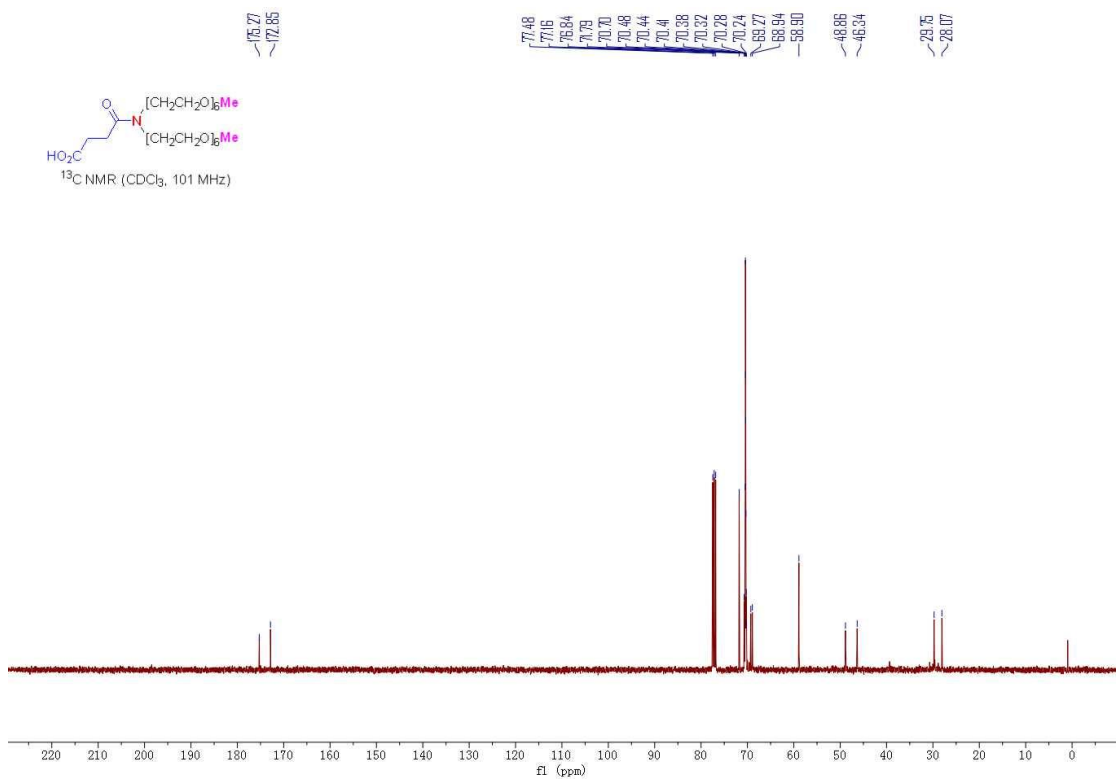
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¹H NMR of compound 24

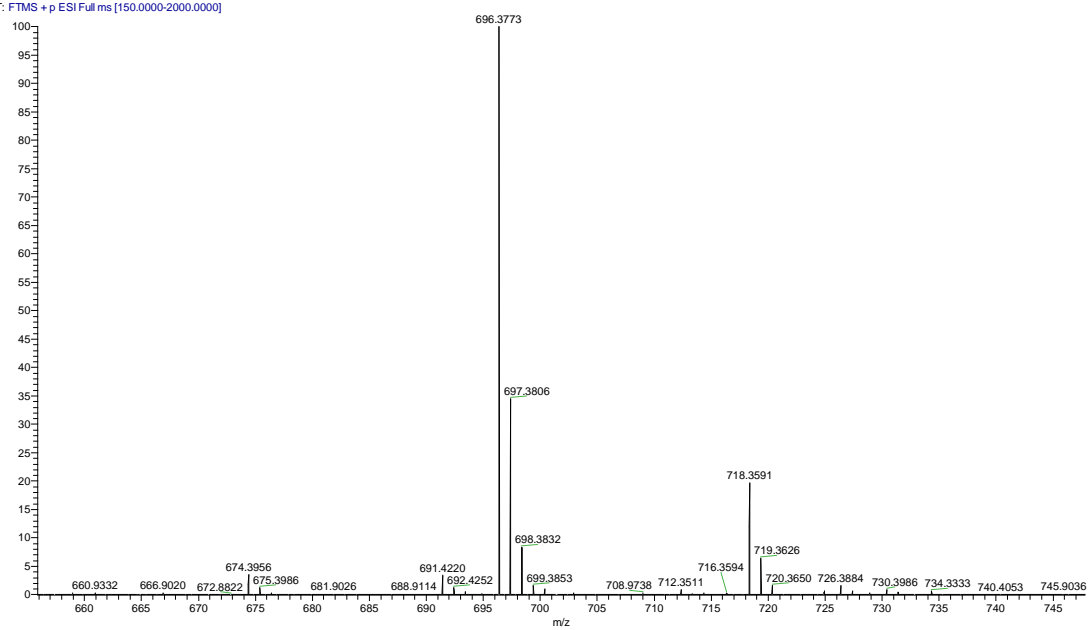


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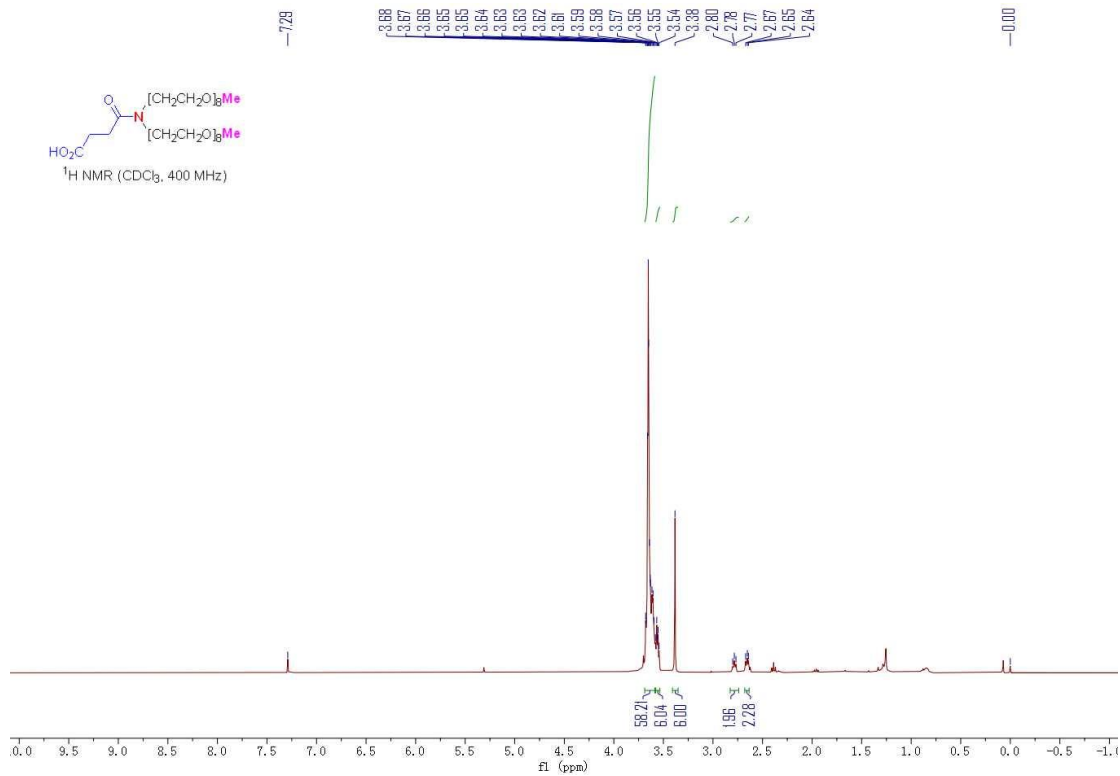
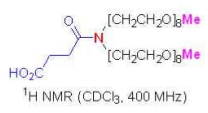


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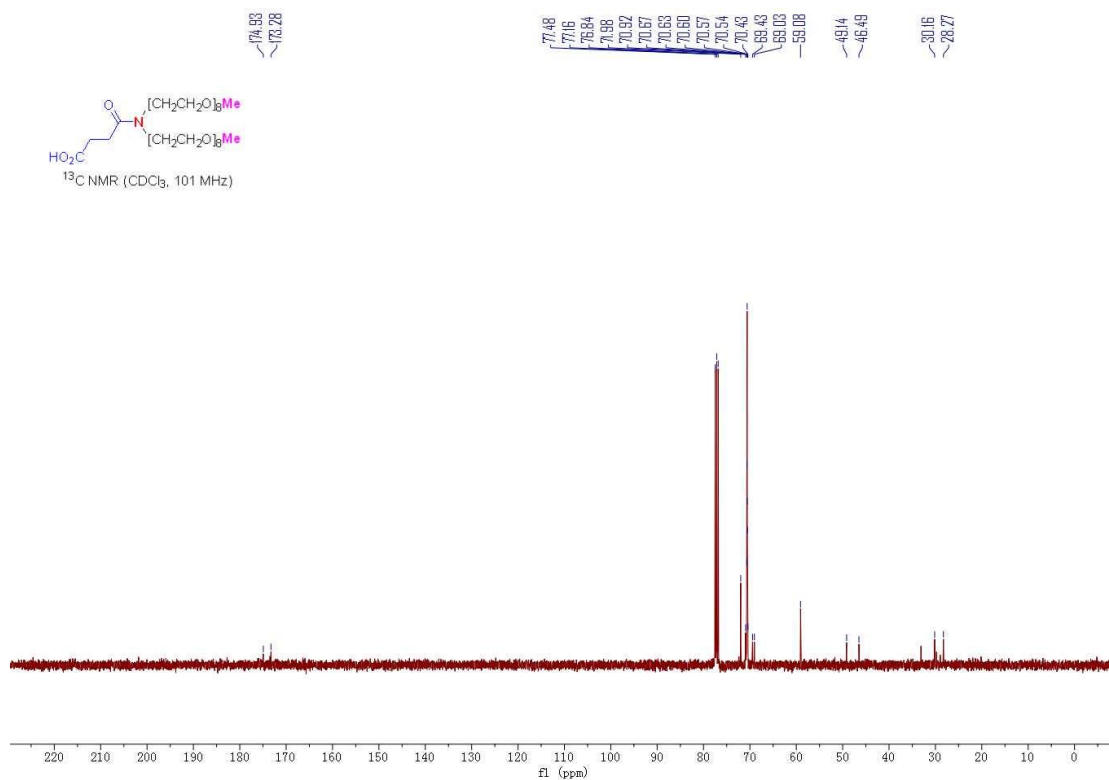
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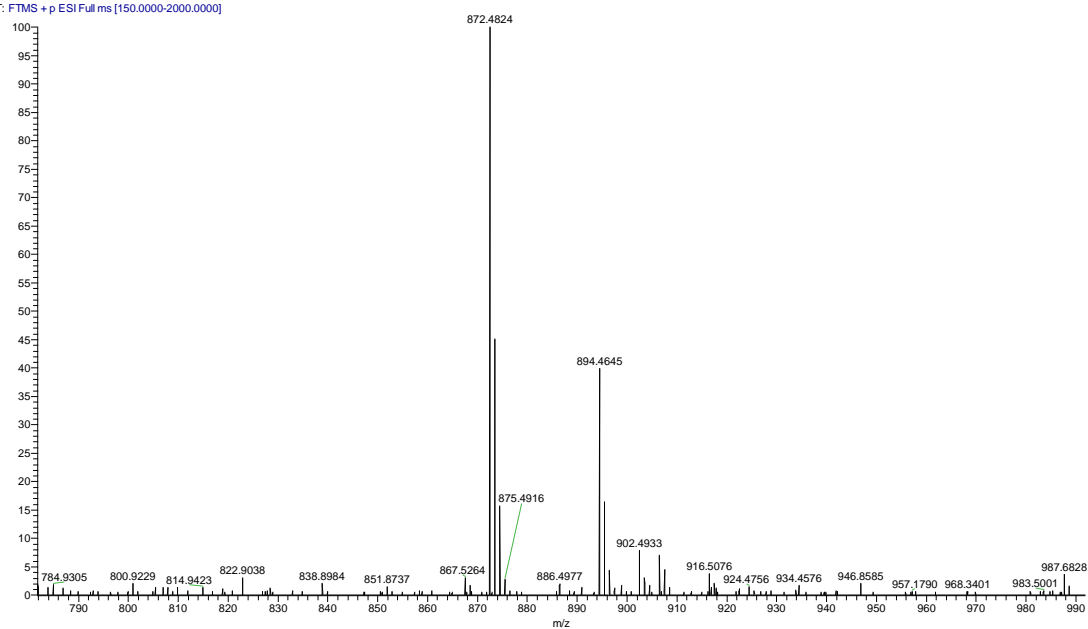


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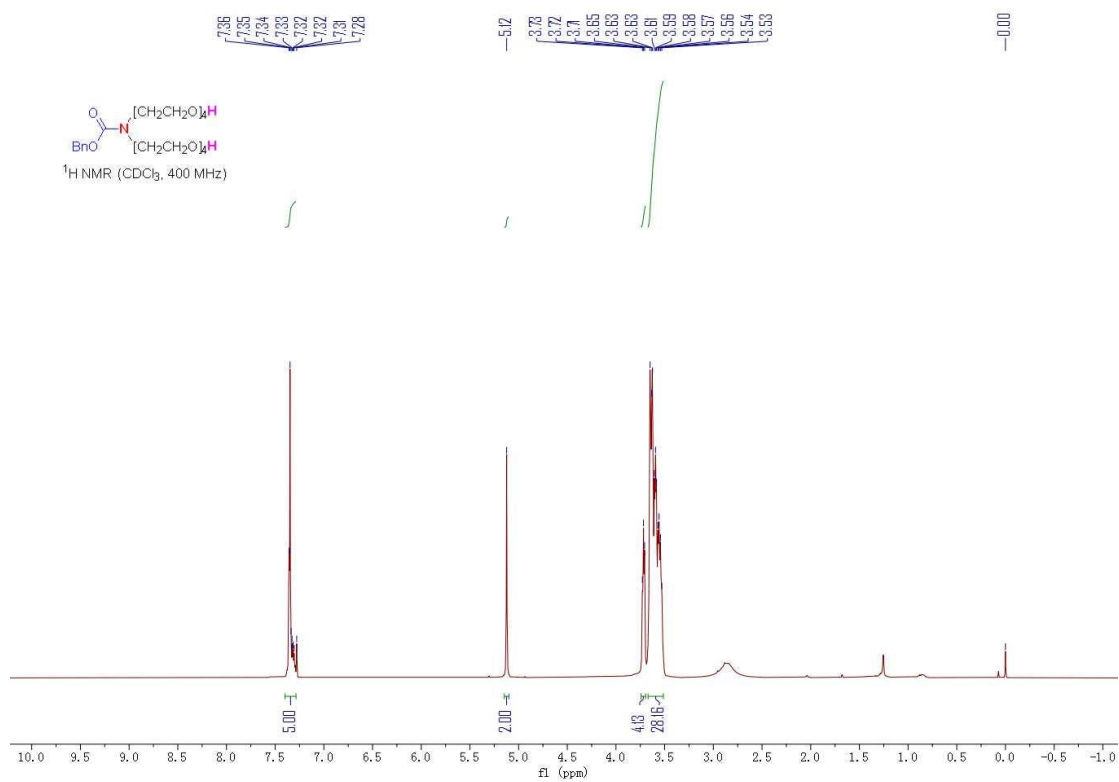


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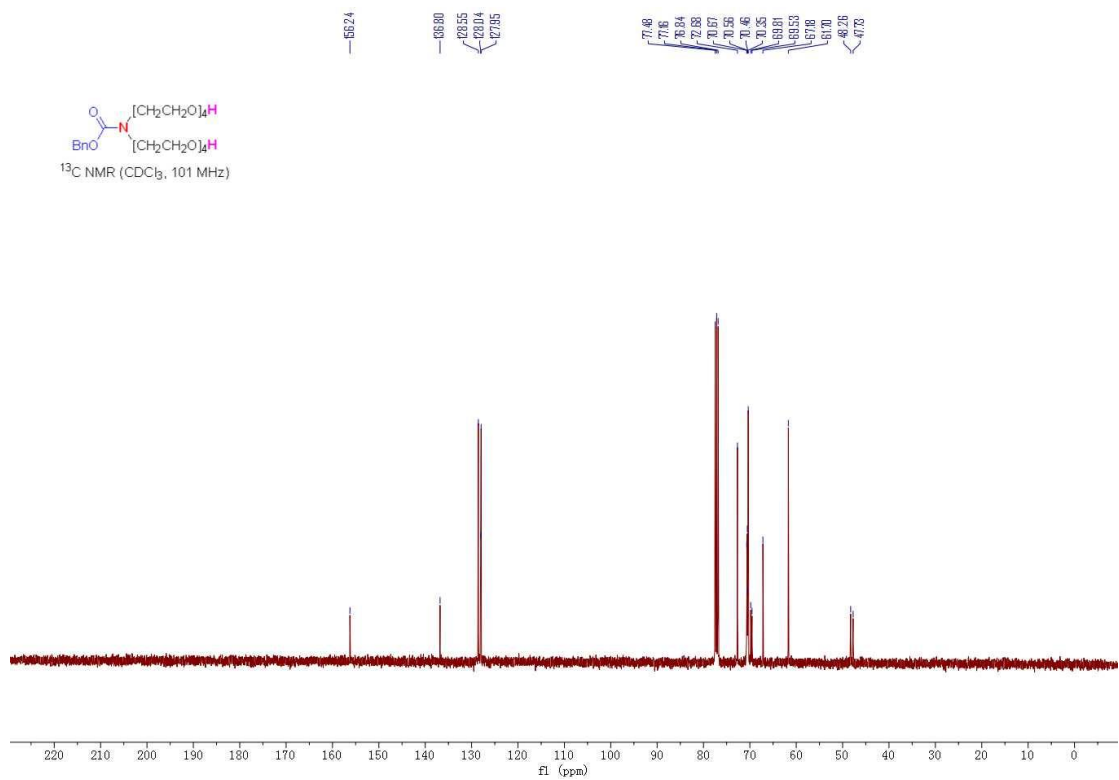
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¹H NMR of compound 26

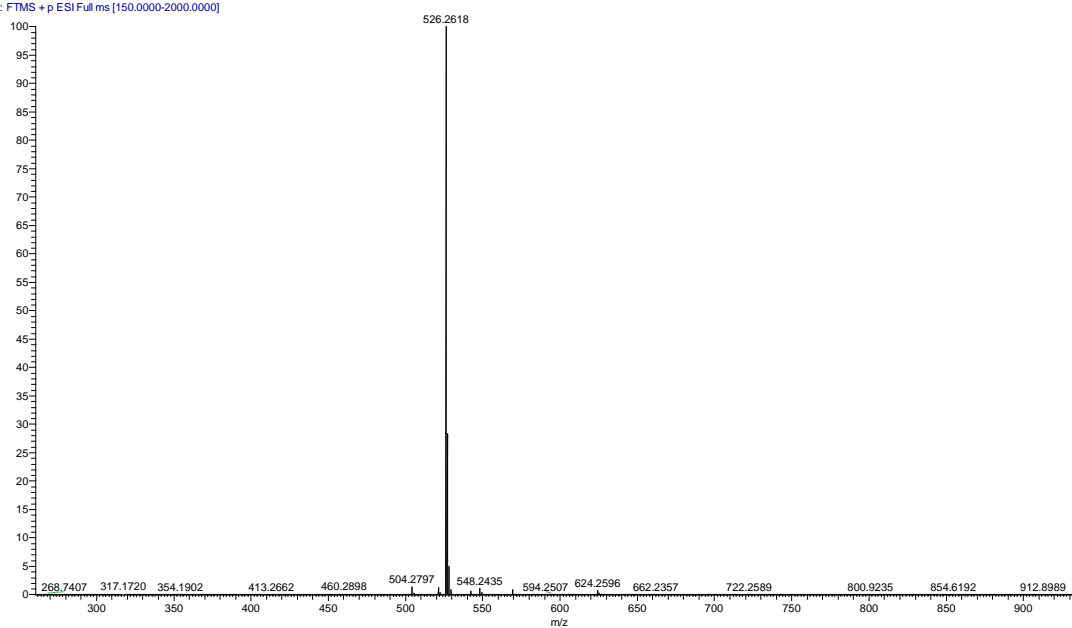


¹³C NMR of compound 26

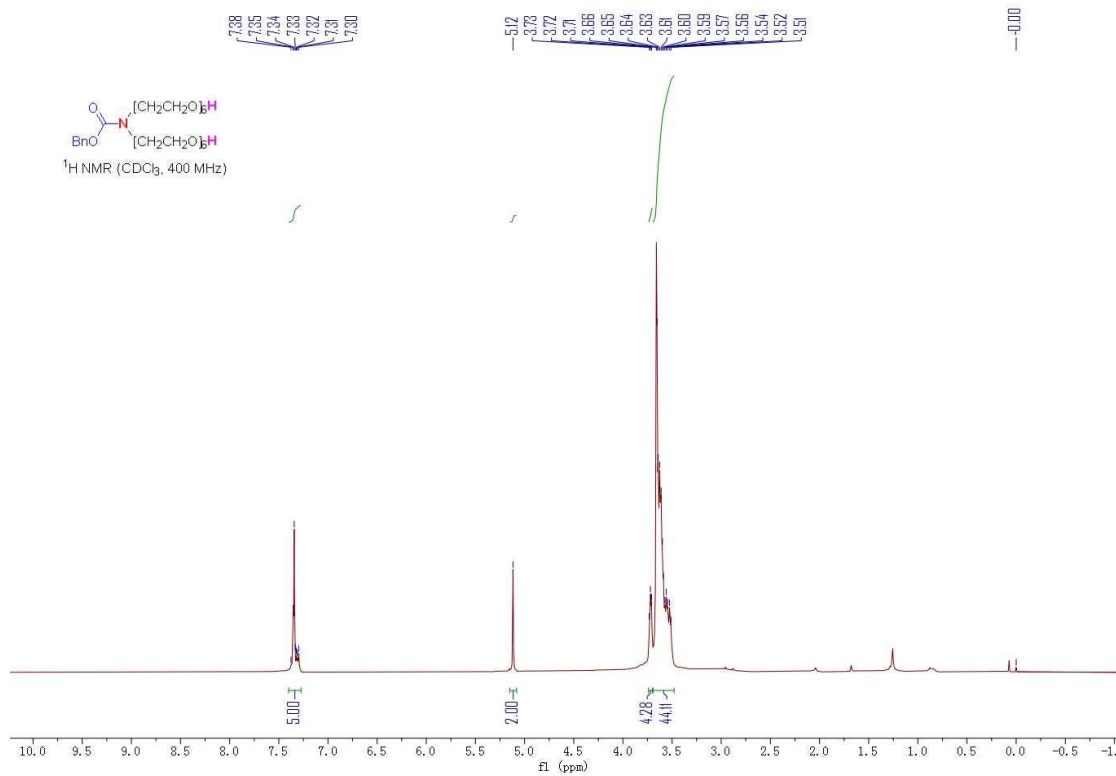


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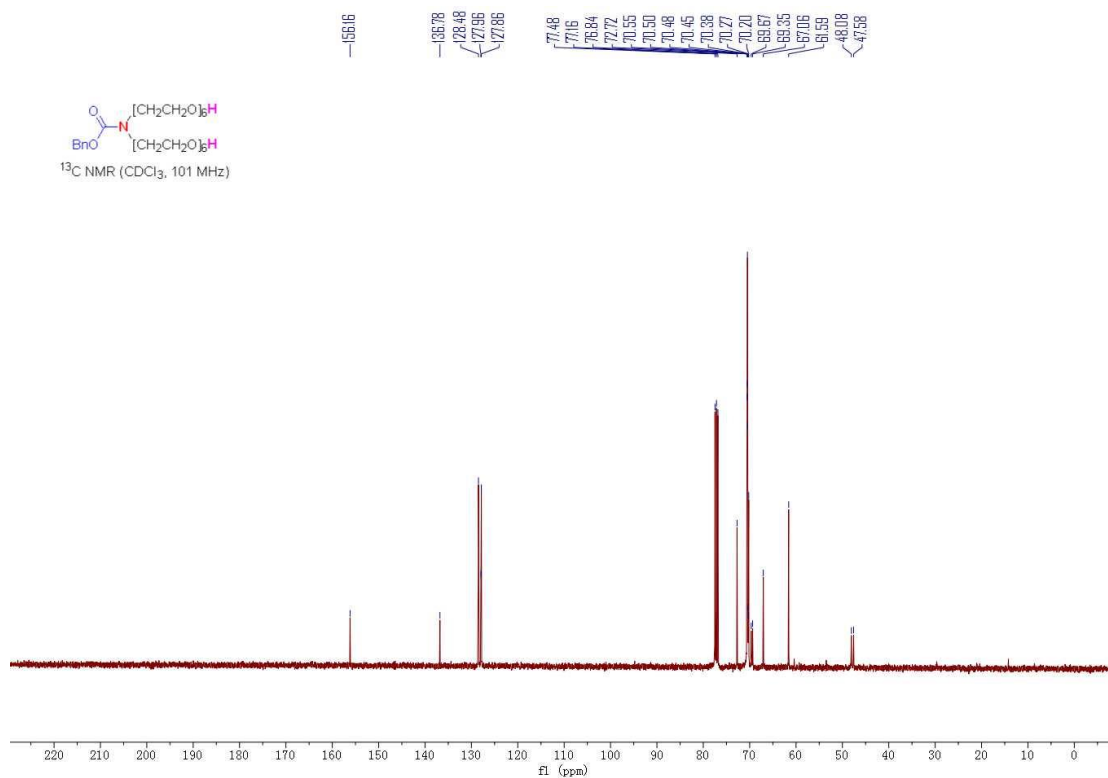
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T: FTMS + p ESI Full ms [150.0000-2000.0000]



¹H NMR of compound 27

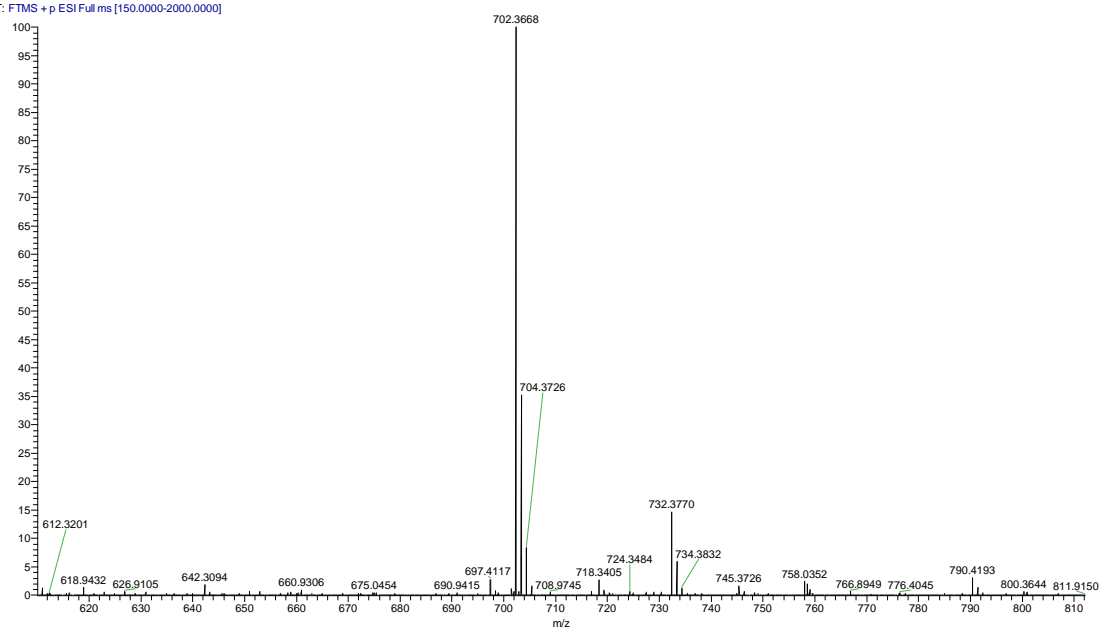


¹³C NMR of compound 27

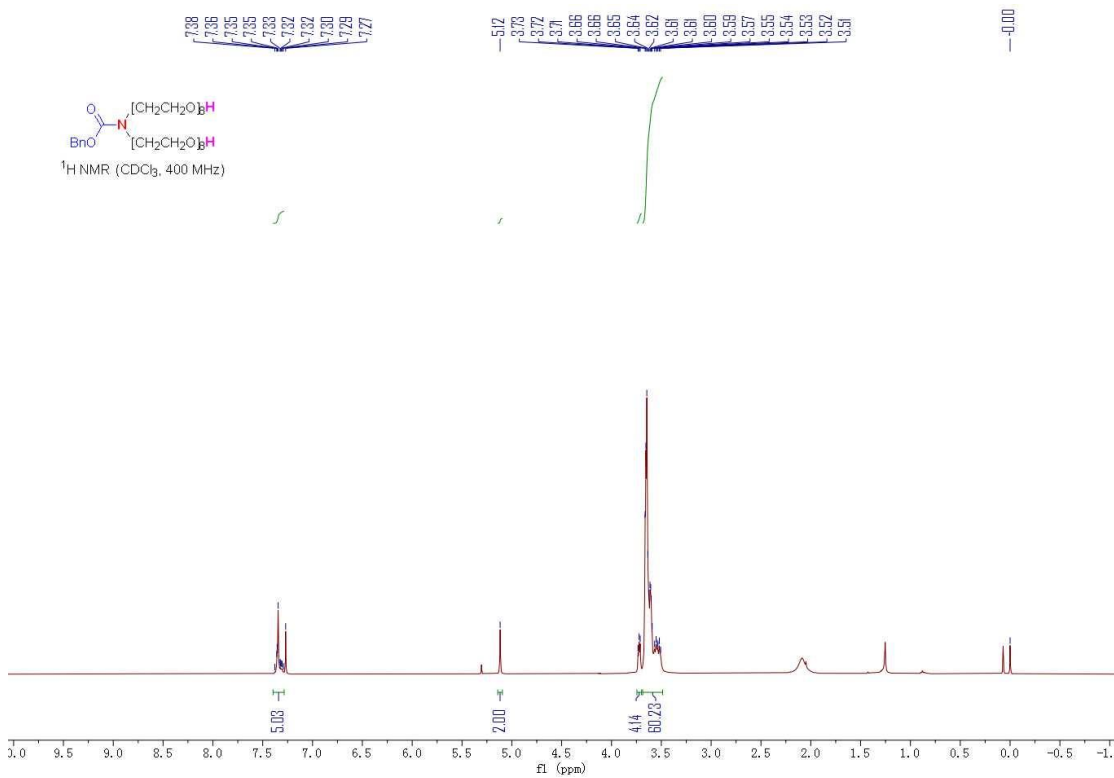


HRMS of compound 27

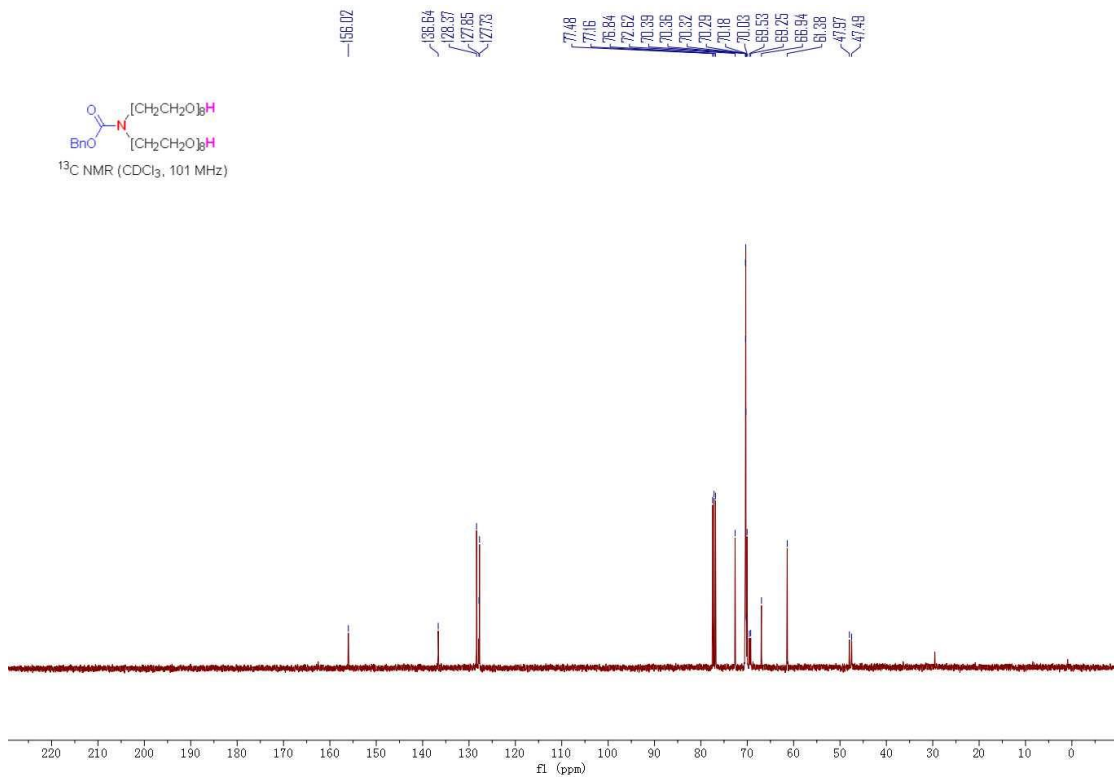
8 #715 RT: 7.62 AV: 1 NL: 1.16E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



¹H NMR of compound 28

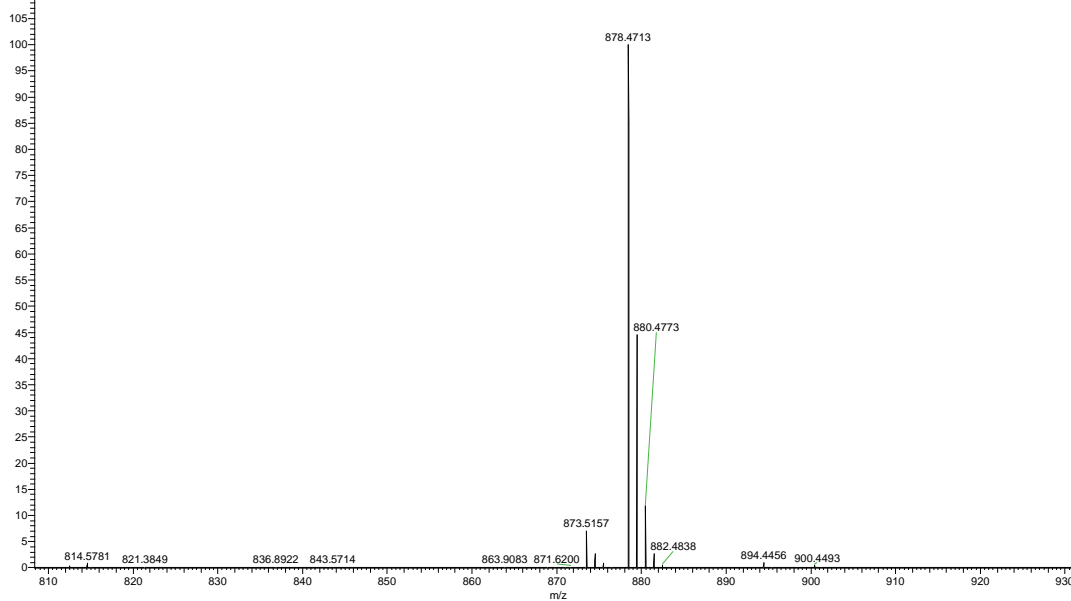


¹³C NMR of compound 28

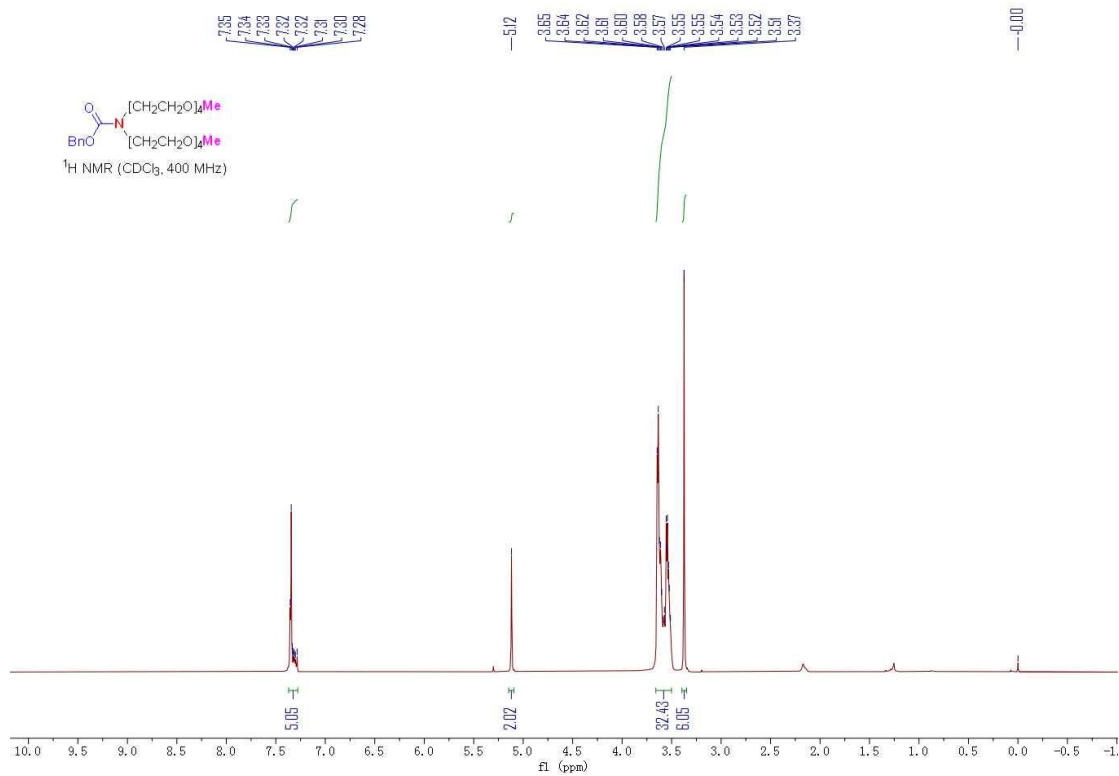


HRMS of compound 28

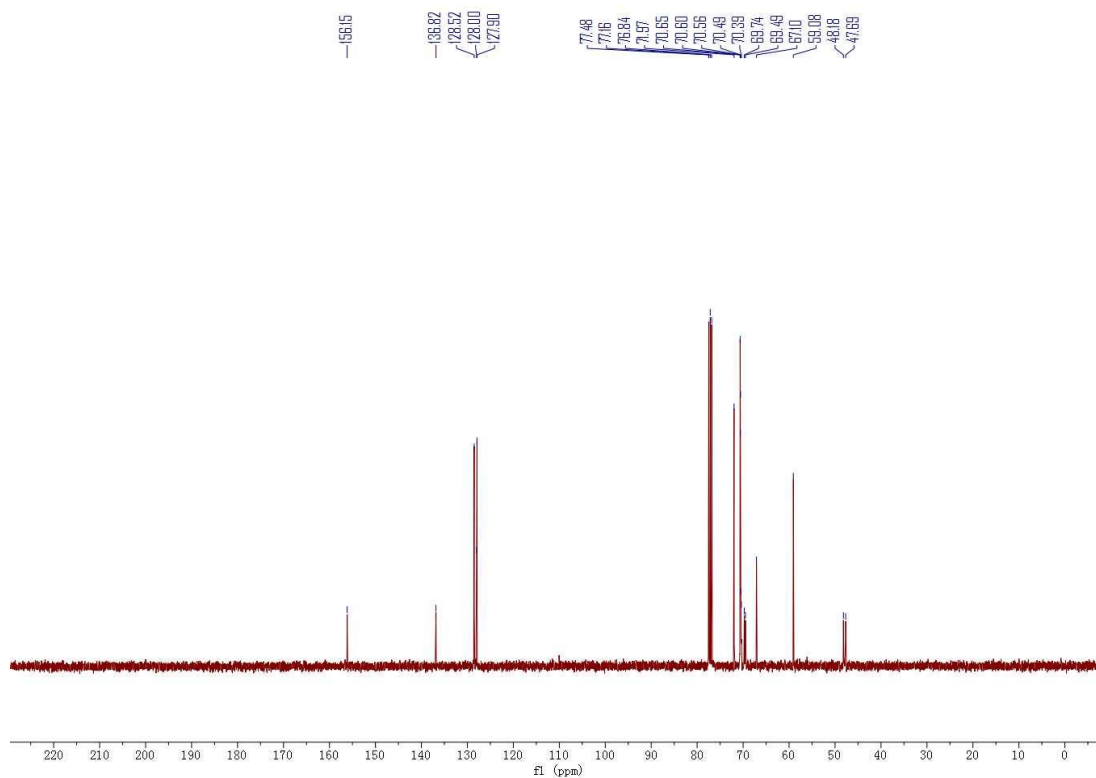
2 #717 RT: 7.77 AV: 1 NL: 2.94E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



¹H NMR of compound 29

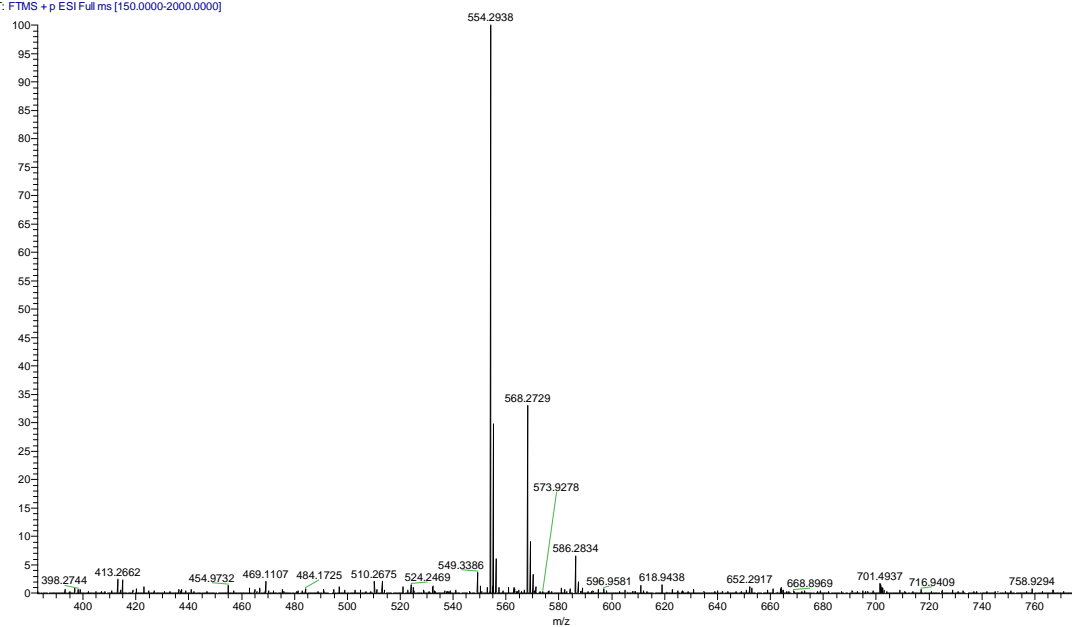


¹³C NMR of compound **29**

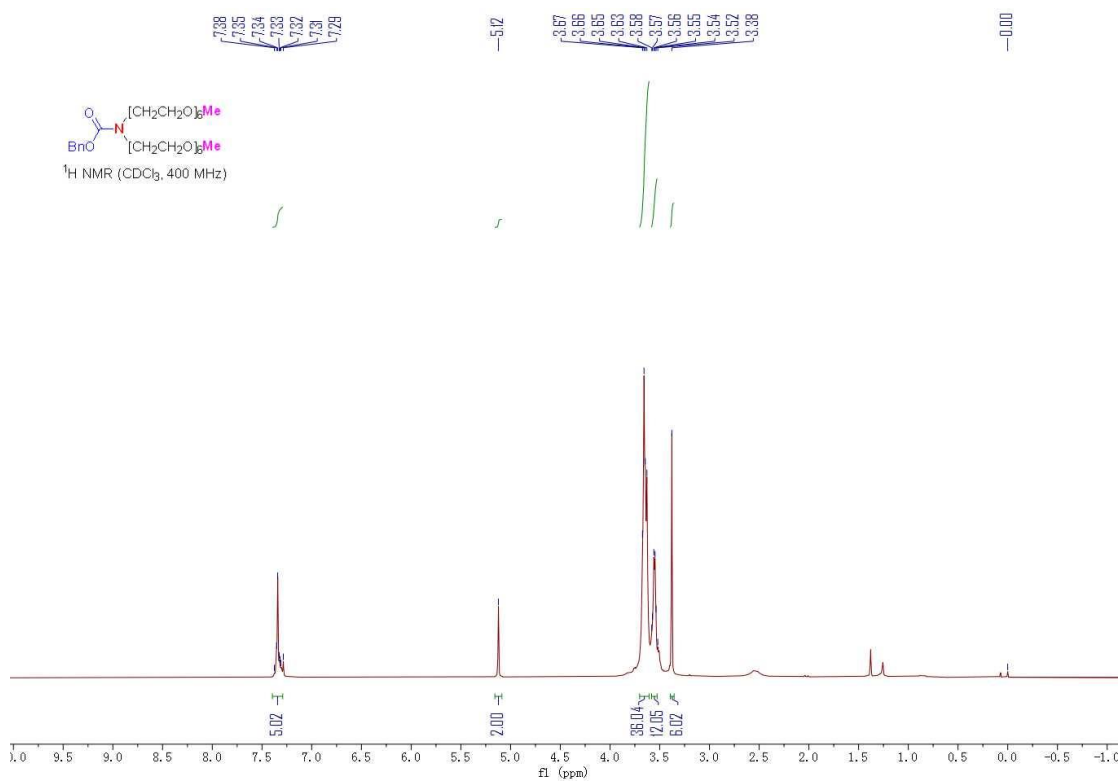


HRMS of compound **29**

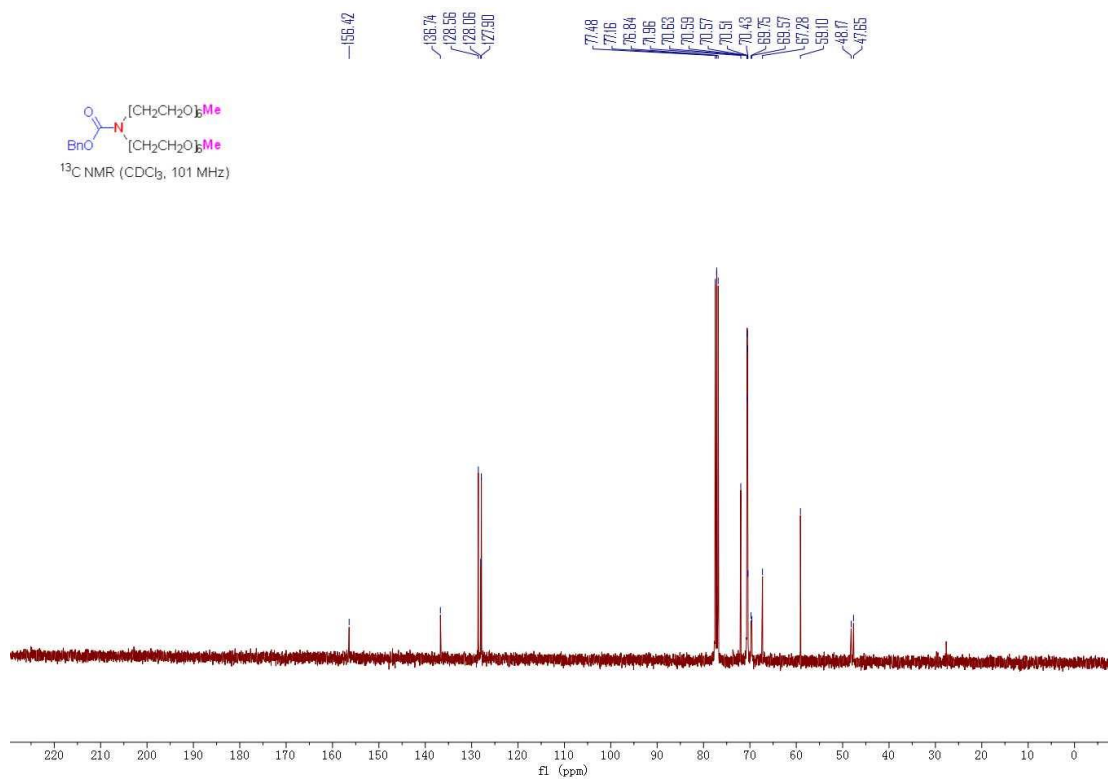
17 #759 RT: 7.96 AV: 1 NL: 1.29E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



^1H NMR of compound **30**

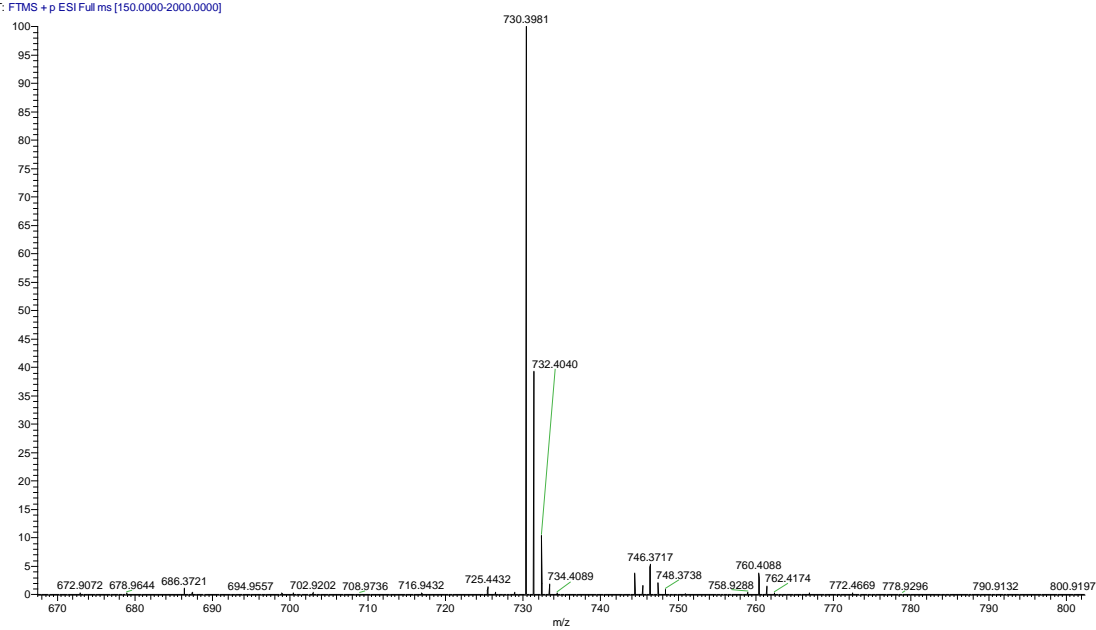


^{13}C NMR of compound **30**

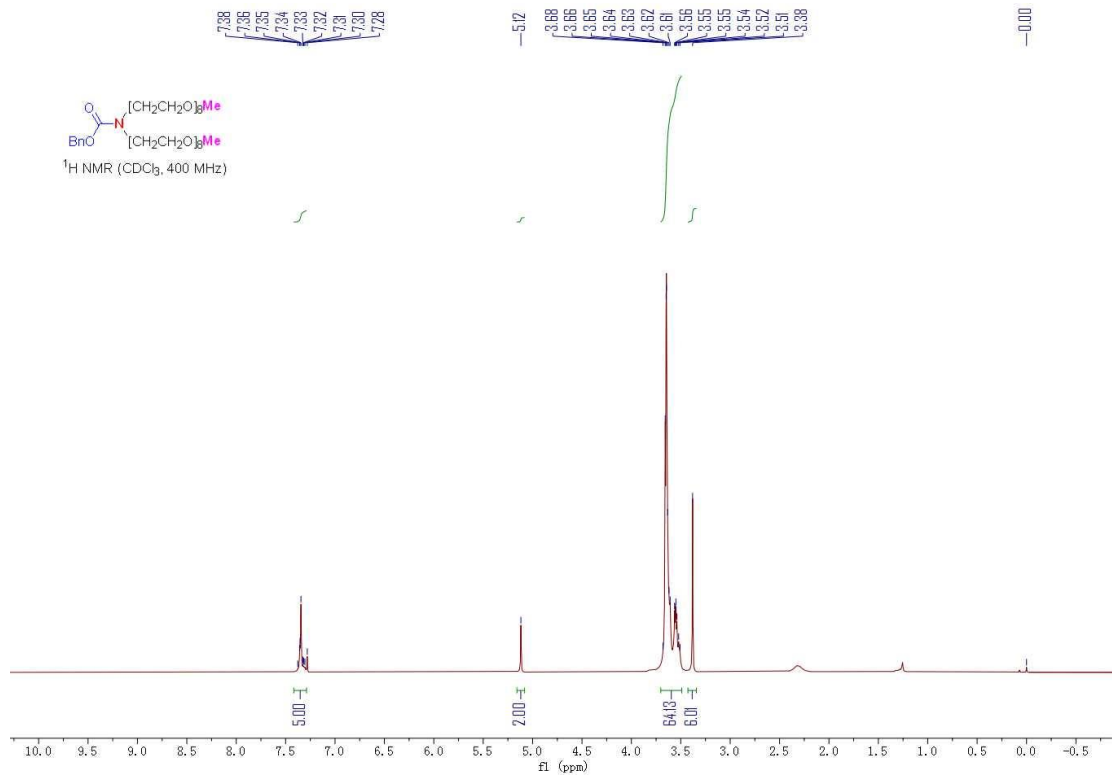


HRMS of compound 30

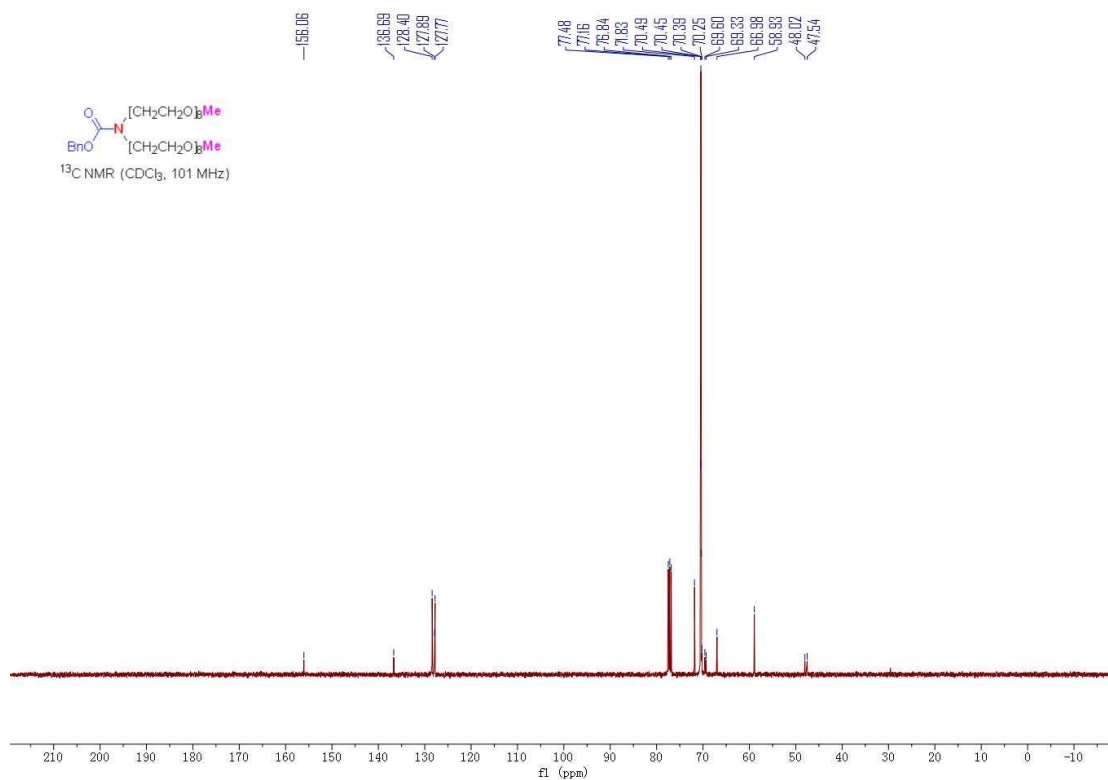
10 #793 RT: 8.40 AV: 1 NL: 2.36E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



¹H NMR of compound 31

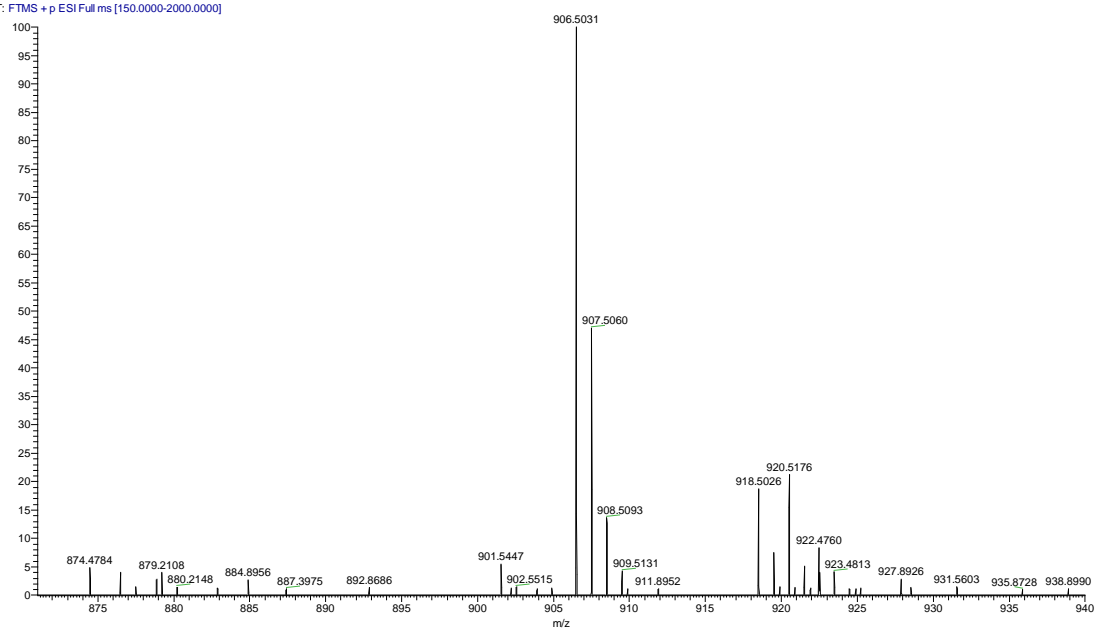


¹³C NMR of compound **31**

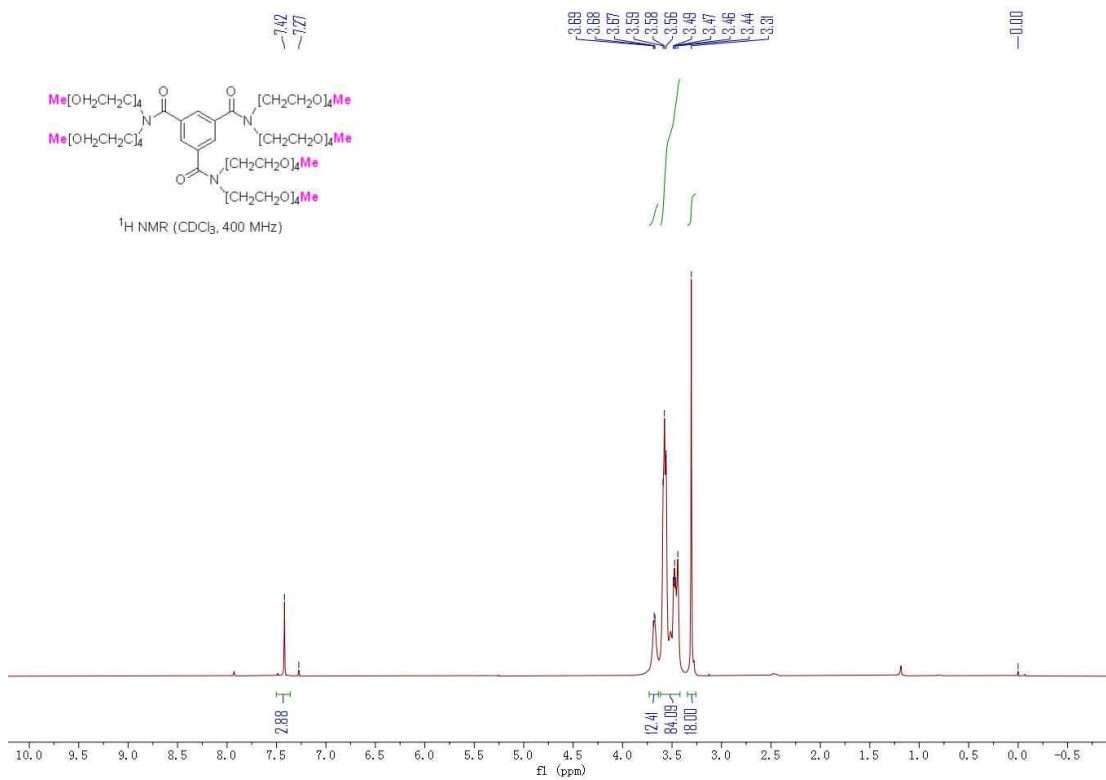


HRMS of compound **31**

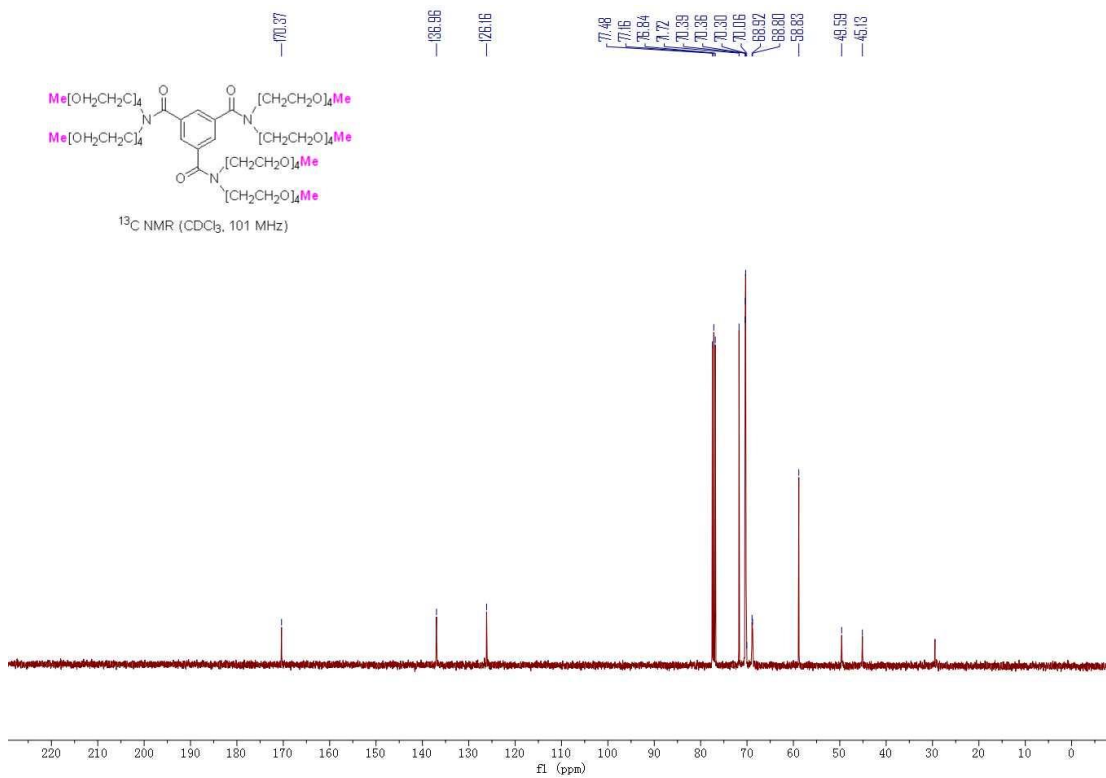
5 #799 RT: 8.59 AV: 1 NL: 1.64E6
T: FTMS + p ESI Full ms [150.0000-2000.0000]



¹H NMR of compound 32

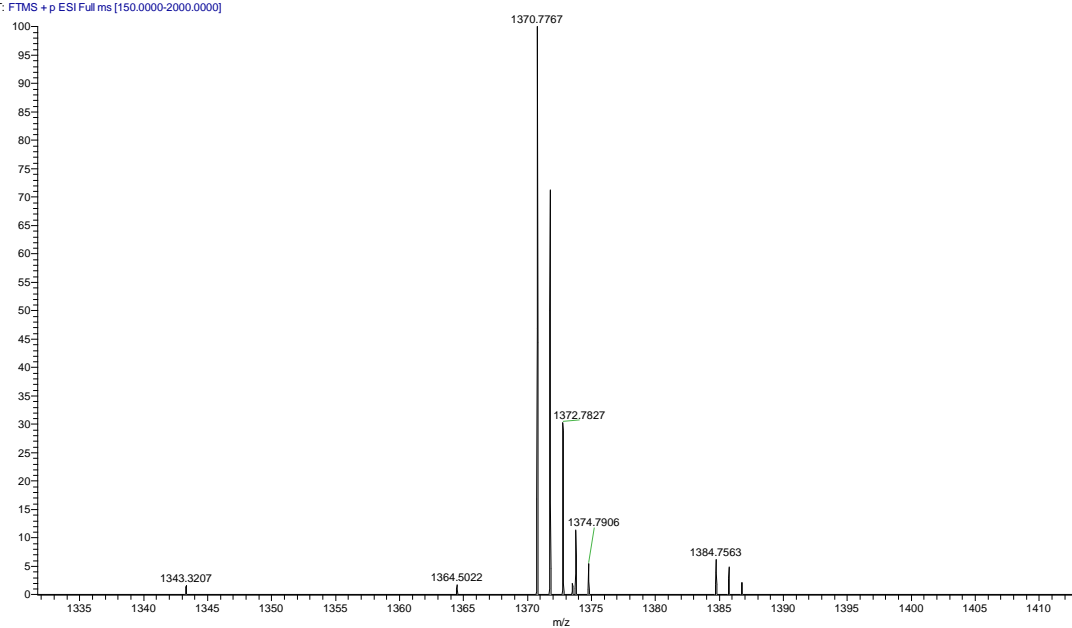


¹³C NMR of compound 32

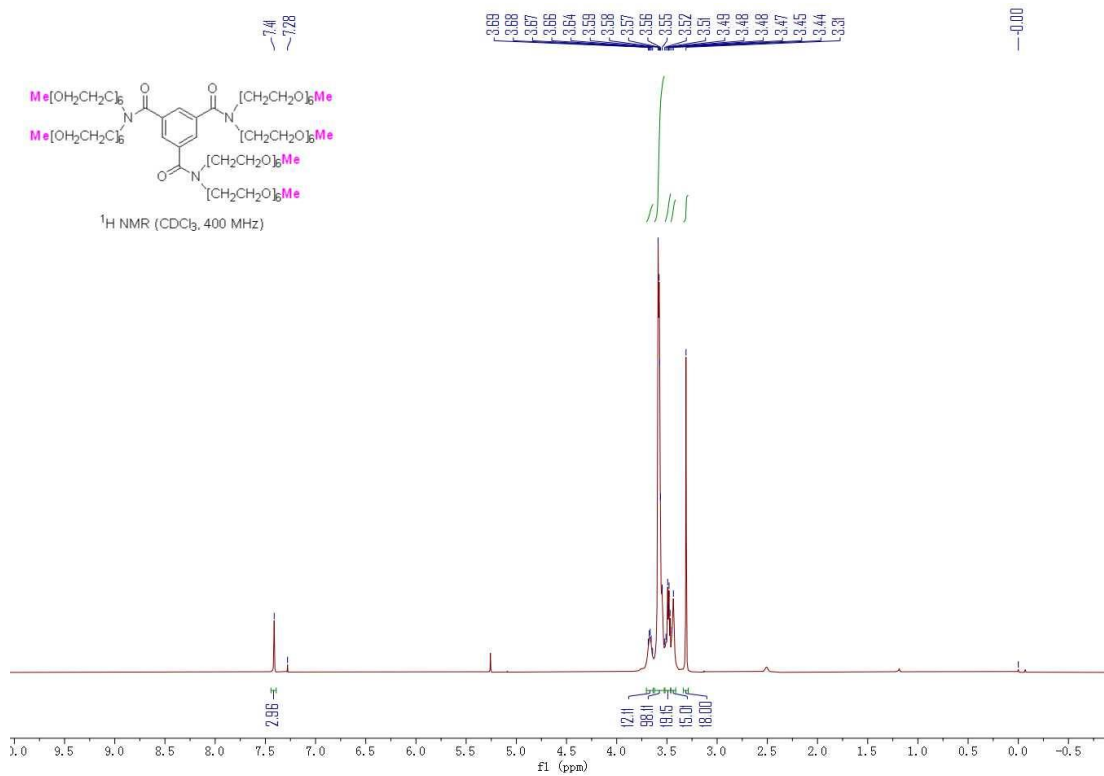


HRMS of compound 32

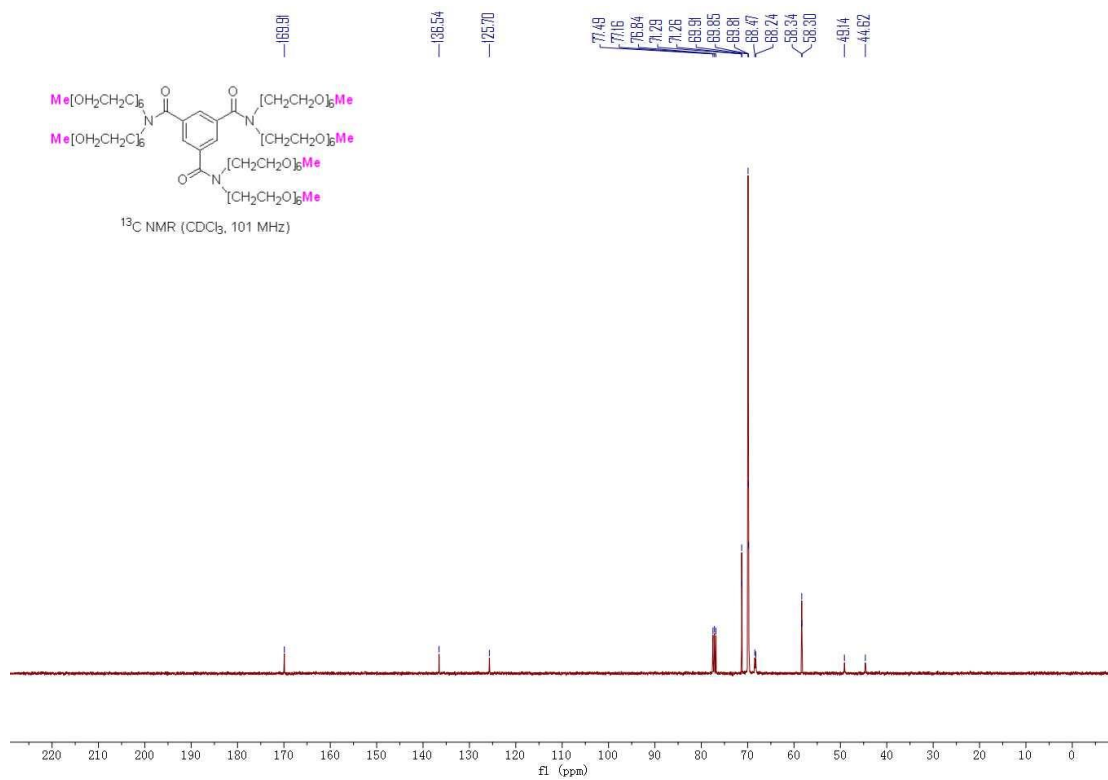
20 #745 RT: 7.71 AV: 1 NL: 1.49E6
T: FTMS + p ESI Full ms [150.0000-2000.0000]



¹H NMR of compound 33

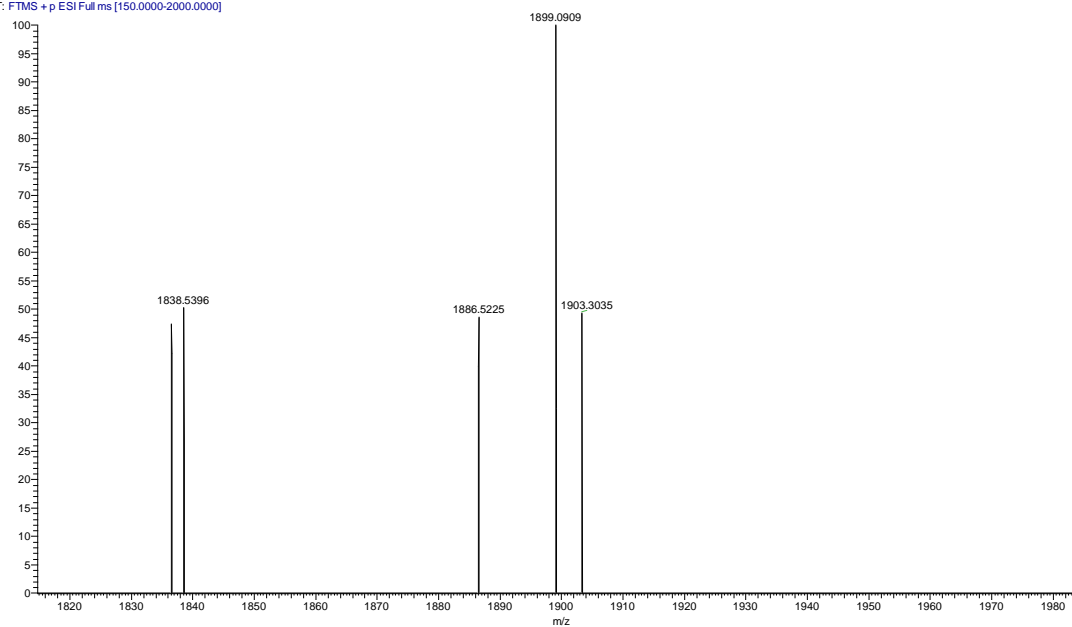


¹³C NMR of compound **33**

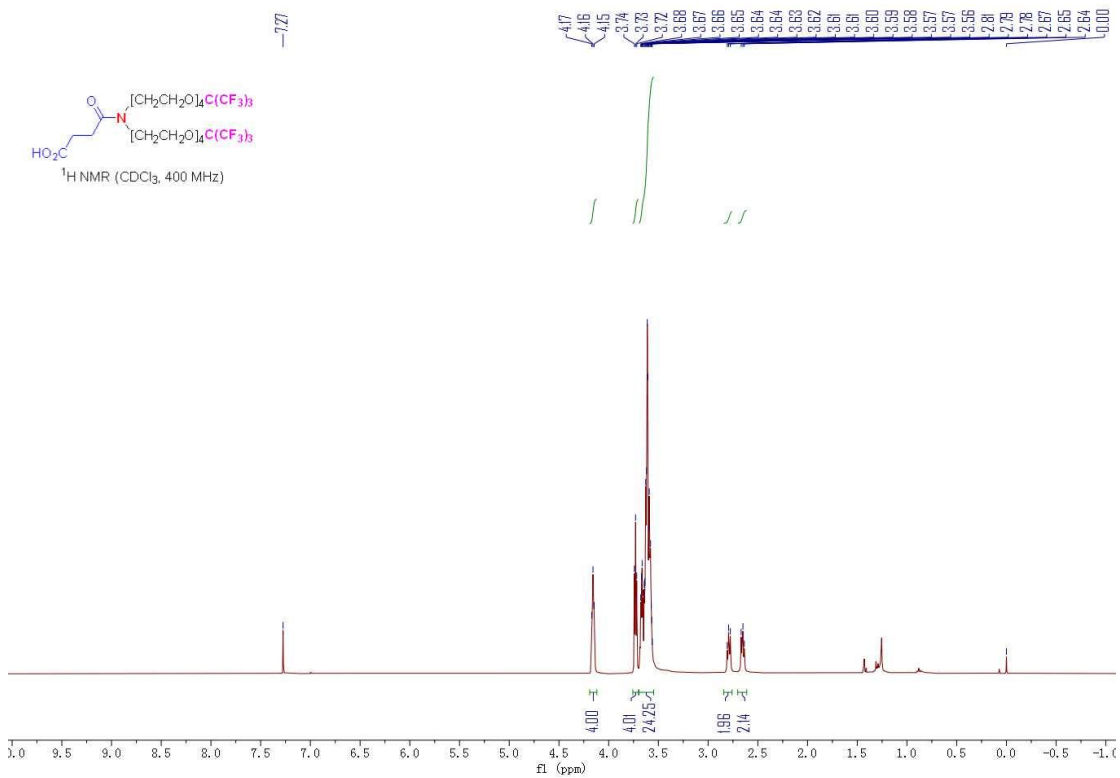


HRMS of compound **33**

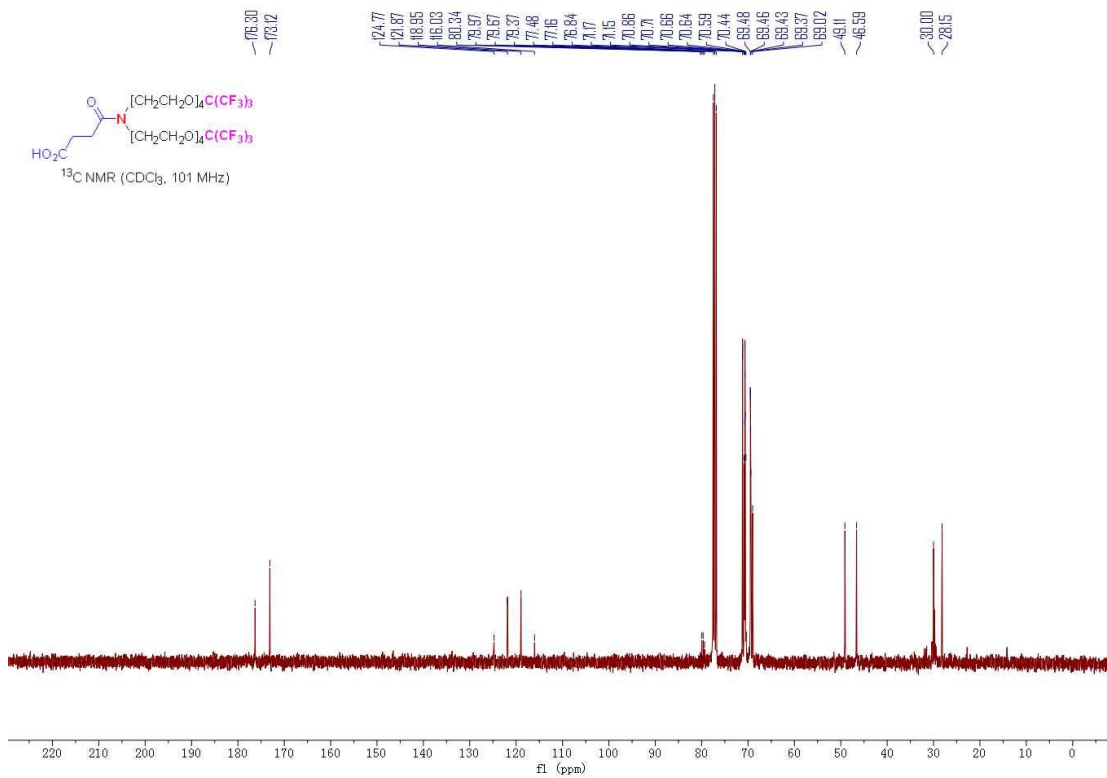
13 #1427 RT: 14.73 AV: 1 NL: 2.94E4
T: FTMS + p ESI Full ms [150.0000-2000.0000]



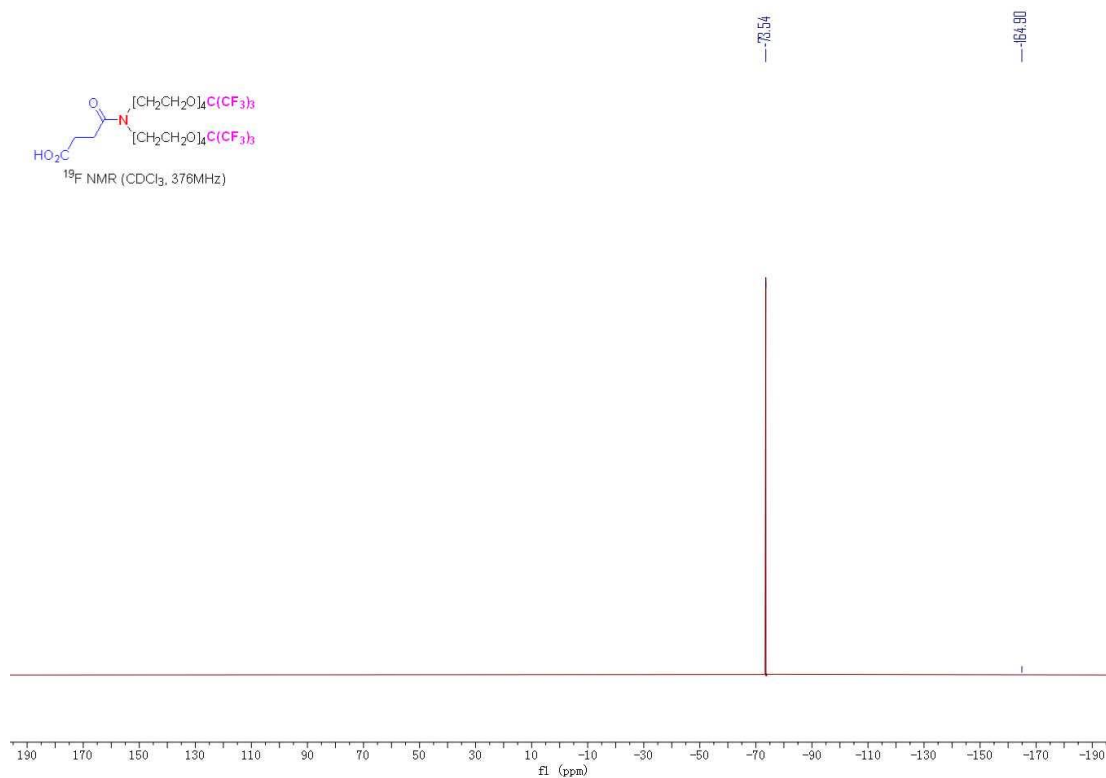
¹H NMR of compound 34



¹³C NMR of compound 34

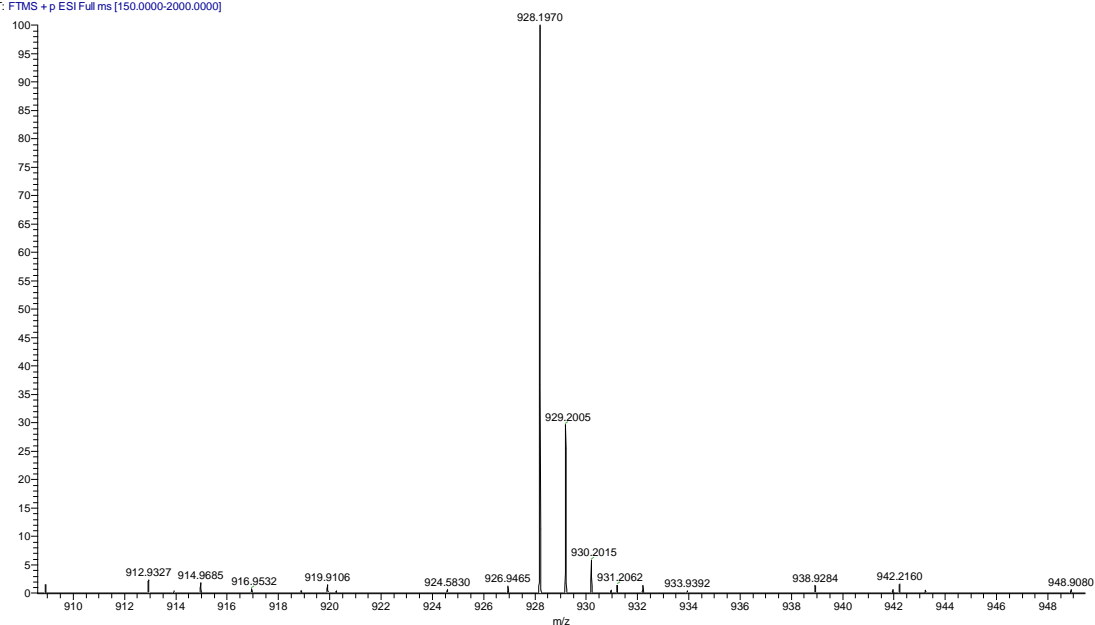


¹⁹F NMR of compound 34

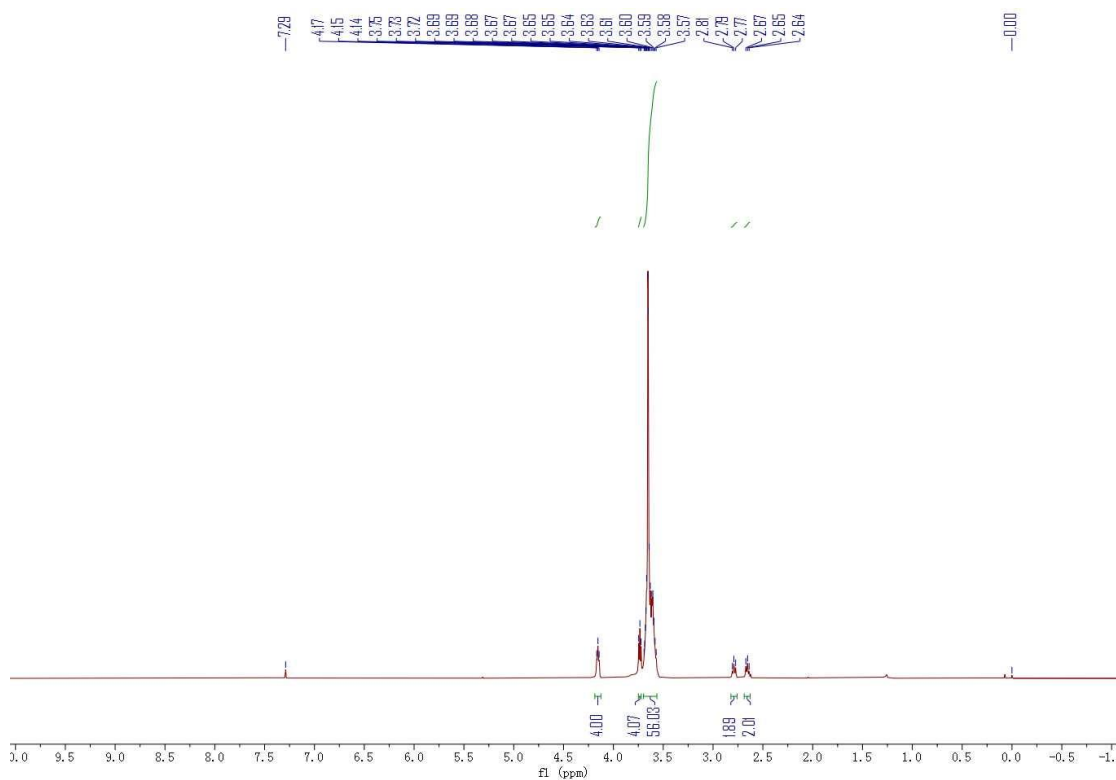


HRMS of compound 34

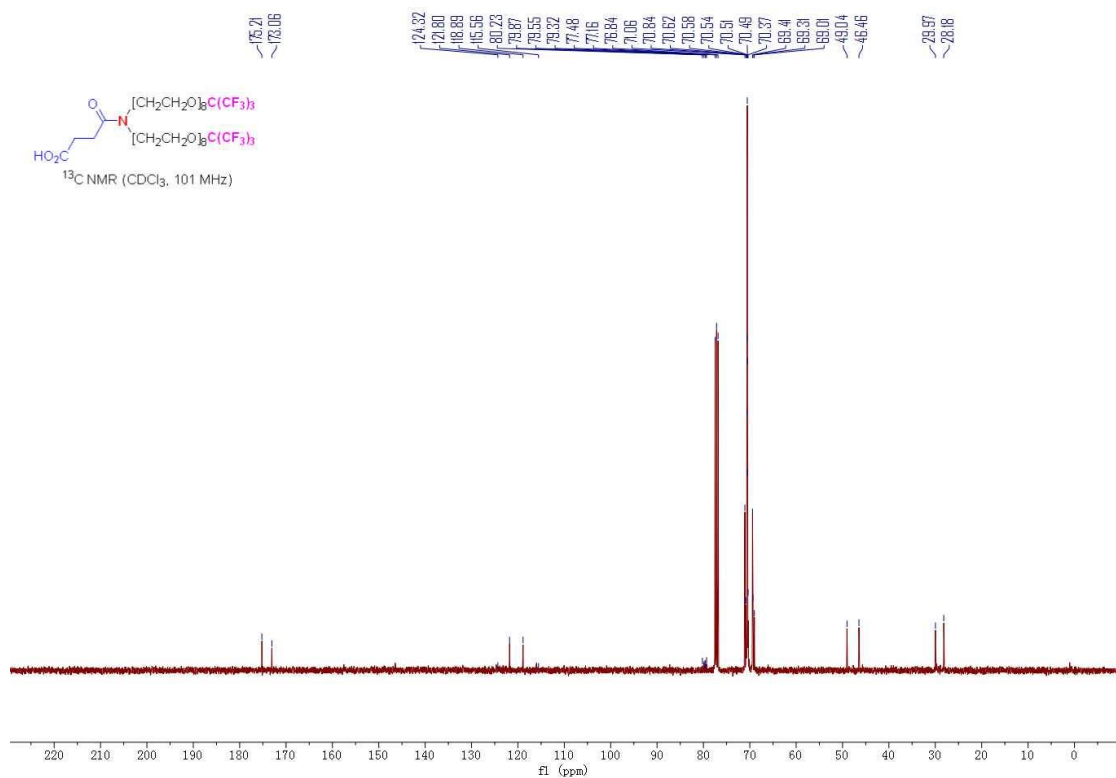
25 #955 RT: 9.60 AV: 1 NL: 5.72E6
T: FTMS + p ESI Full ms [150.0000-2000.0000]



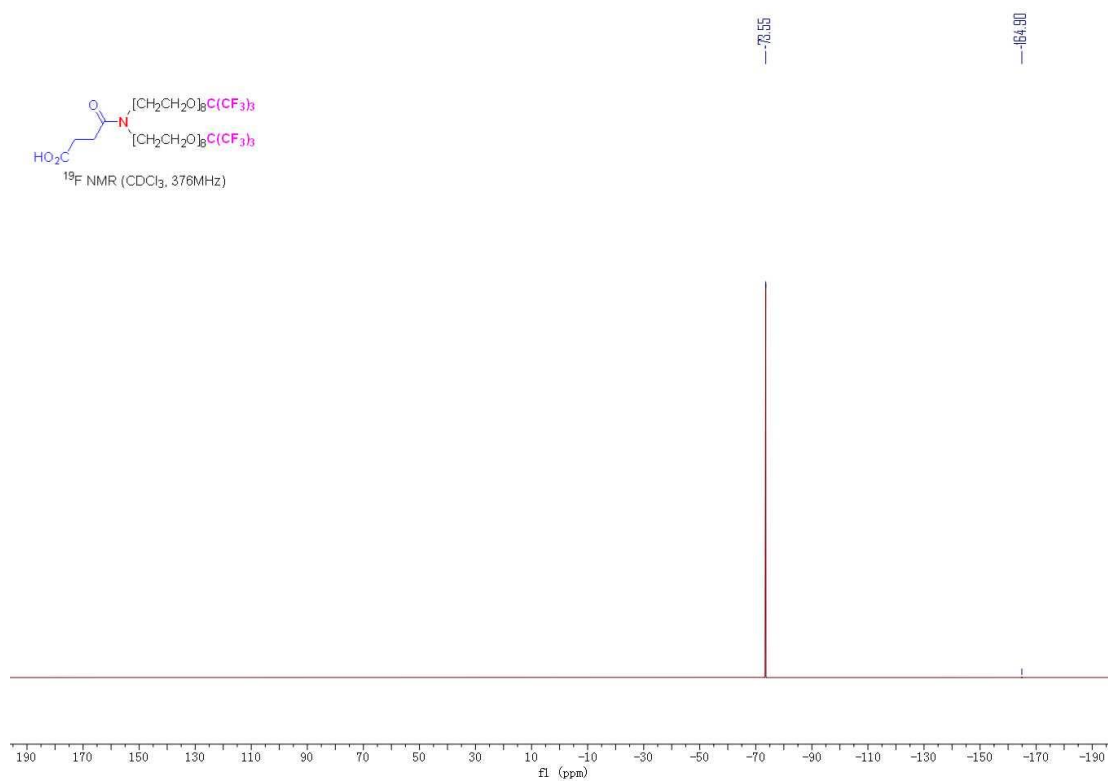
¹H NMR of compound 35



¹³C NMR of compound 35

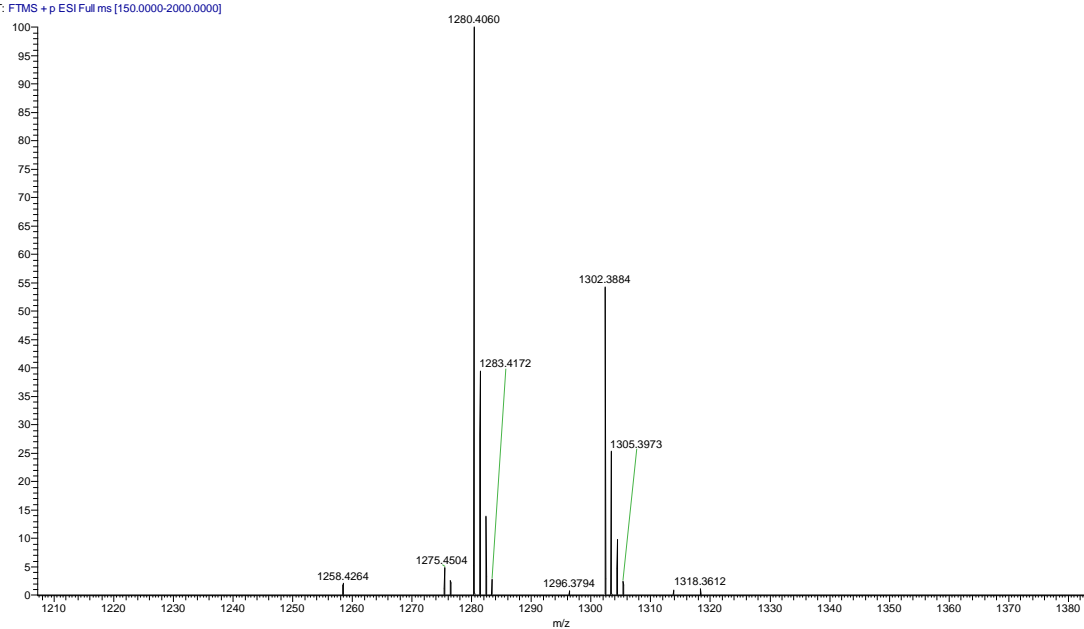


¹⁹F NMR of compound **35**

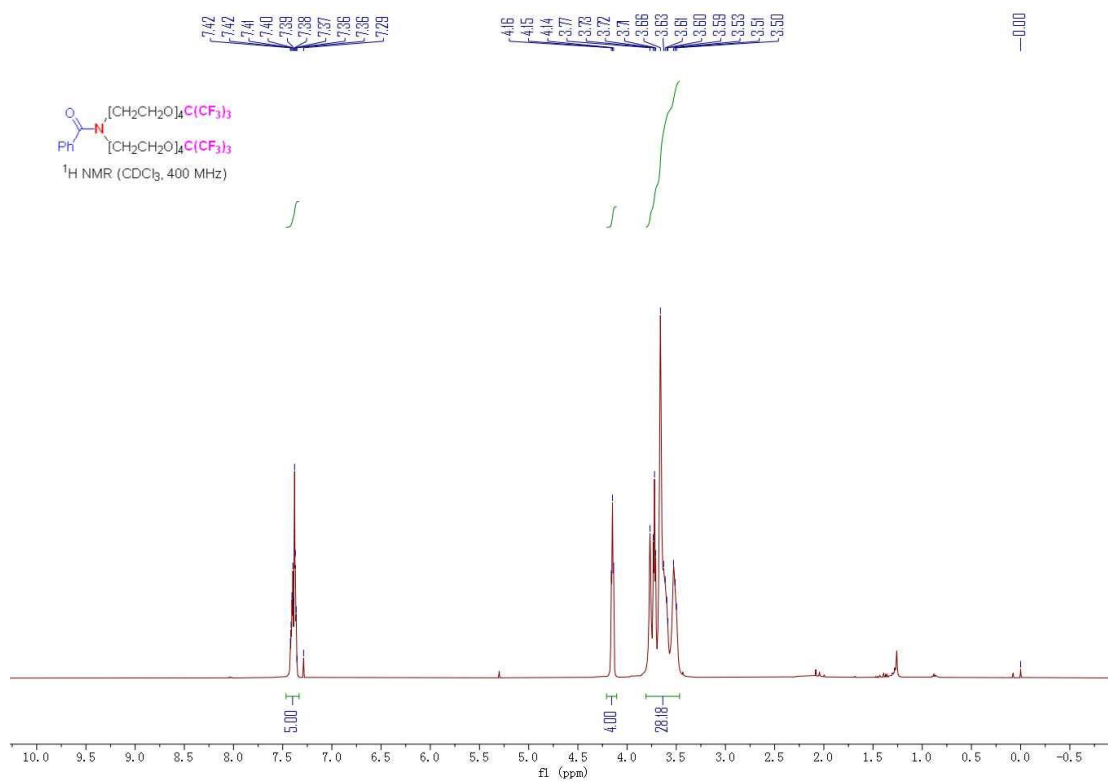


HRMS of compound **35**

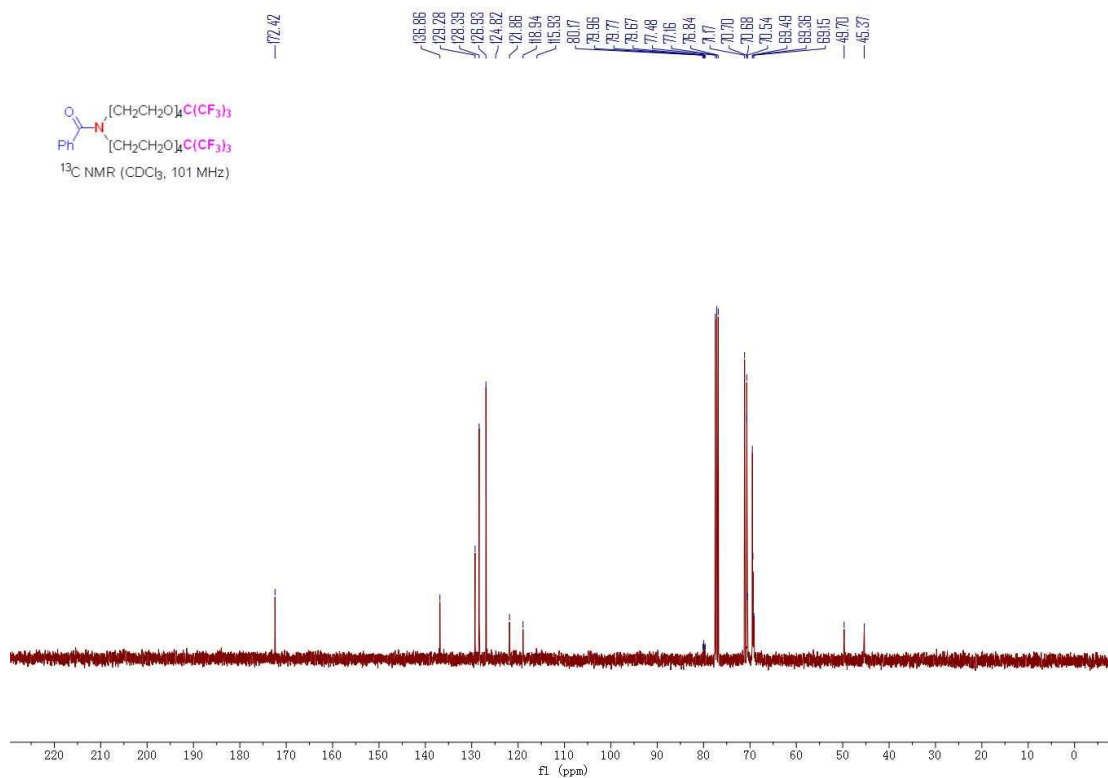
20 #947 RT: 9.51 AV: 1 NL: 4.50E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



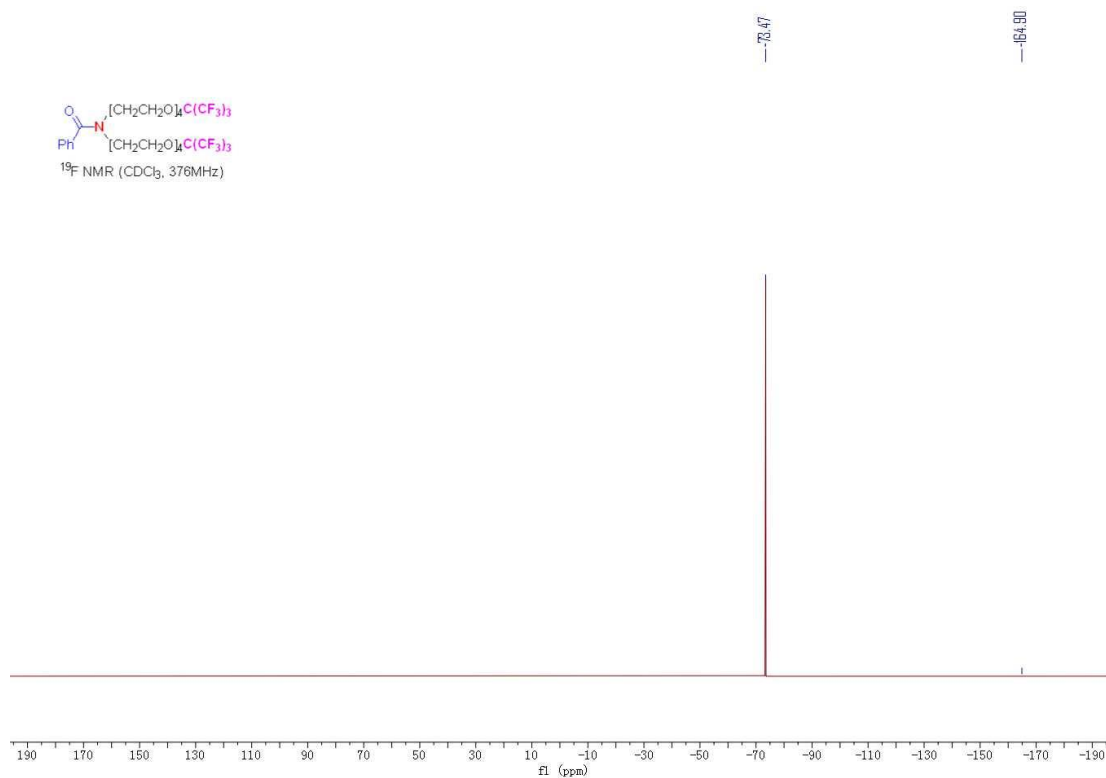
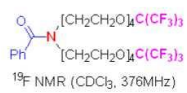
¹H NMR of compound 36



¹³C NMR of compound 36

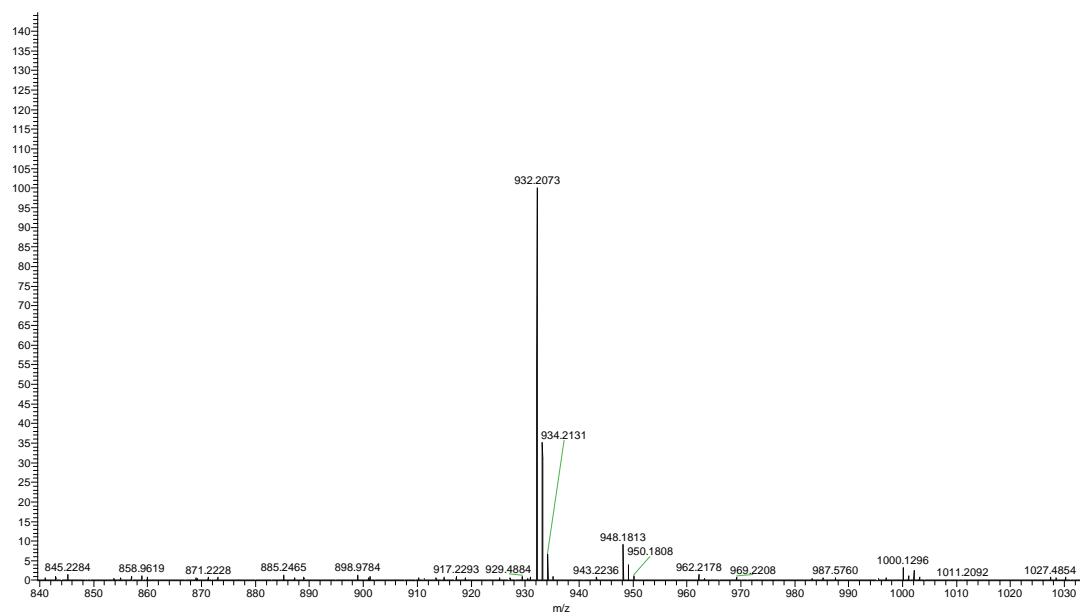


¹⁹F NMR of compound 36

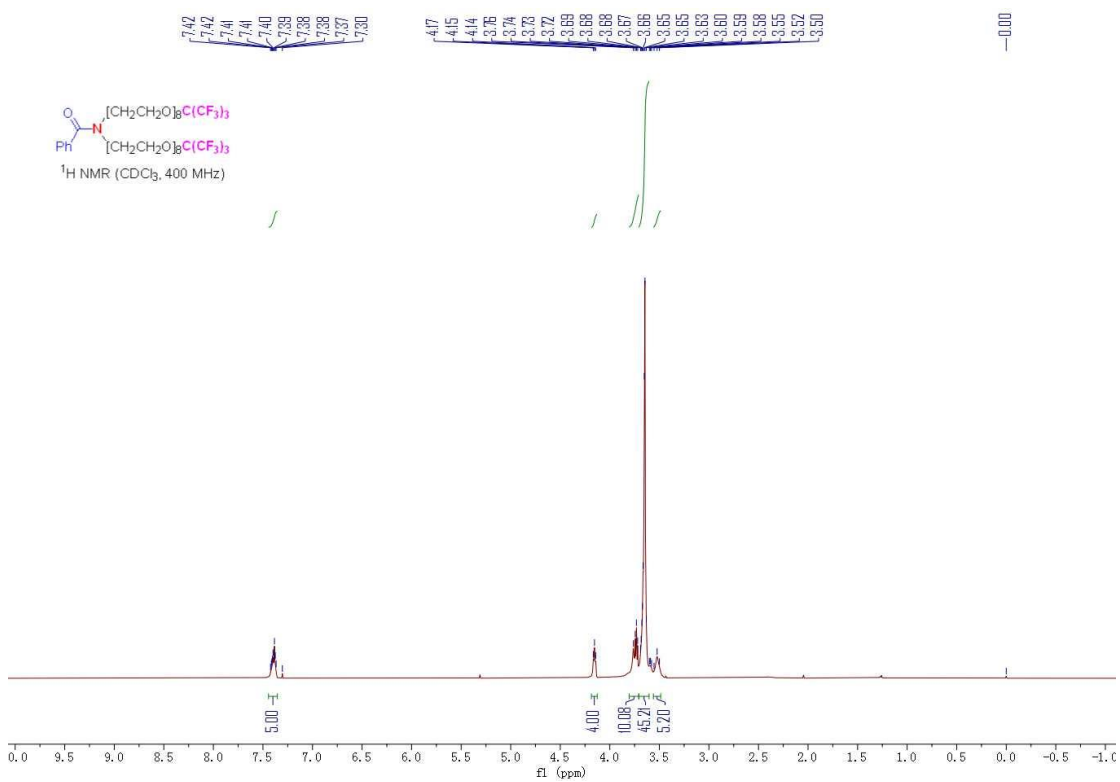


HRMS of compound 36

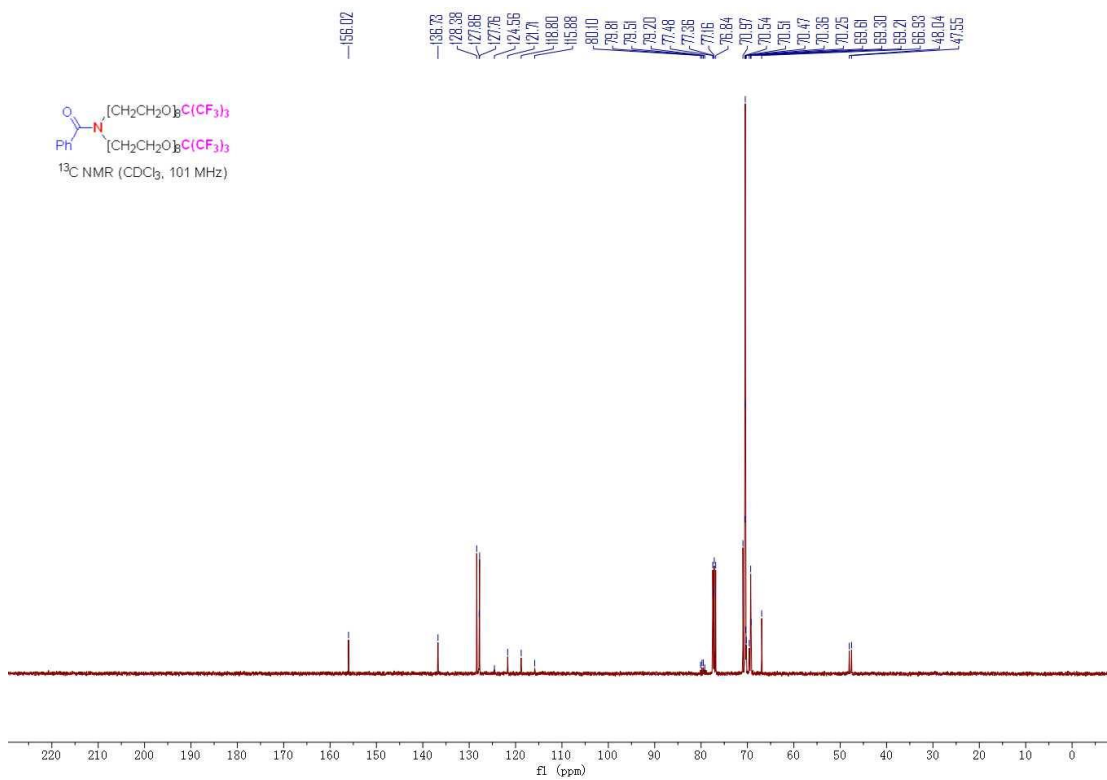
24#1099 RT: 11.00 AV: 1 NL: 1.45E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



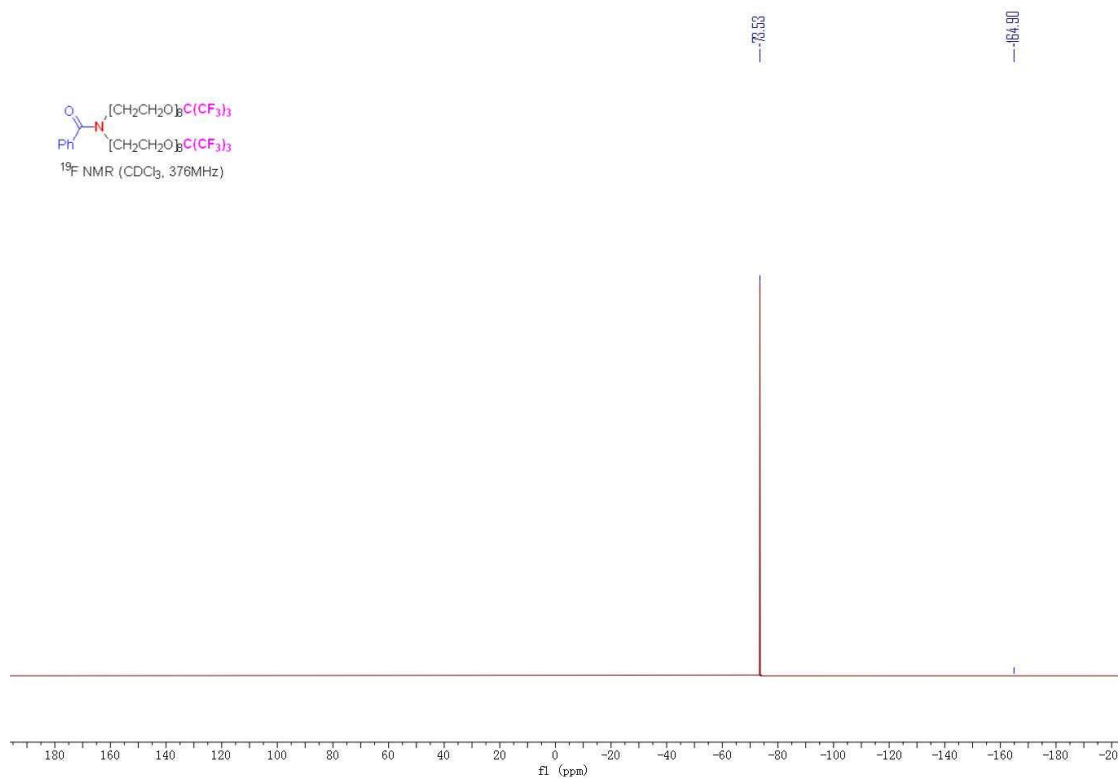
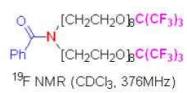
¹H NMR of compound **37**



¹³C NMR of compound **37**

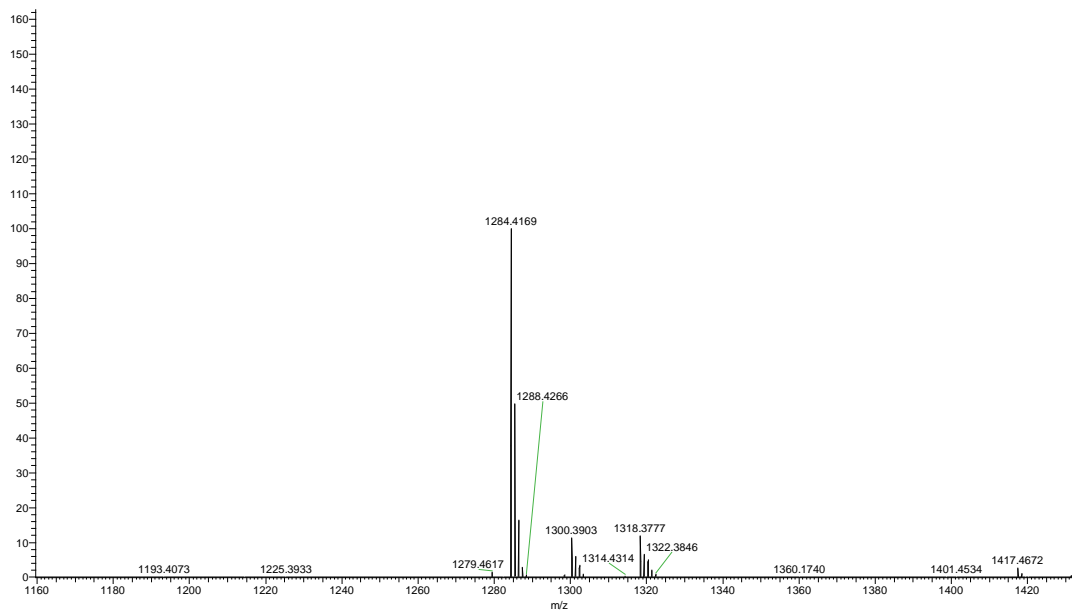


¹⁹F NMR of compound 37

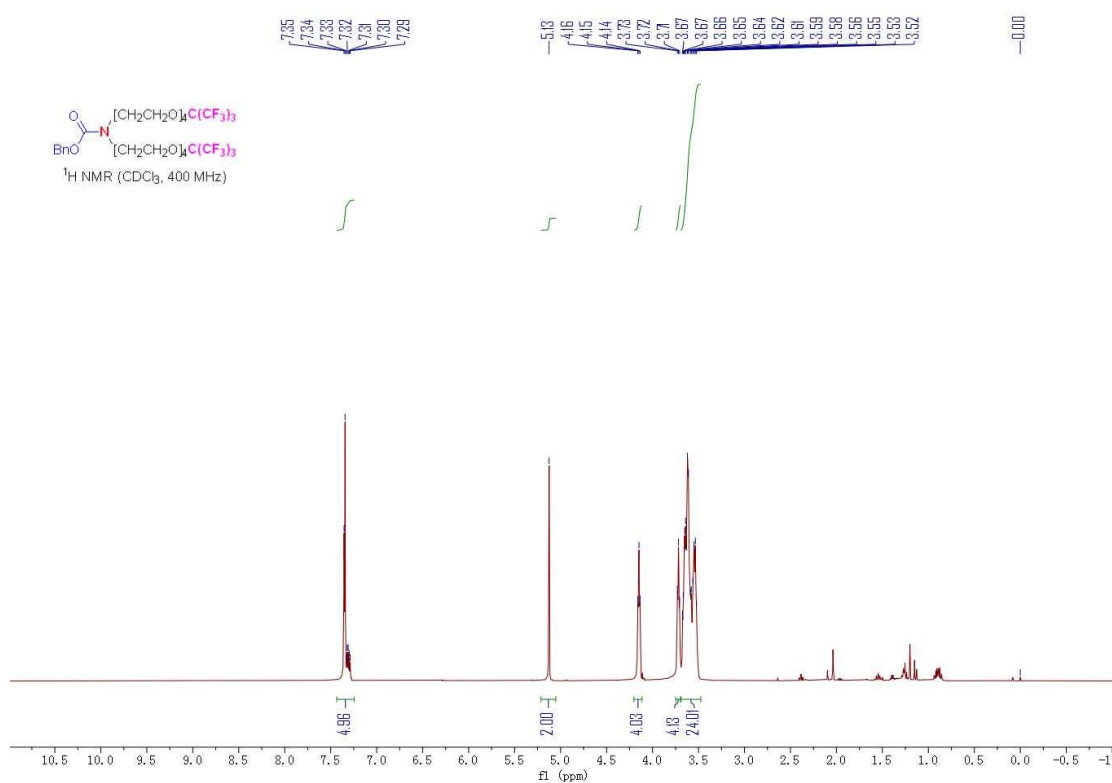
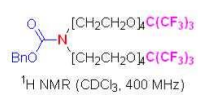


HRMS of compound 37

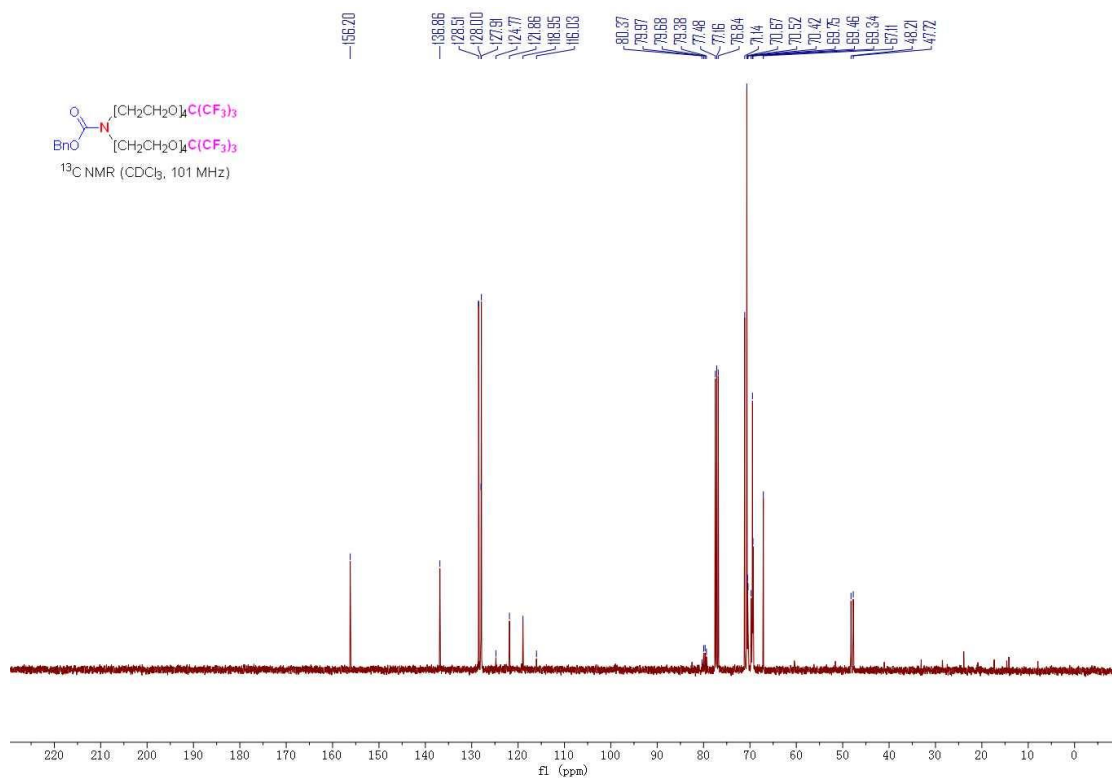
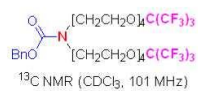
19#1059 RT: 10.62 AV: 1 NL: 2.73E7
T: FTMS + p ESI Full ms [150.0000-2000.0000]



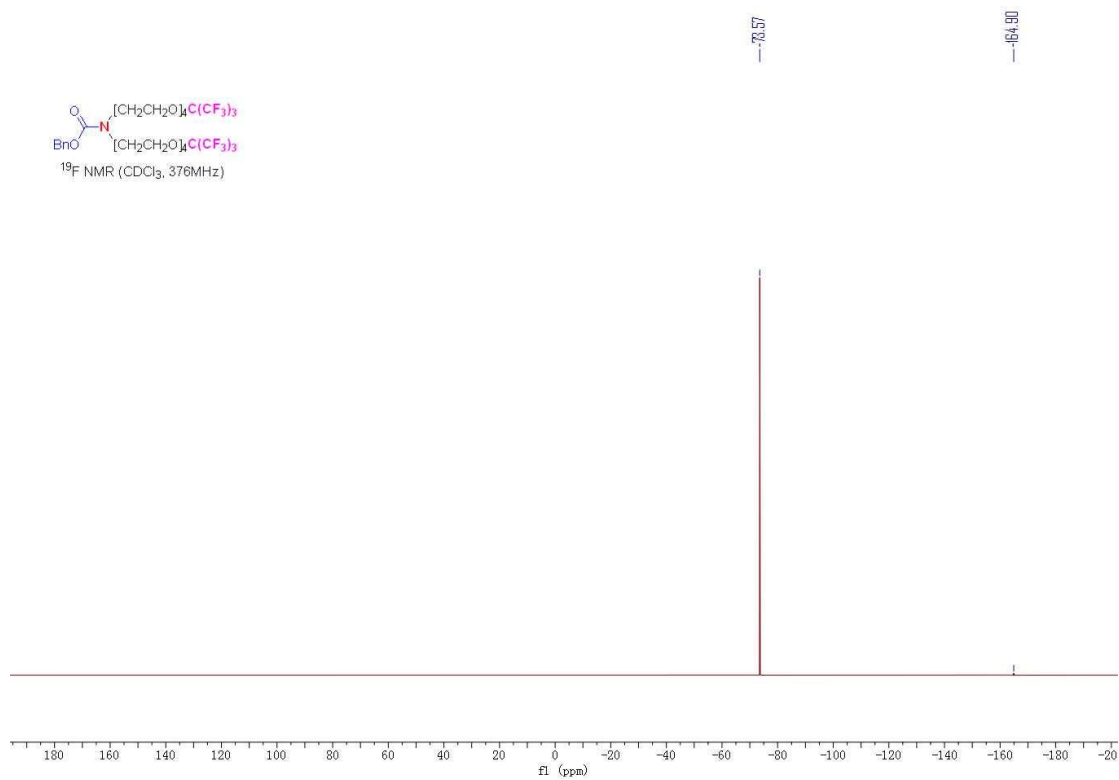
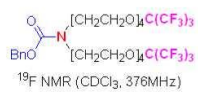
¹H NMR of compound 38



¹³C NMR of compound 38

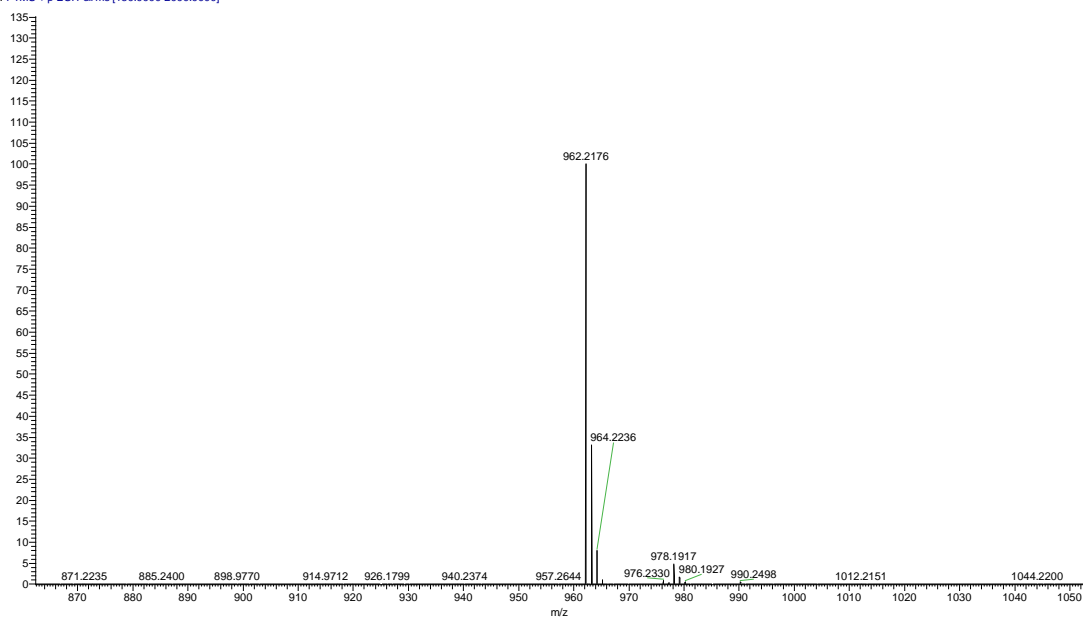


¹⁹F NMR of compound 38

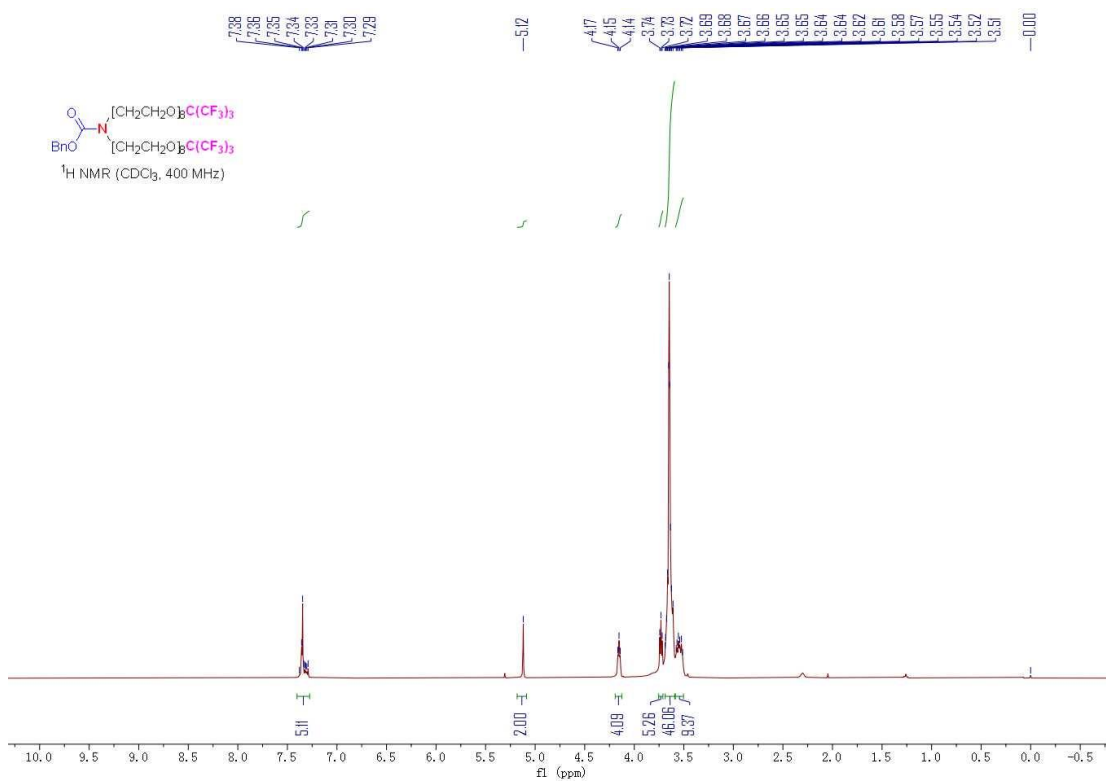


HRMS of compound 38

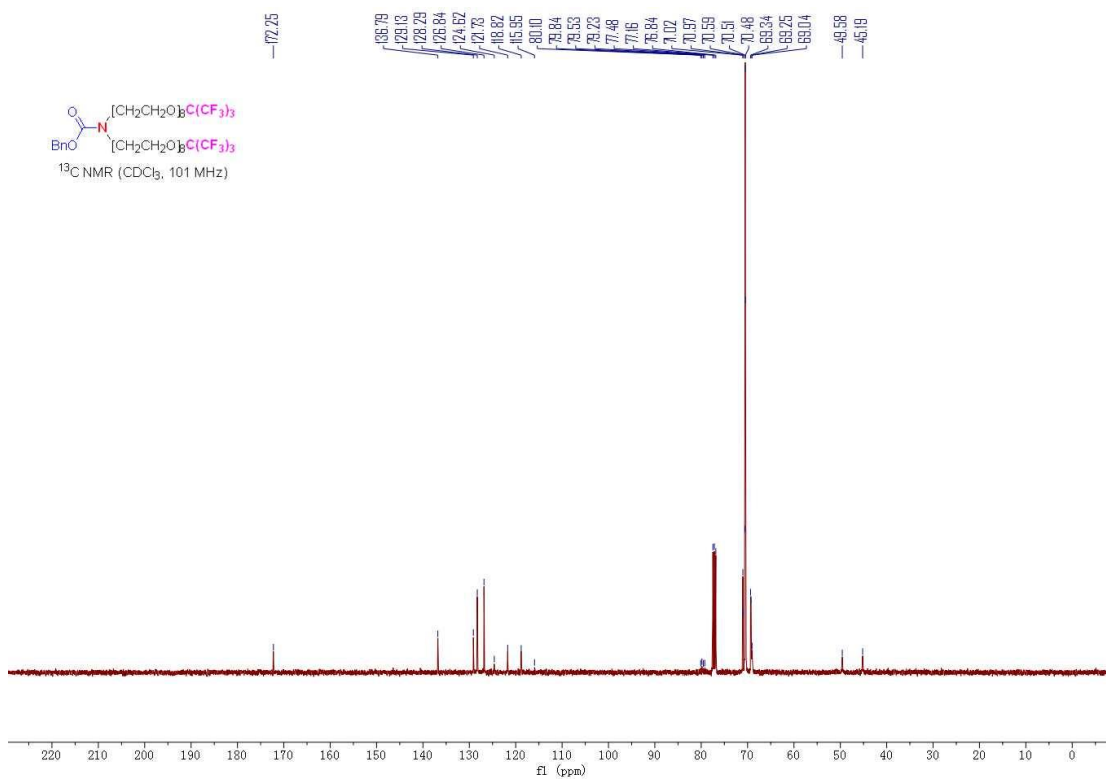
23 #1099 RT: 10.99 AV: 1 NL: 1.06E8
T: FTMS + p ESI Full ms [150.0000-2000.0000]



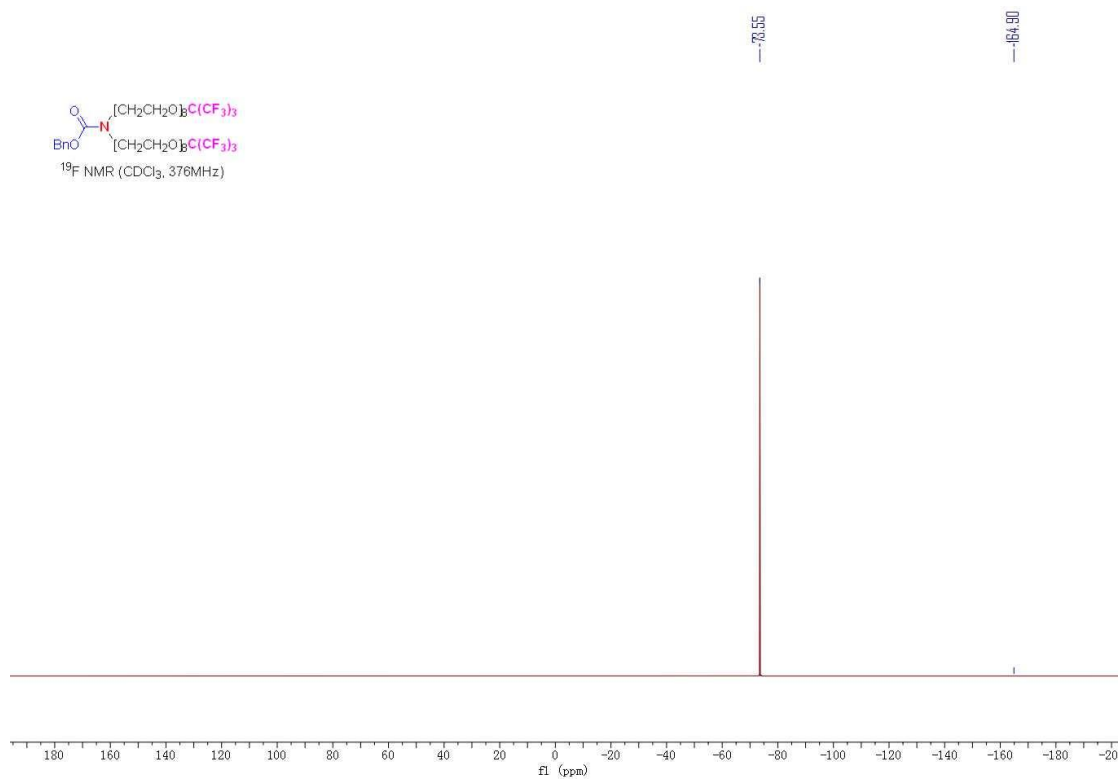
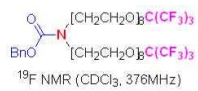
¹H NMR of compound **39**



¹³C NMR of compound **39**

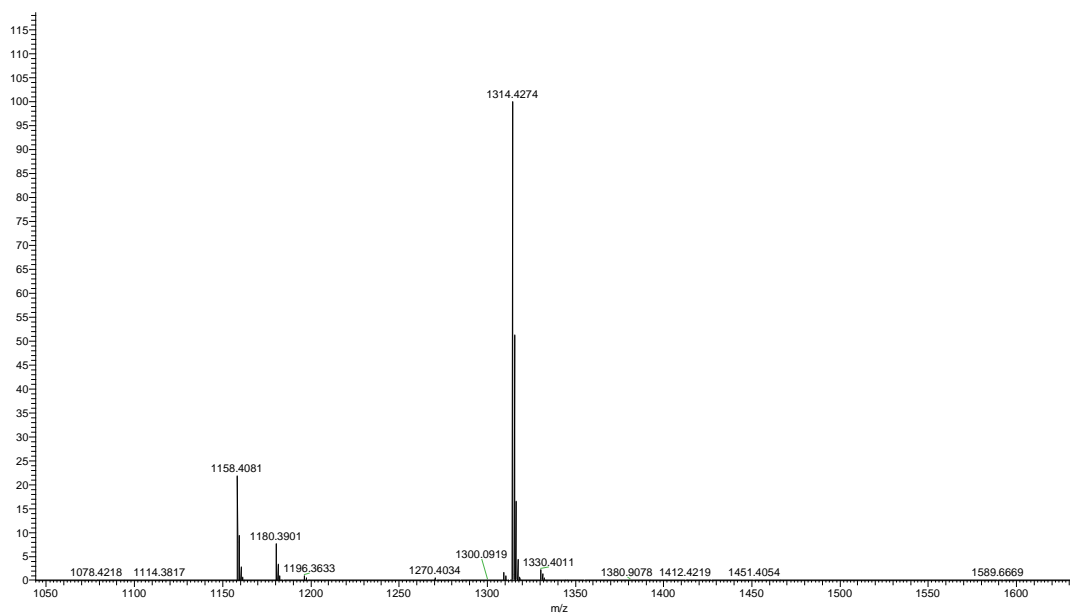


^{19}F NMR of compound **39**

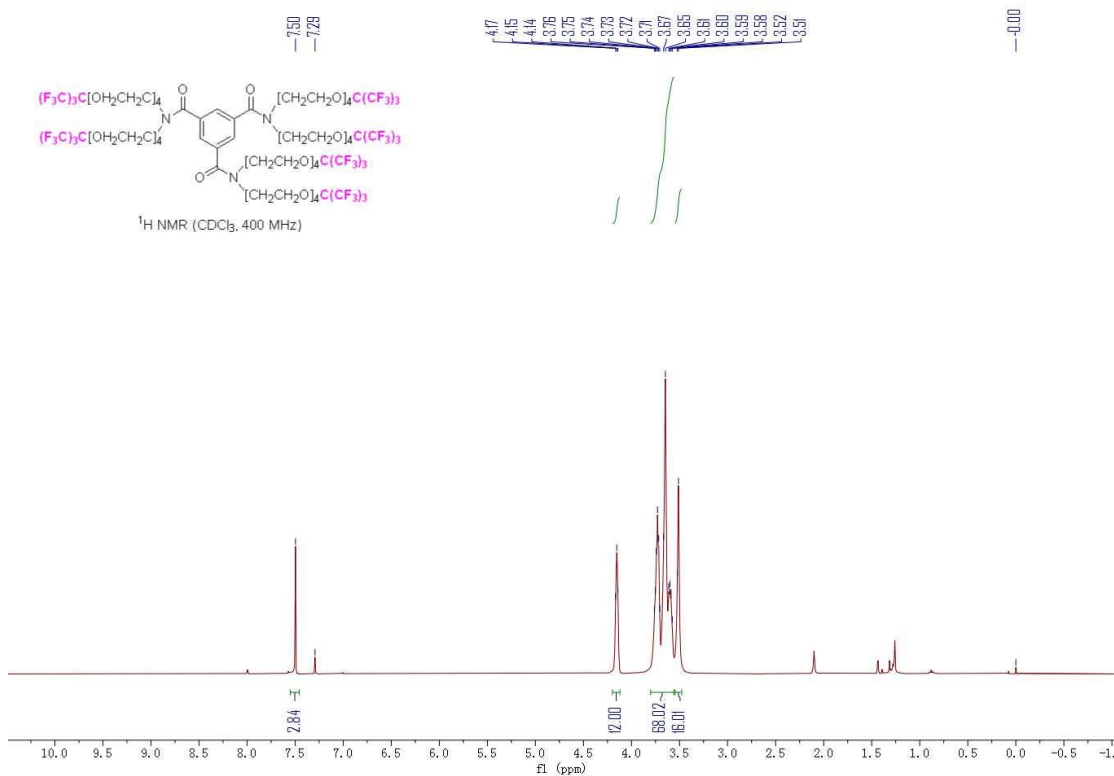


HRMS of compound **39**

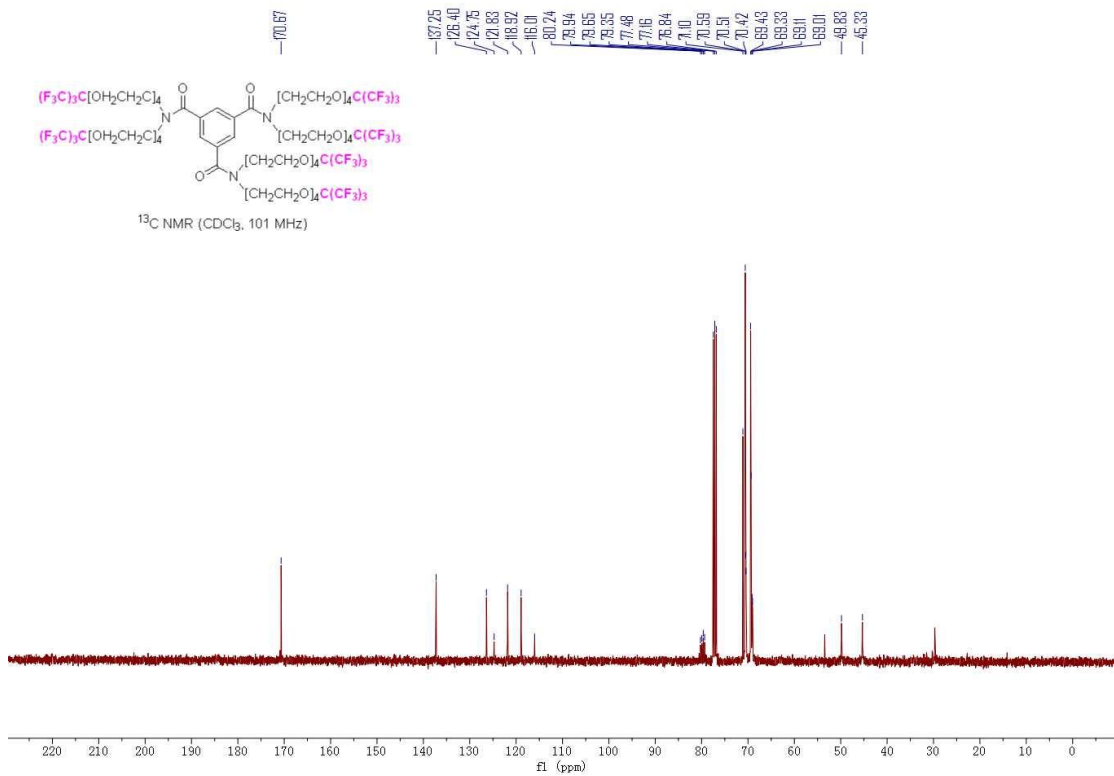
18 #1073 RT: 10.72 AV: 1 NL: 1.28E9
T: FTMS + p ESI Full ms [150.0000-2000.0000]



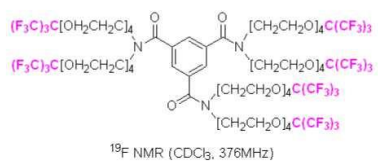
¹H NMR of compound 40



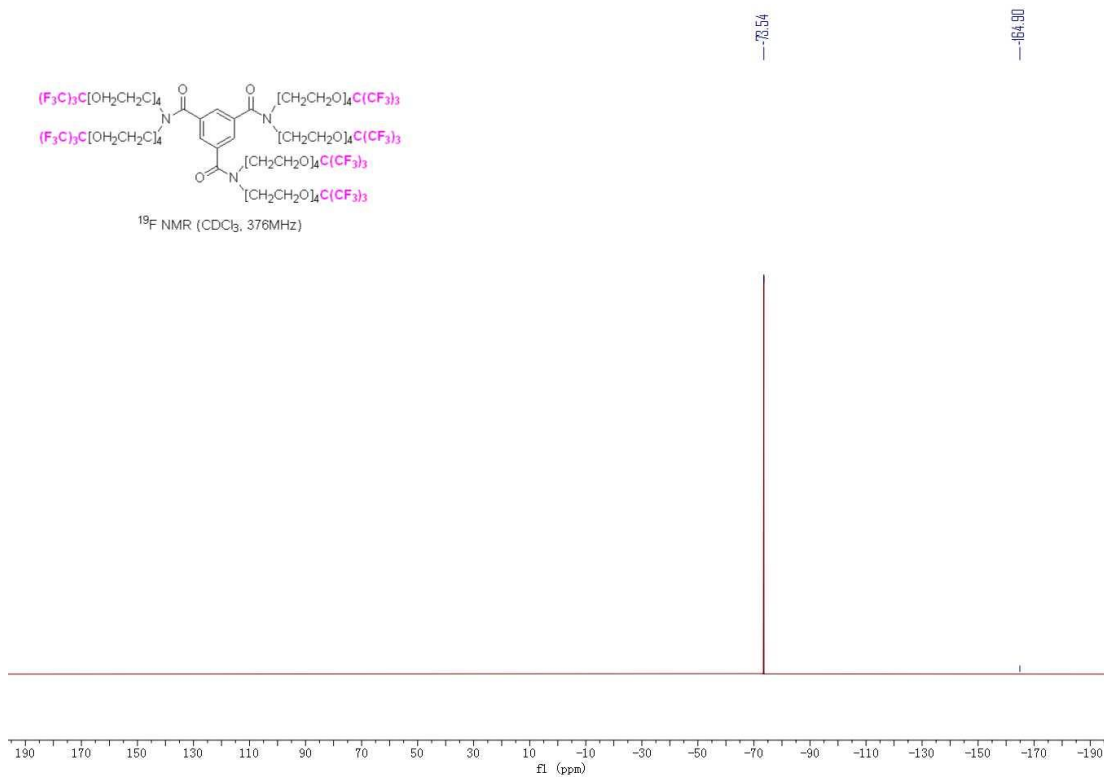
¹³C NMR of compound 40



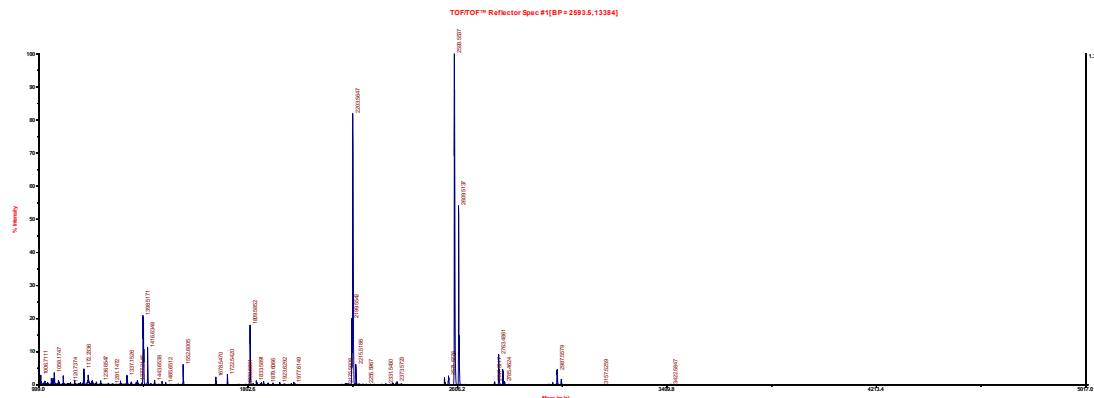
¹⁹F NMR of compound 40



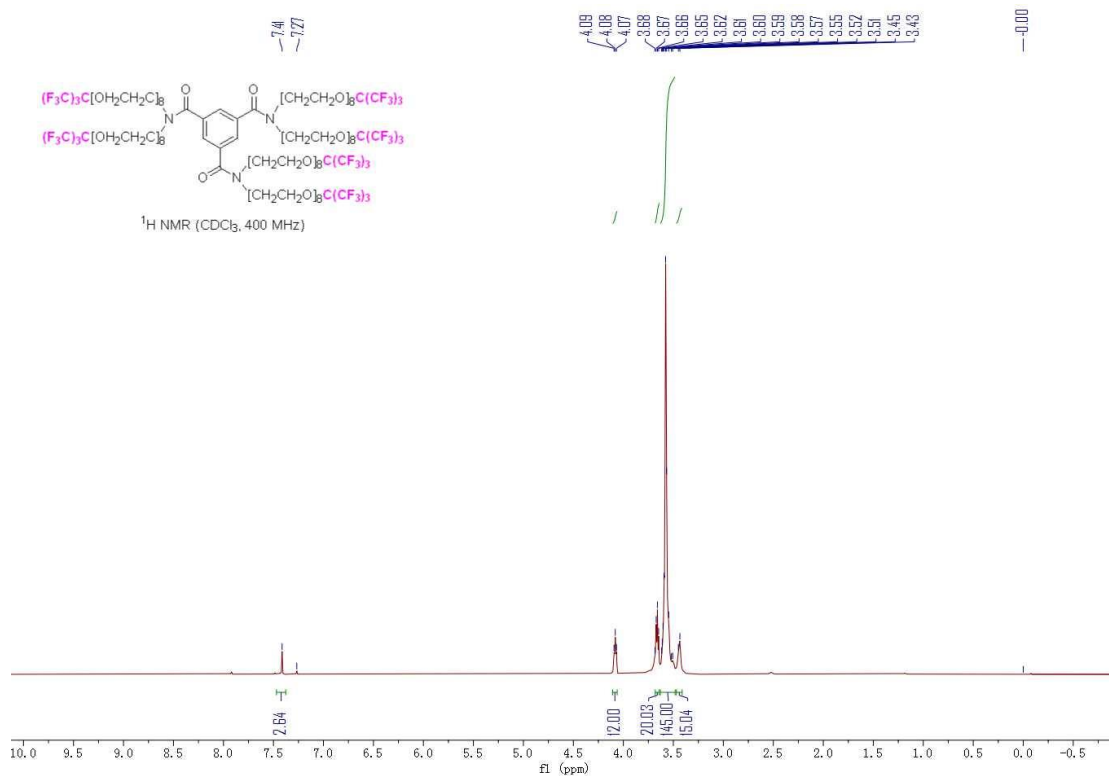
¹⁹F NMR (CDCl₃, 376MHz)



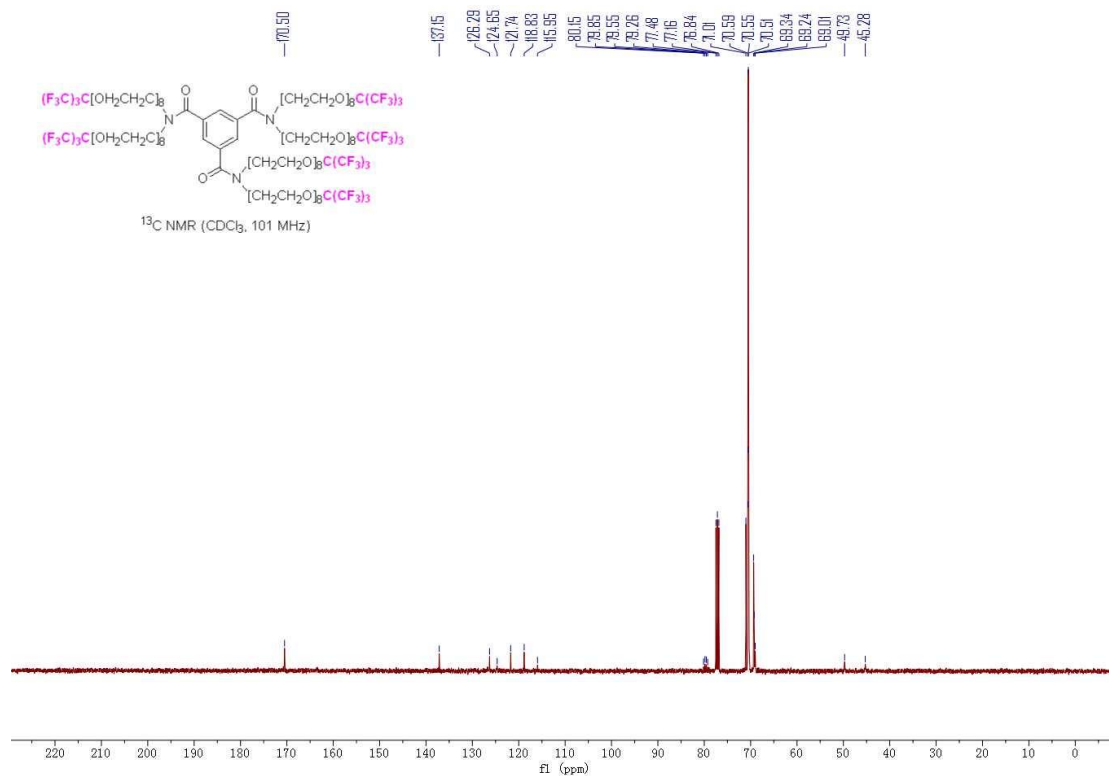
HRMS of compound 40



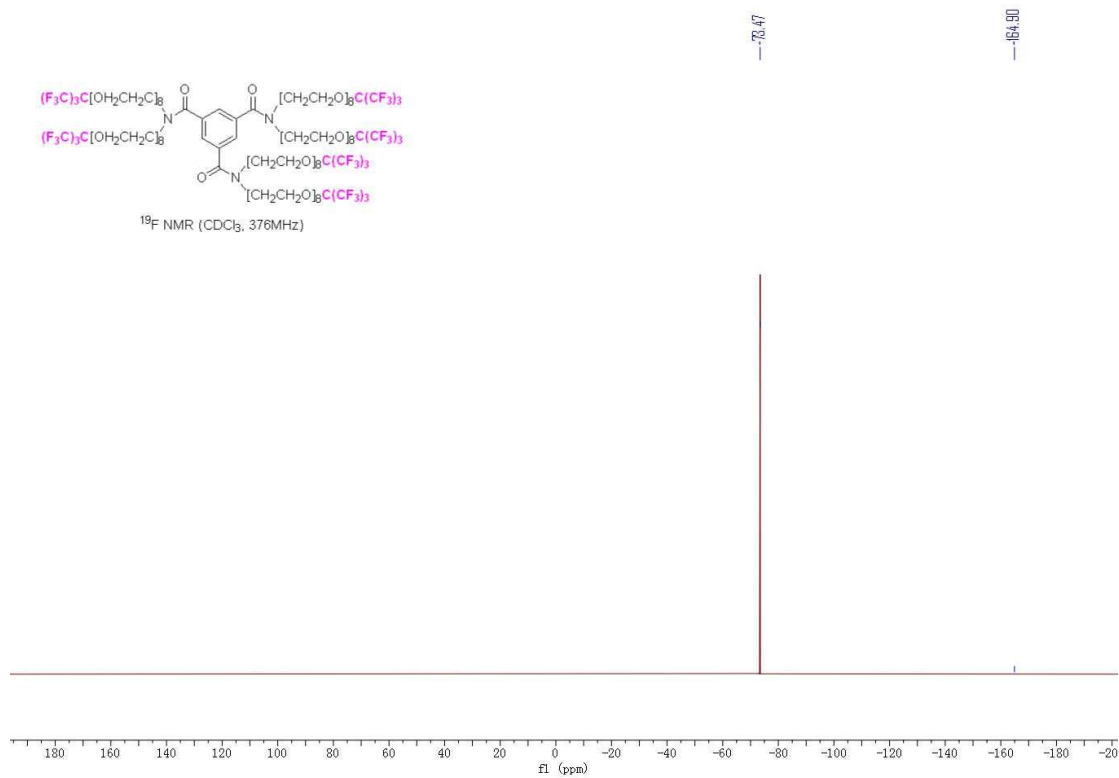
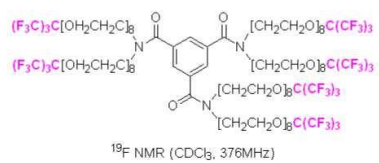
¹H NMR of compound 41



¹³C NMR of compound 41



¹⁹F NMR of compound 41



HRMS of compound 41

