Biothiol Xenon MRI Sensor Based on Thiol-Addition Reaction

Shengjun Yang,[†] Weiping Jiang,[†] Lili Ren,[†] Yaping Yuan,[†] Bin Zhang,[†] Qing Luo,[†] Qianni Guo,[†] Louis-S. Bouchard,[‡] Maili Liu,[†] and Xin Zhou^{*,†}

[†]Key Laboratory of Magnetic Resonance in Biological Systems, State Key Laboratory of Magnetic Resonance and Atomic and Molecular Physics, National Center for Magnetic Resonance in Wuhan, Wuhan Institute of Physics and Mathematics, Chinese Academy of Sciences, Wuhan, 430071, China

[‡]Department of Chemistry and Biochemistry, California NanoSystems Institute, The Molecular Biology Institute, University of California, Los Angeles, California 90095, United States

*Email: xinzhou@wipm.ac.cn

Supporting Information

Contents:

- 1. Characterization of cryptophane 1
- 2. Time dependent ¹²⁹Xe NMR signal of cryptophane 1 interacting with biothiols
- 3. Biosensor 1 in response to the low Cys concentration
- 4. Biosensor 1 in response to Cys in HEPES buffer with 10% DMSO
- 5. Absorption spectra of cryptophane 1 treated with 10 equiv of Cys
- 6. Biosensor 1 in response to Hcy
- 7. Biosensor 1 in response to GSH
- 8. TOF-MS spectrum of reaction product between cryptophane 1 and Cys
- 9. ¹H NMR spectral comparison between cryptophane 1 and [1-Cys] adduct
- 10. ¹²⁹Xe NMR chemical shift comparison of cryptophane 1, cryptophane 2 and [1-Cys] adduct
- 11. ¹²⁹Xe spectra of cryptophane 1 in the presence of various analytes
- 12. Sensitivity of biosensor 1
- 13. Biosensor 1 in response to GSH and Hcy in bovine serum solution

1. Characterization of cryptophane 1:



CDCl₃.



Figure S2. HR MS-ESI of cryptophane 1.

2. Time dependent ¹²⁹Xe NMR signal of cryptophane 1 interacting with biothiols:



Figure S3. Time-dependent ¹²⁹Xe NMR signal intensity at δ = 76.9 ppm (left) and the corresponding concentration change (right) of cryptophane 1 (200 μ M) after treated with 3 equiv Cys (\blacksquare), Hcy (\bullet) and GSH (\blacktriangle), respectively.

3. Biosensor 1 in response to the low Cys concentration:



Figure S4. ¹²⁹Xe chemical shift change of cryptophane 1 (200 μ M) upon addition of 1 equiv Cys.

4. Biosensor 1 in response to Cys in HEPES buffer with 10% DMSO:



Figure S5. Time-dependent ¹²⁹Xe NMR spectra change of cryptophane 1 (10 μ M) upon addition of 10 equiv Cys in 20 mM HEPES buffer (pH 7.4) with 10% DMSO at

25 °C. Under this conditions, the chemical shift of dissolved free Xe was at δ = 201.4 ppm and chemical shift of caged Xe in cryptophane 1 was at δ = 72.8 ppm, respectively. After addition of 10 equiv Cys, a new signal at 71.2 ppm appeared. The result means this reaction can proceed in low cryptophane concentration and DMSO content. All spectra were obtained with 32 scans.



5. Absorption spectra of cryptophane 1 treated with 10 equiv of Cys:

Figure S6. Absorption spectra of cryptophane 1 (10 μ M) after treated with 10 equiv Cys. The spectra were recorded at 0, 10 and 50 min.

6. Biosensor 1 in response to Hcy:

						76.	9 ppm								
		~~~~~	0 n	nin			$\bigwedge$								
			<u>8 n</u>	nin		75.7 ppm									
	12 min														
	16 min														
19 min															
	28 min														
	40 min														
	55 min						$\sim$		~~~~~			~~~~~			
	70 min						$\sim$								
	90 min														
	120 min 180 min														
90		86		82	80	78	76 f1 (pr	74 om)	72	70	68	66	64	62	60

Figure S7. Time-dependent ¹²⁹Xe NMR spectra change of cryptophane 1 (200  $\mu$ M) upon addition of 3 equiv Hcy. All spectra were obtained with a single scan.

#### 76.9 ppm 0 min 75.7 ppm 7 min 10 min 16 min 26 min 36 min 51 min 66 min 81 min 96 min 136 min 236 min 76 74 f1 (ppm) 86 84 82 78 72 70 90 88 80 68 66 64 62 Figure S8. Time-dependent ¹²⁹Xe NMR spectra change of cryptophane 1 (200 $\mu$ M)

60

S-7

#### 7. Biosensor 1 in response to GSH:

upon addition of 3 equiv GSH. All spectra were obtained with a single scan.



8. TOF-MS spectrum of reaction product between cryptophane 1 and Cys:

Figure S9. TOF-MS spectrum of reaction product between cryptophane 1 and Cys.





Figure S10. ¹H NMR spectral comparison between cryptophane 1 and its Cys adduct. (a) cryptophane 1 only; (b) [1-Cys] adduct.





Figure S11. (a) The reaction of cryptophane 1 with Cys and (b) ¹²⁹Xe NMR chemical shift of cryptophane 1, cryptophane 2 and [1-Cys] adduct. All spectra were obtained under the same test conditions. The result indicates that the reaction of cryptophane 1 with Cys do not undergo the intramolecular cyclization.

11. ¹²⁹Xe spectra of cryptophane 1 in the presence of various analytes:



Figure S12. Single-scan ¹²⁹Xe NMR spectra of caged xenon of cryptophane 1 (200  $\mu$ M) in the presence of various analytes (the concentration of Cys, Hcy and GSH were 600  $\mu$ M, the rest analytes were 2000  $\mu$ M). The result indicates that other analytes including amino acids and sulfur-containing molecular: 4-methoxy thiophenol, HSCH₂CH₂OH, hydrogen sulfide, sulfite, sulfate, thiosulfate, and hydrogen sulfite caused no obvious ¹²⁹Xe NMR chemical shift change of cryptophane 1.

#### 12. Sensitivity of biosensor 1:

(a)



Figure S13. (a) Single scan ¹²⁹Xe NMR spectrum of caged xenon of cryptophane 1 at a concentration of 10  $\mu$ M. (b) 4096 signal averages enable the detection of ¹²⁹Xe NMR of caged xenon of cryptophane 1 at a concentration of 80 nM after treatment with Cys. In this experiment, Xe gas was introduced by continuous-flow at the rate of 0.08 standard liters per minute.



13. Biosensor 1 in response to GSH and Hcy in bovine serum solution:

Figure S14. ¹²⁹Xe NMR spectra of cryptophane 1 (40  $\mu$ M) in 20 mM HEPES buffer (pH 7.4) solution (containing 10% bovine serum and 20% DMSO, v/v) at (a) 10 min, (b) 180 min, (c) 400 min and in the presence of (d) GSH, (e) Hcy. The ¹²⁹Xe NMR chemical shift of caged xenon is (a) 74.1 ppm, (b) 74.1 ppm, (c) 74.1 ppm, (d) 73.0 ppm and (e) 72.7 ppm, respectively. All spectra were obtained with 16 scans and the line width is 20 Hz.